Illustrated Key to the Longhorned Woodboring Beetles of the Eastern United States

Lingafelter







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Front Cover Photos—top, *Strangalia strigosa* Newman [by Eugenio Nearns]; bottom left, *Romulus globosus* Knull [by Roy Morris]; bottom right, *Aegomorphus morrisii* (Uhler) [by Eugenio Nearns].

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Abstract: A fully illustrated key with over 800 habitus and character photographs is presented to allow the easy identification of eastern U. S. Cerambycidae. Of the 400 species of Cerambycidae that occur east of the Rocky Mountains (but excluding southern and western Texas), 377 species are treated in the key. Only uncommonly collected or isolated taxa from the Great Plains, Great Lakes Region, or extreme upper New England are excluded. Nine invasive Cerambycidae known or suspected to be established in the eastern U. S. are also included in the key. The key includes 417 couplets that are arranged such that most taxa will key out in less than 20 couplets and 10 minutes. The key uses only easily seen external characters, never requires dissection, and never requires both sexes of a species to be available. It emphasizes ease of identification over constraining genera, tribes, or subfamilies to remain together. Unless otherwise specified, all nomenclature follows the latest checklist of Cerambycidae of the Western Hemisphere by Monné & Hovore (2006).

This illustrated key includes most of the Cerambycidae of the eastern United States (as defined in Yanega, 1996:11) with the exception of those species occurring only in the boreal forest (taiga) around the Great Lakes and extreme upper New England (Maine and northern portions of New Hampshire, Vermont, and New York) and a few other rarely collected species. This represents 377 of the 400 known species east of the Rocky Mountains but excluding southern and western Texas (about one-third of the United States fauna). Every currently known species in the Southeastern and mid-Atlantic United States (Tennessee, Kentucky, North Carolina, South Carolina, Virginia, West Virginia, Maryland, Delaware, Louisiana, Alabama, Mississippi, Georgia, and Florida) is included, and this key can be used with full confidence for those states. Given that many of the included taxa also occur in central USA and Canada, it will be useful for identifying 90% of those species that occur east of the Rocky Mountains, with the exception of southern and western Texas (see Fig. 1 for confidence intervals). The map indicates confidence intervals based on percentages of species

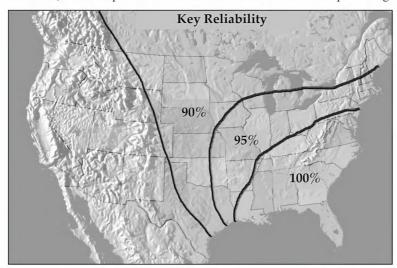


Figure 1. Confidence intervals reflecting percentage of species from given regions that are included in the key.

included and able to be identified for the given region, but the confidence intervals will actually be much higher if considering success based on specimens encountered for the given regions, since those species excluded are rare, isolated, and uncommonly encountered. The nine invasive species confirmed for the eastern United States are also included in the key. The key should not be used for identifying species from southern or western Texas, or the Rocky Mountains and localities west, as it will have a reliability of less than 50% for those regions.

The key was constructed in a practical way to facilitate identification without the need for dissection, without the need for having both sexes of a species available, and without the need for expert knowledge of morphology. It was produced by first generating a 10 drawer synoptic collection of about 1,500 specimens representing the 377 species covered. These species were sorted and subsorted based on easy to see characters wherever possible, and divided into equally sized subsets wherever possible. Using a digital automontage camera system, over 800 images were taken and enhanced in Photoshop® and these images of diagnostic characters accompany most couplets. After the key are short synopses and dorsal habitus photos for nearly every species. If I could maintain a natural grouping in the key, I did; however, subfamilies and tribes are not necessarily treated together as they are in the Cerambycidae of North America monograph series (Linsley and Chemsak, 1961-1995). That was a monumental work, but since taxa were constrained to key together even if some characters did not support that, users of the keys were often frustrated by wrong turns. There are many instances in that work where a tribal or genus character is unreliable later when keying one of its included species. As examples of how I have avoided that situation: the aberrant asemine, Atimia, fits with the Cerambycinae based on external features that do not require dissection (the shape of the palpi); the unusual Necydalis, likewise, is removed from the other lepturine genera because of its unusual morphology (short from and gena). In many cases, characters can reasonably be interpreted in different ways (or may be expressed differently among several specimens). Where I determined this was the case, I have the species occurring in two (or more) places in the key. For example, Romulus globosus Knull may or may not have spines on the antennae. Therefore, it keys with its elaphidiine congeners and elsewhere in the key. Several acanthocinine genera have pronotal tubercles that could reasonably be interpreted as acute or rounded, so they are treated in both sections of the key beyond that couplet. About 50 species occur at least twice in the key to account for the occasional character that could be interpreted in two ways, or is simply variable within species, so the user is not penalized for making a "wrong" turn.

For identifying eastern United States Cerambycidae, Yanega's 1996 field guide to Northeastern Longhorned Beetles is an excellent resource. Users of this key may wonder why they should bother keying anything out since Doug Yanega prepared the field guide with illustrations of all northeastern species. First, this key covers all the species in the southeast U. S. which were not covered in Yanega (1996). Second, when running through a key that has considered all species in a given region and accounted for morphological variation within species, the user gains a certainty after the identification process that is absent from comparing to pictures and perhaps glossing over some important character details. Yanega (1996) does include a few of the rare or isolated taxa that occur in extreme upper New England or the Great Lakes region that are not included in this key, so it remains an indispensible resource. Third, this is the only key that includes known invasive North American Cerambycidae as well as native species. Given the onslaught of invasive organisms, their economic cost, having a resource that facilitates their identification is highly beneficial to all of us. As a final justification for this key: You will get a definitive name on each specimen from the Midatlantic or southeast United States in 10 minutes or less, nearly every time. No other resource will enable this speed and certainty with identifications.

I have included a general morphological atlas (Figure 2) that defines the structures used in the key, along with some other general anatomical features of Cerambycidae. By no means should you memorize these structures, as many of them are clearly shown in the hundreds of automontage photographs that follow throughout the key. It merely serves as a reference figure. Note that the figures in the key are not numbered, but their number is inferred and referred to as the couplet number immediately above the figure. To avoid unnecessary duplication, in some cases a figure is needed in several couplets and the user may be instructed to refer to an earlier appearance of that figure.

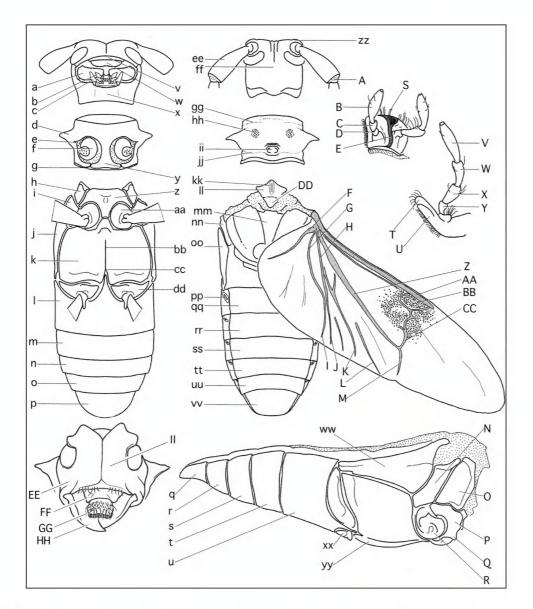


Figure 2. Morphological atlas of a generalized cerambycid. a, mandible; b, labium; c, maxilla; d, lateral pronotal tubercle; e, lateral (external) procoxal cavity; f, procoxa; g, posterior procoxal margin (closed); h, mesepisternum; i, mesepimeron; j, metepisternum; k, metasternum; l, ventrite 1; m, ventrite 2; n, ventrite 3; o, ventrite 4; p-q, ventrite 5; r, ventrite 4; s, ventrite 3; t, ventrite 2; u, ventrite 1; v, clypeus; w, labrum; x, gular region; y, prosternal intercoxal process; z, mesosternum; aa, mesocoxa; bb, metasternal sulcus; cc, metasternal intercoxal apex; dd, metacoxa; ee, scape (first antennomere); ff, vertex of head; gg, anterior constriction of pronotum; hh, anterolateral patch (sometimes patch of hairs, punctures, or raised callus at this position); ii, posteromedial pronotal callus; jj, posterior pronotal constriction; kk, stridulatory region of mesonotum; II, mesonotum (attached to mesoprescutum or scutellum); mm, metanotum; nn, mesepimeron; oo, metepisternum; pp, first abdominal spiracle; qq, tergite 2; rr, tergite 3; ss, tergite 4; tt, tergite 5; uu, tergite 6; vv, tergite 7; ww, metepisternum; xx, metacoxa; yy, metasternum; zz, antennal tubercle; A, scape cicatrix; B, apical labial palpomere (palpomere 4); C, penultimate labial palpomere (palpomere 3); D, palpomere 2; E, labial ridge or crest; F, AP vein; G, AA vein; H, CuA vein; I, AA3+4 vein; J, CuA3+4 vein; K, MP4 vein; L, MP3 vein; M, Medial Spur vein; N, mesepimeron; O, mesepisternum; P, mesosternum; Q, mesosternal process or tubercle; R, mesocoxa; S, marginal (apical) setae of labium; T, galea; U, lacinia; V, apical maxillary palpomere (palpomere 5); W, penultimate maxillary palpomere (palpomere 4); X, maxillary palpomere 3; Y, maxillary palpomere 2; Z, MP vein; AA, RA vein; BB, Radial Cell; CC, RP-MP vein; DD, scutellum; EE, gena; FF, clypeus; GG, labrum; HH, mandible; II, frons.

Acknowledgments

Special thanks to Elisabeth Roberts who assembled a 10-drawer synoptic collection of nearly 400 species (with up to 20 specimens of each to reflect morphological variation) from the Smithsonian Institution collection. This collection served as the core resource for developing the key and providing the subjects for the auto-montage images. Lisa also took many of the full dorsal habitus photos, mostly using the Microptics system. Special thanks to Gino Nearns, Charyn Micheli, Norm Woodley, Serge Laplante, Deblyn Mead, and Annie Ray for helping to test the key and for their moral support and encouragement. Michael Gates, Mike Thomas, and Allen Norrbom also reviewed this key. I thank Amanda Hodges, Eric Day, and all the students of the 2006 Southern Plant Pest Diagnostic Network Workshop in Blacksburg, Virginia.

The students in this workshop thoroughly tested an earlier version of the key and recommended many areas for refinement. I appreciate the help Gino Nearns provided during that course. For sending some images and specimens of Florida specialties, mostly from the Florida State Collection of Arthropods (FSCA), I thank Mike Thomas, Paul Skelley, and Gino Nearns. Mike Thomas, Roy Morris, Shane Hill, and Gino Nearns developed the very useful Cerambycidae of Florida website that I found quite helpful while working on this key. Thanks to Piotr Naskrecki, Brian Farrell, and Phil Perkins for their joint effort in developing the important Harvard Holotype Image Database that helped resolve some species distinctions. Natalie Allen assisted me by entering data on the seasonality and hosts for many species, and taking some dorsal habitus photos.

Special thanks also to David Smith for providing a huge amount of eastern U. S. longhorned beetles from his West Virginia malaise trap samples. These were invaluable specimens for teaching the identification workshop in Blacksburg, and also for enhancing our knowledge of distribution and seasonality of eastern U. S. longhorned woodboring beetles.

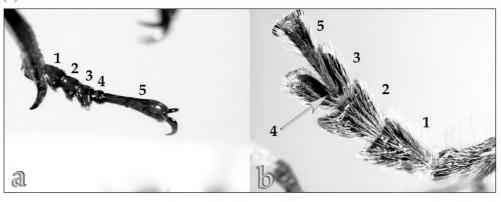
Lastly, I am very appreciative to Terry Seeno for performing the tedious page layout, formatting, and final editing and to Michael Schauff for locating funds within USDA to pay for most of the printing cost.

Illustrated Key to the Longhorned Woodboring Beetles of the Eastern United States

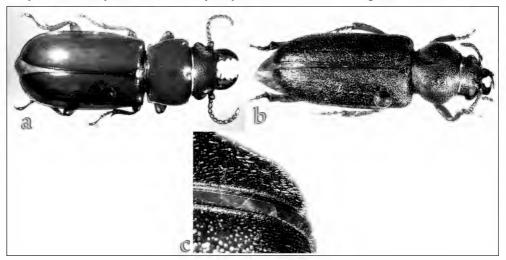
1. Antennal insertions at fronto-clypeal margin, near mandibular insertion (a, b). Acute lateral pronotal tubercles present (b). Maxillary palpi long, about as long as first three protarsomeres combined (b). Antennal scape curved at base and longer than head (b). Prosternal process narrow, not expanded at apex (c). Elytral apices bispinose and patterned as in (d) (Disteniinae)

Distenia undata (Fabricius)



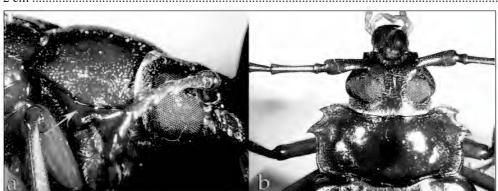


- **3(2).** Body glabrous, smooth, shiny, red; carabid-like appearance (a) (Parandrinae)......4

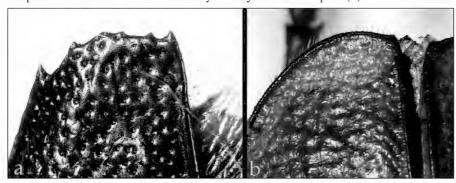


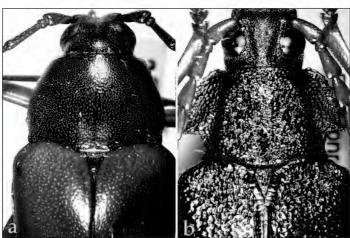


5'. Pronotum without defined, complete lateral margin. Mandibles not very prominent in most species. Elytra usually with pubescence. Species of variable size, although most are less than 2 cm _______21

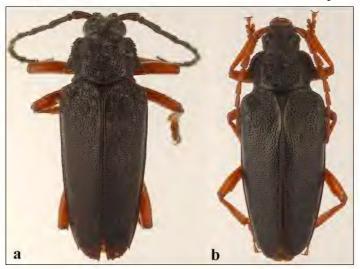


- 6'. Elytral apices rounded to suture which may or may not have a spine (b)......9



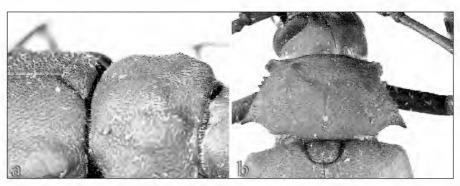


- 8(7'). Antennae black (a) Elateropsis rugosus Gahan



- **9(6).** Femora densely pubescent with long hairs. Thorax densely pubescent ventrally with long hairs (and dorsally in males) that are not appressed (a)............ *Tragosoma depsarium* (Linnaeus)
- 9'. Femora mostly glabrous; if venter of thorax is pubescent, then hairs are appressed and short (b).....10

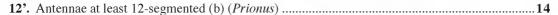




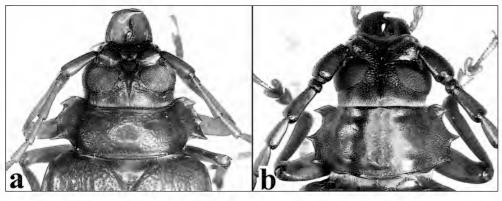
11(10'). Lateral margin of pronotum produced into 2 or 3 main spines or angulate projections (a)...12





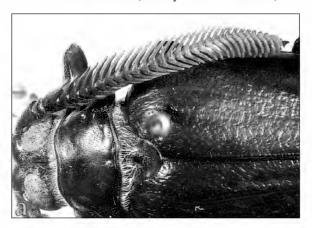






14(12'). Antennae v	with at least 25 anteni	nomeres (often over	er 30) (a) <i>Prioni</i>	s fissicornis Haldeman
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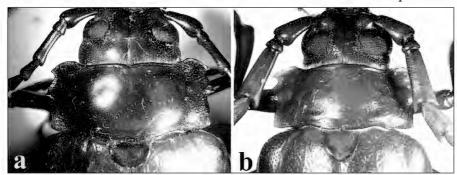




17(16').	Antennae	with mo	re than 1	12 segments	(usually	more than	15).	Antennomeres	beyond
eight	t strongly a	ppendici	ılate (a).				Prior	ius imbricornis	(Linnaeus

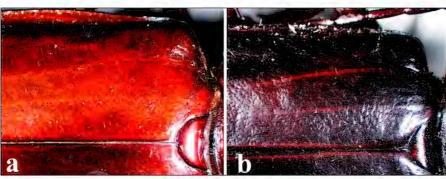




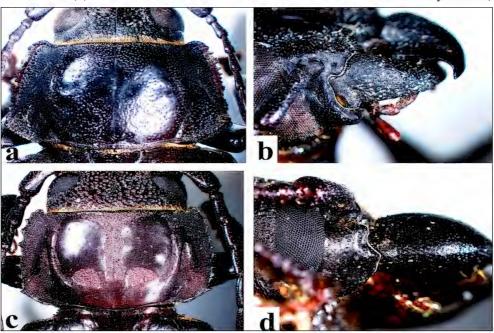


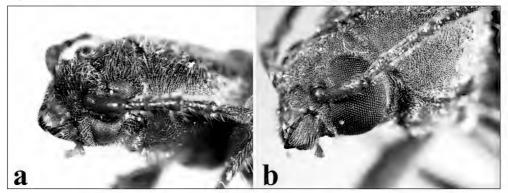
19(11'). Elytral surface smooth, shiny, without punctures or wrinkles (a). Color reddish-brown

Stenodontes chevrolati Gahan



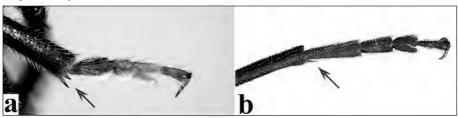
20(19').	Pronotum	distinctly	narrowed	anteriorly	at margin	(a).	Genal	margin	near	mandible	
inser	tion with o	ne or no tu	ibercles (b))		Ar	chodon	ites mela	пори	ıs (Linnae	eus)



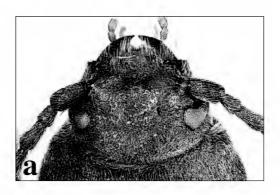


- **23(22).** Pronotum with dense, coarse punctures and microsculpture giving a matte, non-reflective.finish (a). Elytral and venter coloration very similar, brown-gray *Tetropium schwarzianum* Casey





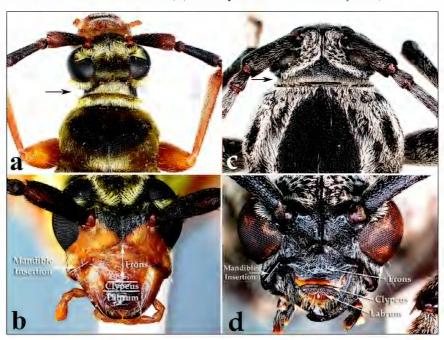
- **25(24').** Antennae thickened at base (especially scape). Interantennal region strongly impressed (no specimens available for imaging)......
 - Asemum australe LeConte



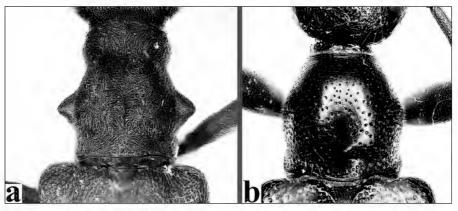


- **26°.** Third tarsomere of posterior legs cleft nearly to base (a) (note: I have elected not to distinguish among the various subspecies of *Arhopalus rusticus* (Linnaeus)).....

27(21'). Pronotum of most species tapering or constricted anteriorly, combined with a narrowing
	of head behind eyes giving a neck-like appearance (a). Frons, gena, and sometimes clypeus
	elongate, giving appearance of mouthparts more elongated or protuberant than in other groups
	(b). Body of most species distinctly tapering posteriorly. Stridulatory plate of mesonotum
	divided by a median line (best seen with dissection) (Lepturinae, except Necydalis)28

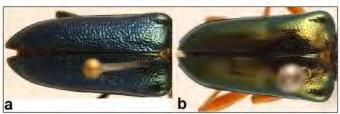


- **28(27).** Pronotum with lateral tubercles (either acute or rounded), defined by constriction immediately anterior and posterior of tubercle (a) (see also 39a-b)......**29**



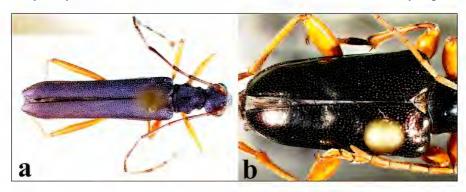
29(28).	Elytra (and	usually	pronotum)	metallic	iridescent	green,	blue,	or dark	purple (perhaps	
app	earing black	except u	nder strong	g illumina	ition)					3	0

- **30'.** Elytra with punctures very shallow (if deep, then elytra narrow, about 4 times longer than wide), regularly distributed, surface appearing smooth between punctures (b)......**33**



- 31'. Pronotum densely punctate, not rugose. Lateral pronotal tubercles moderately projecting......32

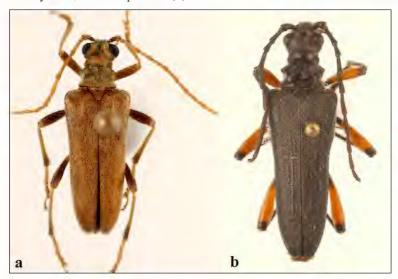


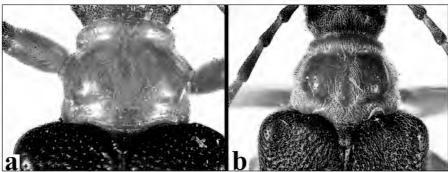


34(29°). Elytra (excluding epipleural edge) unicolorous, without vittae or maculae, or differently colored pubescent spots on dorsal surface
34'. Elytra (excluding epipleural edge) of at least two colors, with vittae or maculae, or differently colored pubescent spots on dorsal surface (or, if appearing as one color, with distinct vittae of pubescence)
35(34). Metepisternum thin, elongate (about 4 times longer than wide) (a)
35'. Metepisternum thick, short (about 2.5 times longer than wide) (b)
36(35'). Pronotum longer than wide; anterior constriction about one-third overall length of pronotum (a)
36'. Pronotum as long as wide; anterior constriction less than one-fourth overall length of pronotum (b)
a
37(36). Antenna with third antennomere subequal to fourth (a)Stenocorus cylindricollis (Say)
37'. Antenna with third antennomere distinctly longer than fourth (b)38

38(37'). Elytra color tawny, light brown, to red-brown (a) Stenocorus cinnamopterus (Randall)

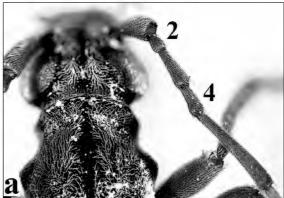


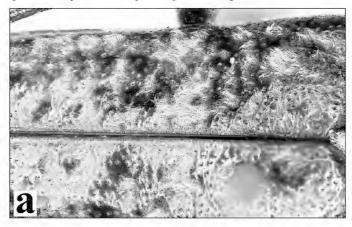






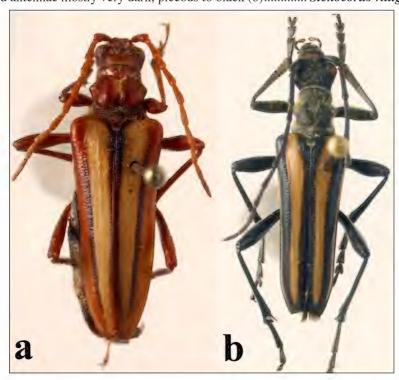
41(40'). Fourth antennomere very short, little more than twice as long as second (a)	
	nte
41'. Fourth antennomere at least 3-4 times as long as second	.42

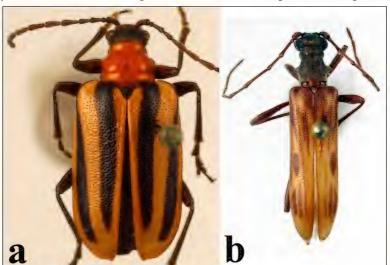


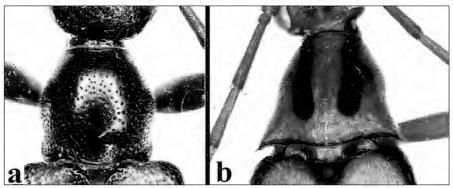




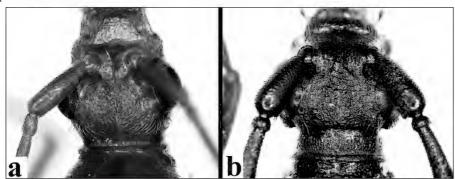




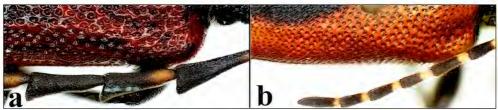


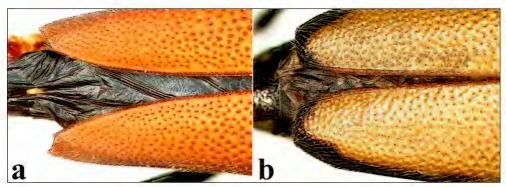


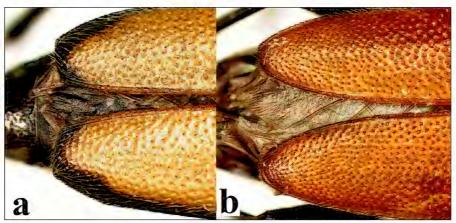
50(49). Head with a slight, gradual constriction behind eyes (a). Elytra vittate do	rsally 5 7
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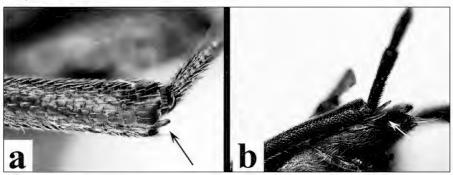
51'. Antennomeres weakly or not serrated (b). Elytral apices rounded, truncate or acuminate, not bidentate. Elytral and pronotal punctures not as pronounced as in former (b)......**52**

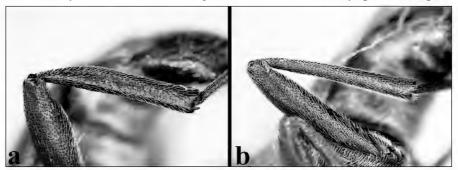




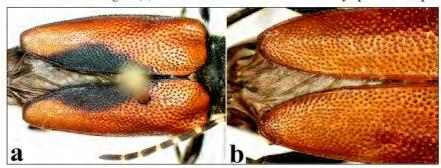


- 54(53'). Hind tibia with one tibial spur (a); last ventrite with deep medial impression (males).......55



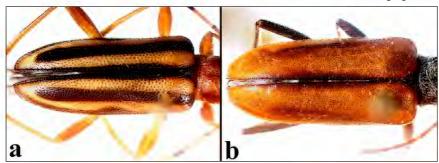


56(54). Elytra with margin (at least) black (a) (this character is not completely reliable, and
V	riant forms exist which cannot be distinguished from B. champlaini)
	Brachyleptura vagans (Olivier)



57(50). Venter pale testaceous. Elytra with 2-3 black vittae (rarely very faint) (a). Males with distinct tooth and curvature in inside margin of front tibiae..... Metacmaeops vittata (Swederus)

57'. Venter dark. Elytra with, at most, a vague sutural and/or epipleural vitta (b). Males without



- 59(58). Head with region at eyes much wider than base (a); basal antennomeres not expanded at
- 59'. Head with region at eyes only slightly wider than base; basal antennomeres strongly expanded at apex; large species, most specimens over 2 cm long.....



60(58'). Elytra uniformly orange colored except for apex which is abruptly black (a).	Middle tarsi
and claws of males thickened compared to hind tarsi (Trigonarthris)	61

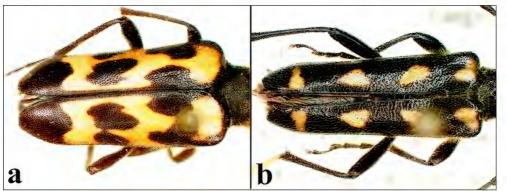


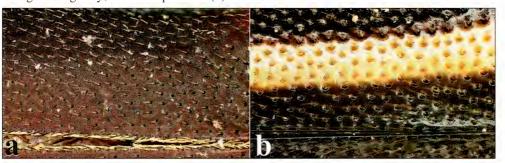




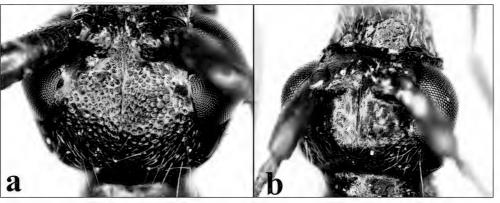


Key
64(63'). Elytra with well defined maculae or vittae of contrasting color from adjacent regions65
64'. Elytra generally unicolorous, sometimes with very poorly delineated maculae with edges that gradate into adjacent color
65(64). Legs uniformly pale reddish-brown
65'. Legs, at least in part, dark brown to piceous or black
66(65). Head, pronotum, and venter covered with dense, golden pubescence (a)
66'. Dense, golden pubescence sparse or absent from head, pronotum, and venter
67(66'). Head and pronotum uniformly reddish-brown (a)
67'. Head and/or pronotum, at least in large part, piceous or black (b) Pidonia densicollis (Casey) a

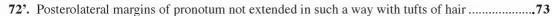


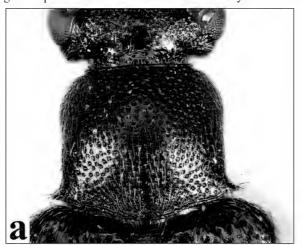


71'. Vertex of head between eyes without noticeable punctures (b)................. Pidonia ruficollis (Say)



72(64'). Posterolateral margins of pronotum extended into long tu	fts of hair giving base of pronotum
a wider appearance (a)	Alosternida chalybaea (Haldeman





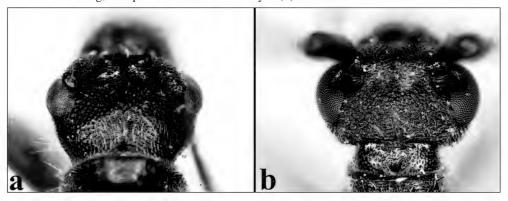


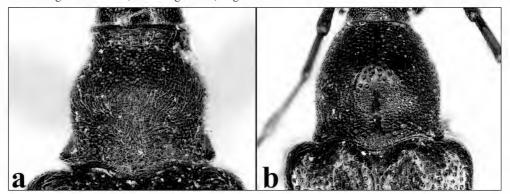


75 (74'). Dorsal apex of metatibia strongly extended along base of first tarsomere (a)

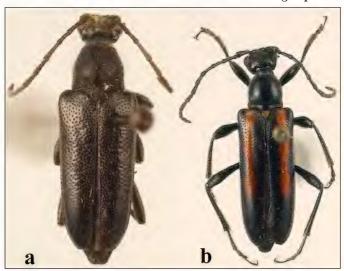


76(75'). Head only slightly and gradually constricted behind eyes (a)Acmaeops proteus (Kirby)

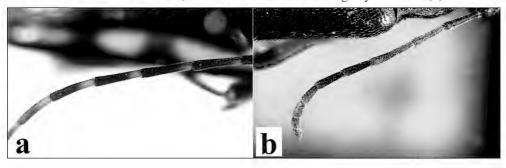




78 (77').			(see b above)			
	 	 	 	noplodera _l	pubera (Say

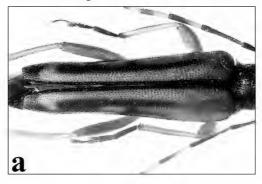


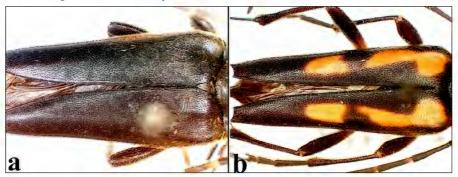
79'. Most antennomeres unicolorous, at most 1-2 antennomeres vaguely bicolored (b)83



80(79). Femora pale testaceous, unicolorous. Elytra long and narrow, 5 times as long as wide (a)

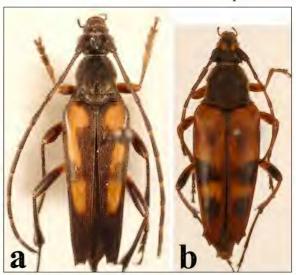
**Analeptura lineola (Say)

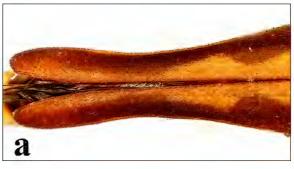




- **82'.** Metatibia bicolored, pale brown at basal three-fourths, dark brown to black at apex (b)......

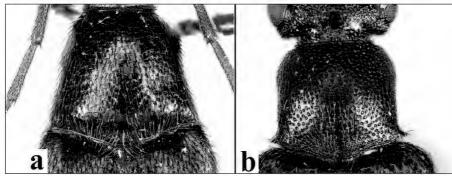
 **Leptura obliterata deleta (LeConte)





Ke	y
4(83'). Elytra uniformly very dark above, appearing black	5
4'. Elytra either with maculae (bicolored), or if unicolorous, not dark (either reddish or testaceous)91	l
5(84). Head very dark	5
5'. Head reddish89	9
6(85). Legs in part pale testaceous	7
6'. Legs uniformly dark	3
7(86). Pronotum often red (if black, then scape with ventral surface reddish). Clypeus, labrum, lower part of frons, basal palpomeres in most part, pale testaceous (a) (note: the uncommon species with a bicolored red and black pronotum, <i>Grammoptera exigua</i> (Newman), is excluded from this key))
7°. Pronotum always black (scape entirely black). Clypeus, labrum, palpi usually very dark (b) Grammoptera subargentata (Kirby))



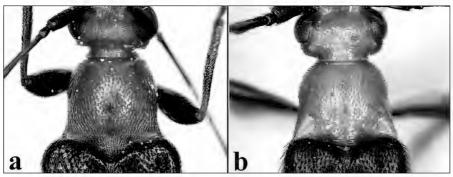


89(8	35').	Antennae and	legs pale	testaceous	to reddish	(a)) Stran	galia	bicolor ((Swederus	3)
------	-------	--------------	-----------	------------	------------	-----	---------	-------	-----------	-----------	----

89'. Antennae and legs very dark......90



90°. Postero-lateral angles of pronotum strongly produced, lateral angles elevated and strongly impressed at base (b). Elytra without strong glossy appearance......



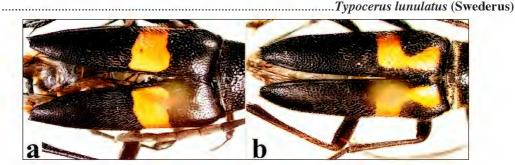
91(84'). Pronotum with broad, black vitta at middle, reddish at sides, with that color continuing to humeri of elytra (and sometimes throughout basal two-thirds of elytra (a, both color forms) (photos courtesy of Florida State Collection of Arthropods).... *Lycochoriolaus lateralis* (Olivier)

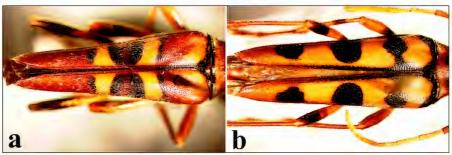


	elytra non metallic yellow (a). Large species, greater than 2 cm long
	without above coloration. Non-metallic colored species. Most species less93
	a
pubescence (a).	pt for black apical fourth or less, maroon and covered with dense velvety carge species, most specimens about 3 cm long
93'. Elytra without a	sove coloration and pubescence. Species less than 2 cm long94
	dle of sides and entire venter light reddish, otherwise black (a)
94'. Elytra without a	sove coloration; venter generally black or dark reddish99
	black except for single yellow to orange macula (either in form of band or sal half of elytra96

96(95). Elytron with single, broad, yell	ow to orange antemedial transverse band (a) (known	only
from Florida	Typocerus fulvocinct	us Knull

96'. Elytron with single right-angle yellow to orange macula at base (b)......

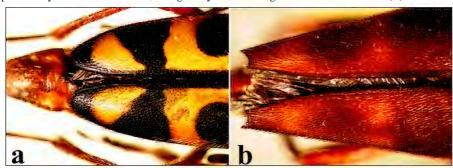




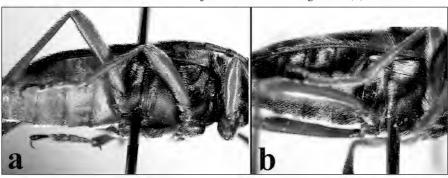


101(97'). Apex of elytra (not just margin edge) black with a distinctly contrasting yellow macula (or
ground color) at its anterior border (a). Elytra further ornamented with a series of approximately
spaced black or yellow maculae
•

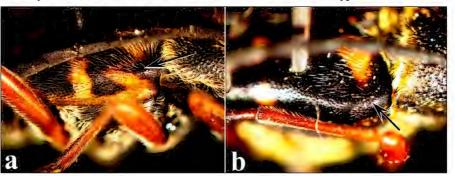








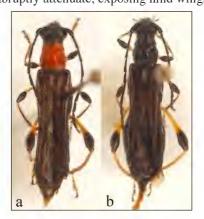






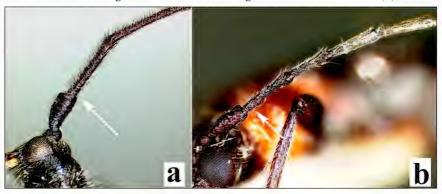
107 °. Pronotum cover	red, especially on posterior	margin, with conspic	uous, dense, golden pubes	scence
(posterior groov	e present on pronotum) (b)			108
a		b		
	e without distinct round o			
	distinct round or oval por			
a		b		
	ith suture forming base o			
	strongly produced outer s	spine; truncate, oblic		ate (b)
a		b		
·	reviated, exposing much o	_		
110'. Elvtra entire, e	exposing, at most, only sm	all portion of hind w	/ing	117

111(1	110). Elytra abruptly attenuate at midpoint, exposing hind wings at middle and apex (a, b)	
		(vier
111'.	Elytra shortened but not abruptly attenuate, exposing hind wings apically	112

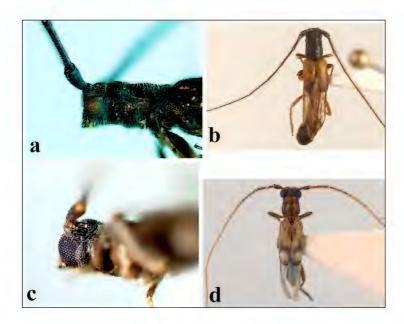




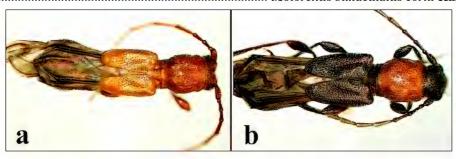
113(112'). Second antennomere very small, less than one-tenth length of third antennomere (a)...114
113'. Second antennomere long, at least one-fourth length of third antennomere (b)......115



114(113). Upper and lower eye lobes completely separated (a, b) *Tessaropa tenuipes* (Haldeman) 114'. Upper and lower eye lobes touching, connected by one row of facets (c, d)......



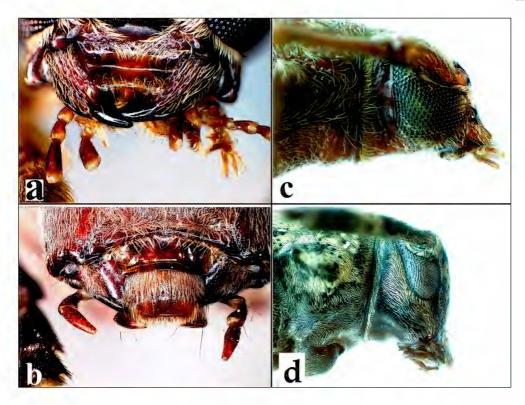




117(110°). Elytron with single antemedial transver	se or oblique, raised ivory callus118
117'. Elytron without single, transverse or oblique,	, raised ivory callus120
118(117). Pronotum with granulate, without define nearly always transverse (a)	ed longitudinal rugae; ivory callus of elytron Euderces reichei reichei LeConte
118'. Pronotum with distinct, longitudinal rugae; (see 119a-c)	ivory callus of elytron nearly always oblique119

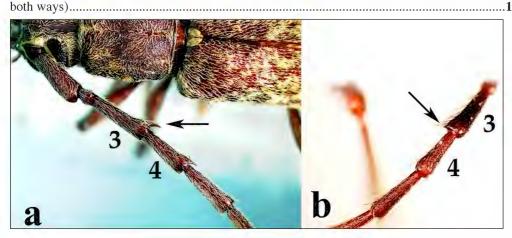


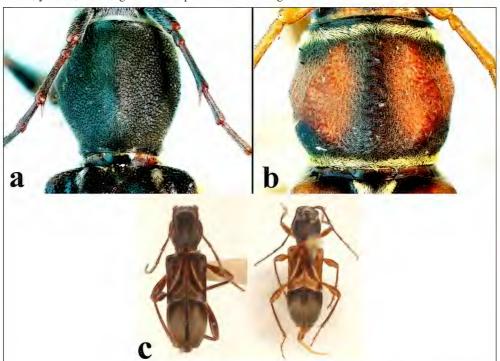




121(120). Antennae with mesal spine on apex of third antennomere (and almost always 2-5 more antennomeres, at least) (a) (Elaphidiini, plus *Megacyllene*, *Glycobius*, *Dryobius*, *Cyrtophorus*)

121'. Antennae without mesal spines. Specimens occasionally with an acute dentiform projection at apex of third antennomere (b), but not a spine (note: species with ambiguous states are treated in both many).

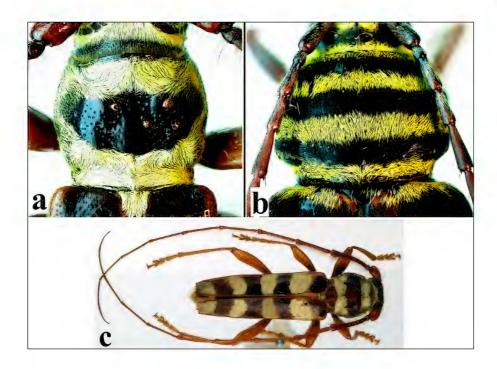


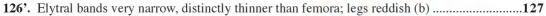


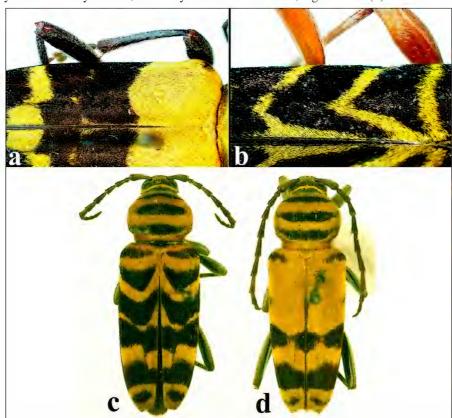
- 124'. Legs mostly glabrous. Bands of pronotal pubescence mostly complete across disk125

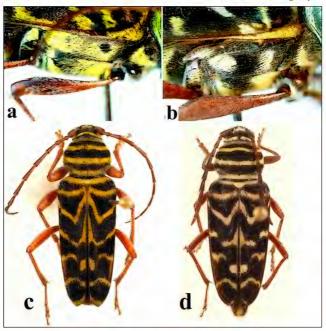


- 125'. Pronotum with 3-4 bands of yellow or white pubescence (b). Elytron with pubescent bands of uneven size, shape, and placement. Antennae at most as long as the body126

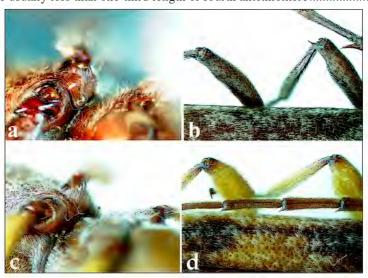




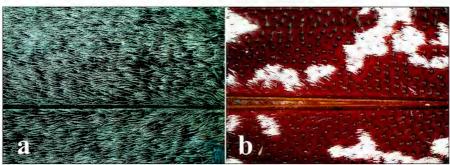


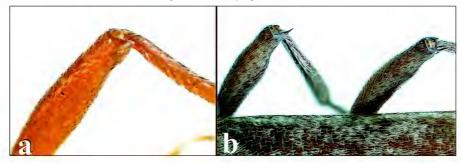


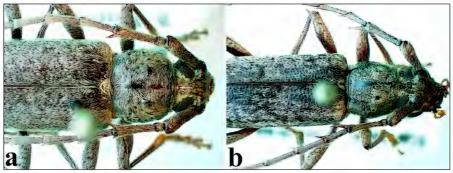
128(122'). Prosternal process straight and level to posterior margin of procoxae, and then abruptly vertical (a). Mesofemoral and metafemoral apices with spine or prominent dentate projection mesally (b). Elytral apices prominently bispinose. Several antennomeres usually bispinose. Spine of third antennomere usually very strong and in most specimens at least one-third length of fourth antennomere (Elaphidion)
129



129(128).	Elytral	pubescence mostly	uniform uniform	without lar	ge, glabrous	, shiny	regions	(a)	130
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132'. Integument color of legs reddish brown (lighter in color than integument elsewhere). Femora mostly covered in closely spaced patches of white pubescence. Outer femoral spines usually weak on meso- and metafemora (c). White pubescence of pronotum very dense around lateral margins of peripheral calli, hiding punctures (d) (known only from Florida in the US)

**Elaphidion irroratum* (Linnaeus)*



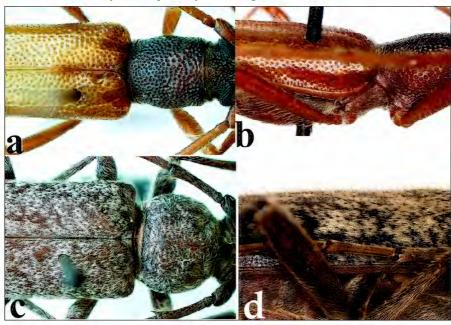
133(128'). Integument testaceous, mostly glabrous, but with distinct patches of very dense, bright white pubescence on head, pronotum and elytra (a) (known only from Florida in the US)

Linsleyonides albomaculatus (Champlain and Knull)

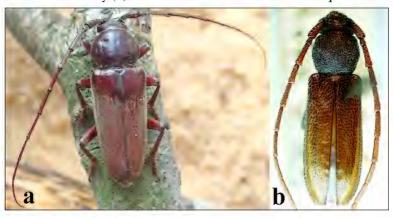


134(133'). Integument mostly testaceous to light reddish brown. Only scattered erect hairs (always	
separate from each other) present on pronotum and elytron, not obscuring any of surface (a, b)	
1	3

134'. Integument mostly dark reddish-brown to black. Pubescence either patchy and more dense in places on pronotum and/or elytra (but these scattered, dense portions may be very sparsely distributed), or uniformly dense, partially obscuring surface (c, d) (see also 147a-b).....141



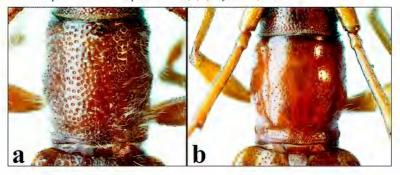
- 136(135). Large, robust longhorn (most > 3 cm). Pronotum without punctures. Spines of antennae very small, appearing dentiform on 3 and 4 mesally (a, photo courtesy Roy Morris) (known
- 136'. Small longhorn (most 1-2 cm). Pronotum with dense punctures. Spines of antennae moderately



137(135').	Pronotum with lateral projec-
tions at	middle and a strong posterior
constrict	ion. Femora pedunculate/clavate
(a) (kno	own only from Florida)
	Stizocera floridana Linsl

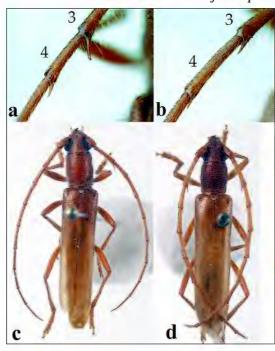


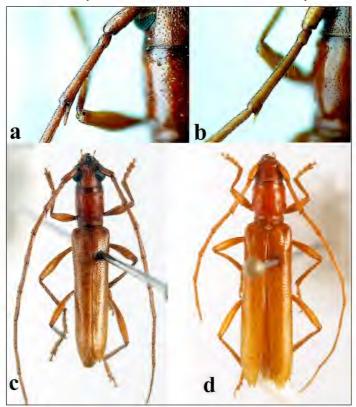
137'. Pronotum without lateral projections, parallel sided or gradually swollen at middle. Femora gradually enlarged, neither clavate



139 (138). Spine of third antennomere almost half length of fourth antennomere; somewhat blunt at apex (a). Pronotum only slightly darker than remaining integument (c)......

139°. Spine of third antennomere about one-fourth length of fourth antennomere or less; usually acute at apex (b). Pronotum distinctly darker than remaining integument (d).....



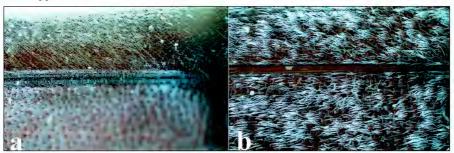


141(134'). Prothorax red or orange except for black circular region at center of disk (rarely entirely red or orange), remainder of body and appendages black (or rarely dark reddish brown)

Stenosphenus notatus (Olivier)

141'. Prothorax light to dark reddish brown, not of contrasting color to remainder of body.......142



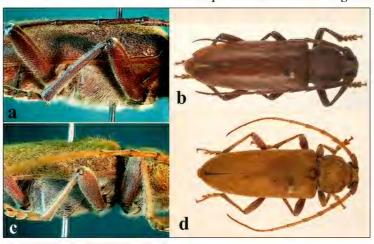


- 143(142). Large, robust beetles, greater than 2 cm in length. Elytral suture with moderate to strong spine ________144

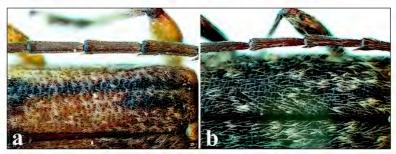


- **144(143).** Body and appendages very dark reddish brown. Ventral pubescence mostly translucent with slight golden sheen (a, b). Last ventral sternite of females with a deep notch at apex *Enaphalodes hispicornis* (Linnaeus)
- **144'.** Body and appendages light reddish brown. Ventral pubescence mostly white and not translucent, distinctly different color from dorsal pubescence (c, d). Last ventral sternite of females with a very shallow notch at apex (known only from Florida)......

Enaphalodes archboldi Lingafelter & Chemsak

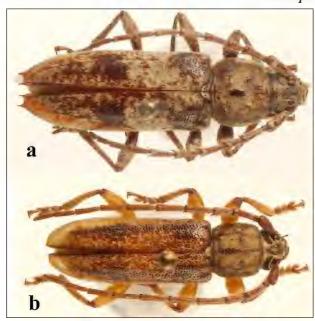


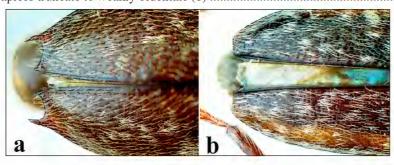
145(142').	Elytral	integument	t with two c	olors (testa	ceous or	reddish bro	own & d	ark brown),	giving
a maci	ulate or	banded app	earance (a)	. Antennae	with str	ong dorsal	carina ((a)	146

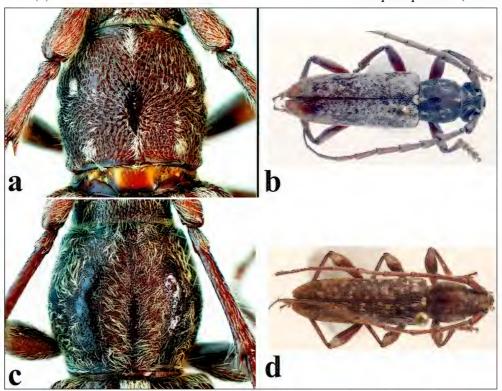


146'. Elytra without apical spines; rounded to suture (b) (known only from Florida in the US).

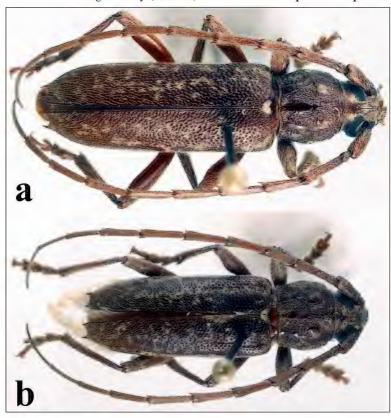
**Anelaphus cinereus (Olivier)

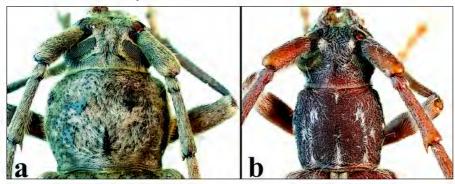


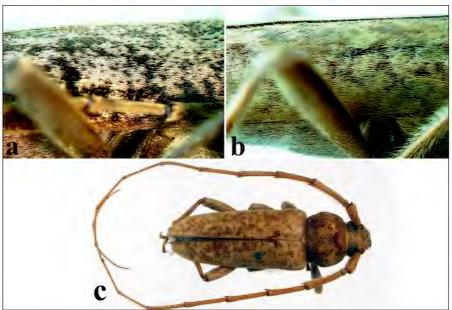




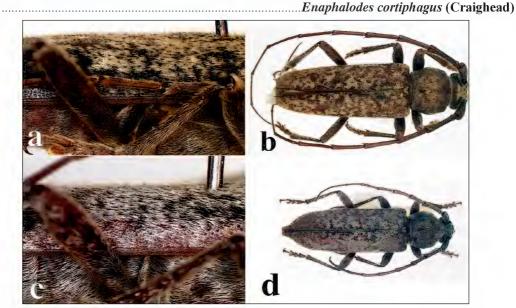




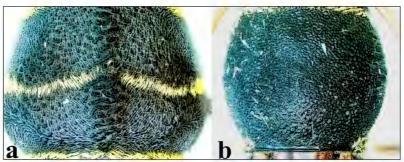




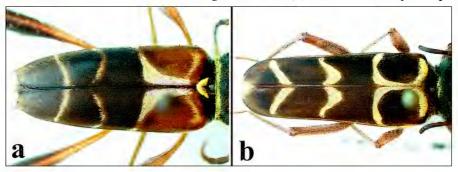
- **153(152).** Erect setae much more abundant (a). Last abdominal sternite of female deeply notched at apex. Elytra with diffuse patches of off-white pubescence (b).. *Enaphalodes atomarius* (Drury)
- **153'.** Erect setae more sparse (c). Last abdominal sternite of female with very shallow notch at apex. Elytra with diffuse patches of white or off-white pubescence (d)......

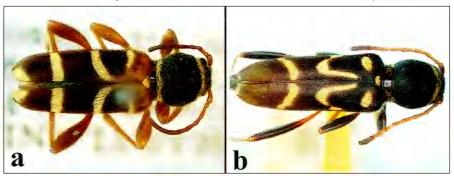


					parallelus (Newn
					100
a			b		
		(Alle)			
c	•		d		
21 2) Propotum	with at least o	ne conspicuous	transverse ha	nd of vellow or	white pubescence
lytra with white	yellow, or ora	ange bands of p	ubescence thro	oughout (e.g., se	ee 123b & 125a-b
	•		•	•	r without bands of
					at least) and some



- **158(157).** Elytron with bold yellow to white band of pubescence that extends from outside edge just antemedially to suture in a straight line and along suture to scutellum in a straight line (a). Epipleuron to base of elytron without pubescent fasciaNeoclytus mucronatus (Fabricius)

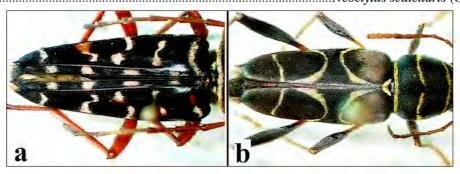


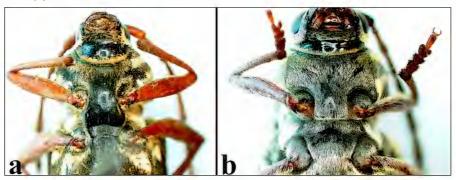


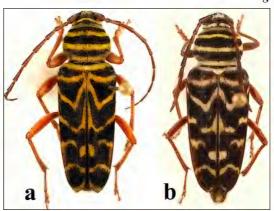








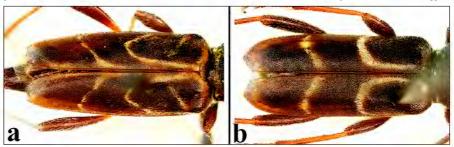


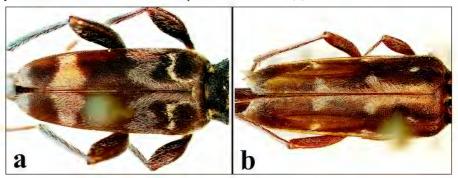


166(155'). Front of head with carinae extending from antennal tubercle straight to frontal margin. Middle of frons with mostly glabrous, raised region bordered by variably developed carinae, often forming a "V" or "U" shape (a). Antennae short, at most reaching only to middle of elytra (<i>Xylotrechus</i>)
166'. Front of head without distinct carinae and swelling (b). Antennae of variable length175
a
167(166). Elytra testaceous, much lighter than pronotum
167'. Elytra reddish brown or very dark brown, of similar color to pronotum
168(167). Pronotum with white or yellow anterior and posterior spots, usually bold (a). Elytron with anterior and antemedial white to yellow narrow fasciae close together, if apparent (a). Lateral portion of mesosternum without bold white to yellow pubescence
a
169(167'). Anterolateral margin of pronotum with distinct, isolated fasciae of yellow or white pubescence (a). Frontal carinae borders and elytra with distinct, narrow fasciae of yellow or white pubescence
169'. Pronotum with diffuse or large patches of pubescence, not isolated in anterolateral region (b). Elytra and frontal carinae margin with fasciae of pubescence more diffuse or in broader bands

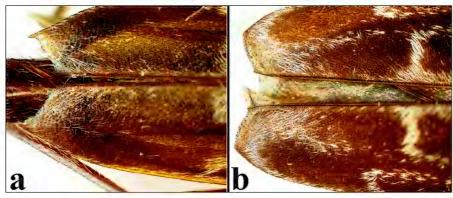
170(169)	. Elytron with anterior fa	scia extending posteriorly	y from near scutellum to lat	eral edge of
elytr	on at basal one-third (a).	Antennae black	Xylotrechus	nitidus (Horn)





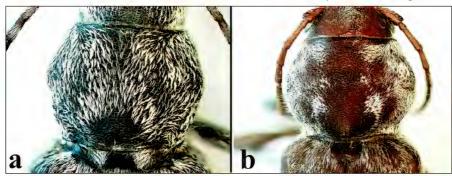


173'. Outer elytral apex at most dentiform (b). Fasciae of elytra not concentrated along suture....174



174'. Pronotal disk center mostly without white or tawny pubescence, exposing granulae (b)......

Xylotrechus integer (Haldeman)

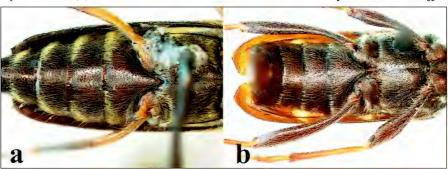


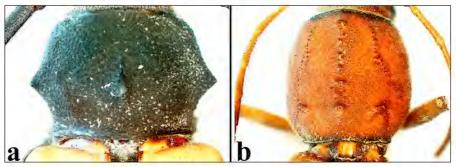
175(166'). Anterior margin of pronotum (and sometimes elsewhere) with an isolated, divided band of yellow or white pubescence (a). Antennae short, not surpassing apical third of elytra176



176'. Legs mostly glabrous. Pronotum densely granulate. Small species, less than 1.5 cm177







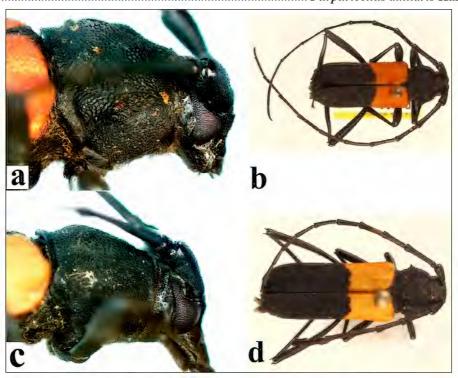
Key
179(178). Elytra bright metallic green or blue (rarely reddish) (a). Femora, except for apex, pale red; remainder of legs black
179'. Elytra without metallic coloration. Legs differently colored (uniform in most species)180
180(179'). Most antennomeres distinctly and boldly bicolored, yellow at base and black at apex (a) (known only from Florida in the US)
180'. Antennae uniformly colored



181(180'). Posterior half, appropriation of elytra does not experience.			
181'. Elytron differently colore	•	. ,	
	same.		

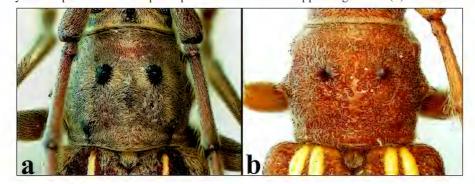
182(181). Scutellum the same color as elytral base, either yellow, orange, or red (see <i>181a</i> above) (known only from Florida in the US)	
182'. Scutellum black, contrasting sharply from color of basal region of elytron	183

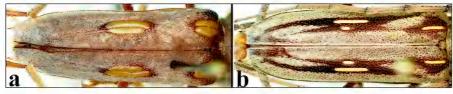
183 (182'). Pronotum with strong, multiple dorsal tubercles throughout disk (a, b)
Purpuricenus paraxillaris MacRae

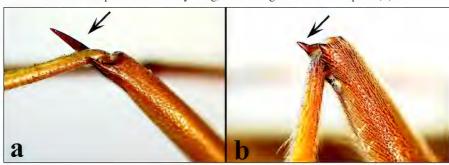


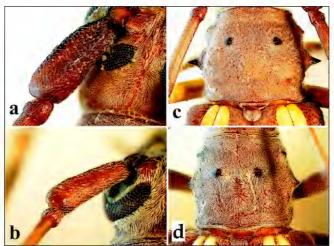




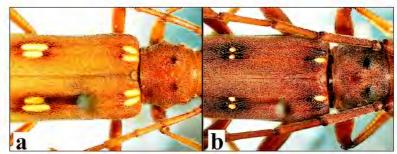








190'. Elytra and pronotum dark brown. Eburneous ridges small and less bold as in former, with distinctly darker peripheral (and sometimes costal) regions (b) (note: these are highly variable and anomalous forms do occur. *E. haldemani* could be a synonym of *E. quadrigeminata*)......



191'. Head and pronotum differently punctured. Femora linear to very slightly enlarged192

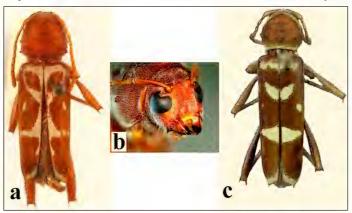


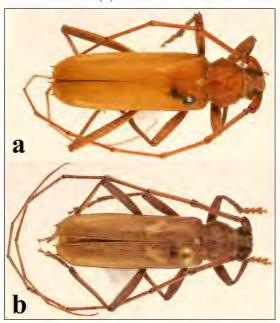
192(191').	Elytra mostly pale green (a) (possible in Florida, not known elsewhere in the U. S.)
192'. Elyti	ra of differing color





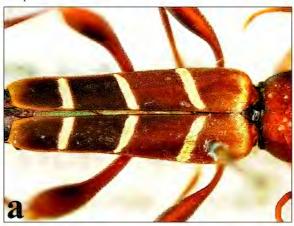




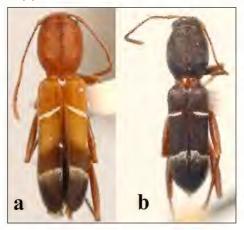


198 (197). Extreme	e anterior margin of pronotum at middle produced into elevat	ted crest, higher than
other ridges (a)). Sides of prothorax with numerous erect, white hairs. Metati	bia of males about as
long as body	Eurys	<i>celis suturalis</i> (Olivier







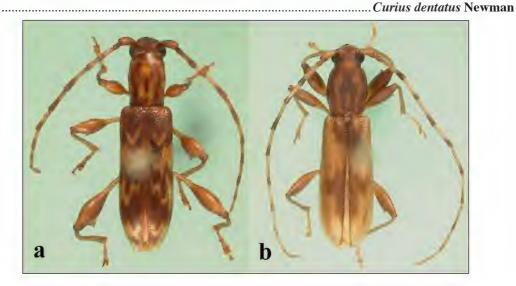


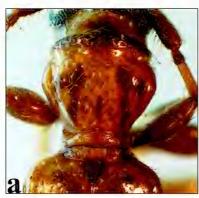




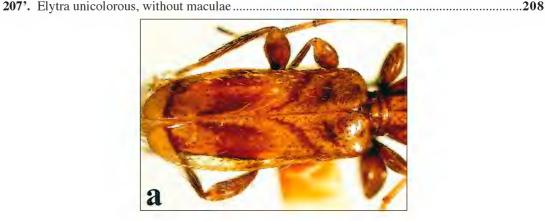
205(204).	Fourth antennomere about half as long as scape (a). Pronotum nearly parallel-
sided	n posterior third (a, photo courtesy Gino Nearns). Integument glossy

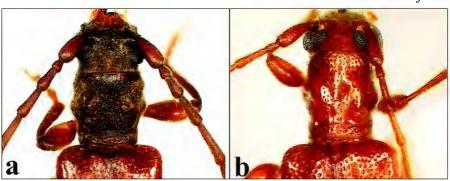
205'. Fourth antennomere about as long as scape or longer (b). Pronotum evenly rounded at sides, without constriction (b, photo courtesy Gino Nearns). Integument not shiny......





207(206).	Elytra maculate	(a)	 Obrium	maculatum	(Olivier)

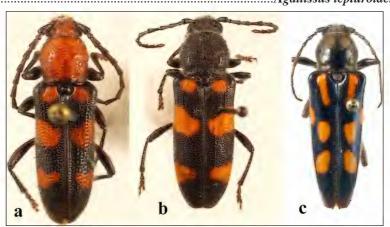






211'. Elytron with appoximate posterior half black and anterior half red or yellow (Heterops dimidi	
a	
212(209'). Basal epipleural region of elytron with abrupt, upward constriction over m (a) (Agallissini)	
212'. Epipleural region of elytron without constriction	215
a	
213(212). Pronotum with four partial to complete, narrow, longitudinal white fasciae (a from Florida and Georgia)	

- 214(213'). Pronotum dull, with dense, coarse, deep punctures (a, b) (known only from Florida and
- 214'. Pronotum shiny with very few, sparse, shallow punctures (c)......



- 215(212'). Femora very long and strongly pedunculate-clavate. Non-clavate portion of metafemur
- 215'. Femora linear, gradually enlarged apically, or clavate, with, at most, a small peduncle216



216(215'). Entire body densely clothed in yellowish pubescence that is much lighter than integument (broken somewhat by pattern of small glabrous maculae on elytra.) Vague broad,

longitudinal vittae formed on pronotum. Pronotum widest anteriorly, gradually tapering posteriorly (a)..... Atimia confusa (Say)

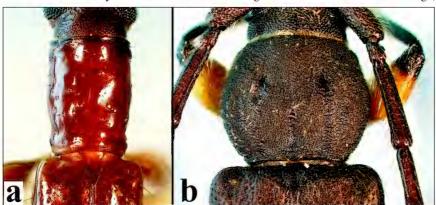
216'. Body mostly glabrous or with diffuse pubescence not obscuring most of surface. Most species without pubescent vittae on pronotum. Pronotum widest at about middle or sub-



217(216'). Pronotum black except for two orange longitudinal vitta	e around broad disk region. Vittae
connect with orange-yellow base of elytra that tapers along epi	pleural edge. Remainder of elytra
black, usually extending up along suture to near scutellum	(a)
	Elytroleptus floridanus (LeConte)
217'. Pronotum and elytral coloration different	218



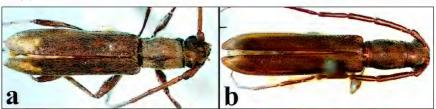
218'. Pronotum short, broadly rounded at sides, little longer than wide or wider than long (b).....221



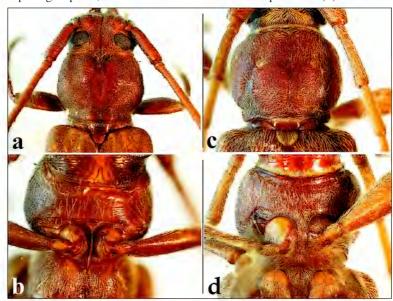


220 (219'). Pronotum and elytra with moderately dense pubescence. Elytra reddish-brown, some-	
times with vague posterior pale macula (a) (known only from Florida)	
Hatangalith as authoris Dlatal	.1

220°. Pronotum and elytra with sparse pubescence. Elytra dark reddish brown to black, without



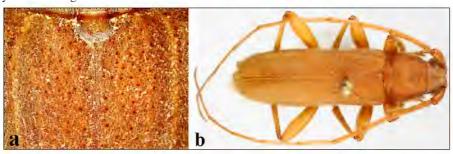
- 221(218'). Entire integument and appendages all uniformly testaceous to light reddish brown. No
- 221'. At least two colors present on appendages, pronotum, and/or elytra, or integument uniformly
- 222 (221). Pronotum with strong and abrupt basal constriction (a). Prosternal process as a thin,
- 222'. Pronotum without strong, abrupt basal constriction (c). Prosternal process either expanded



- 223(222'). Body very dorso-ventrally flattened (a). Tibiae shorter than clavate portion of femora. Femora strongly clavate nearly to base. Pronotum semiquadrate, not evenly rounded at sides....
- 223'. Body not strongly flattened. Tibiae longer than clavate portion of femora, or, if not clavate, about as long as femora or longer. Pronotum approximately, evenly rounded at sides......224

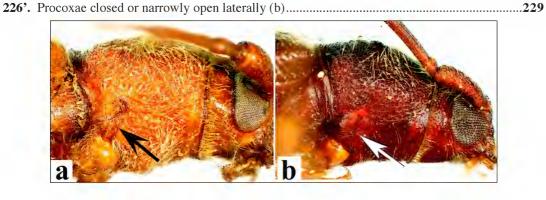


224(223').	Elytra	with	scattered	small	dark	granules/asperites	around	basal	one-third	l (a,	b)
(introdu	iced fro	m Eu	rope into l	North A	Ameri	ca)	Str	romati	um fulvu	m (V	/illers)



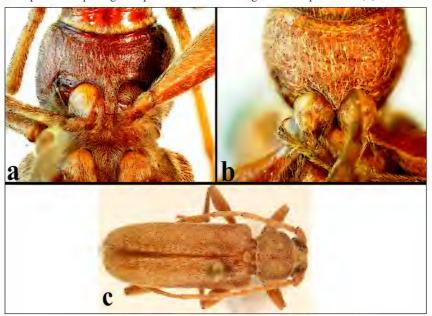
225'. Elytra not antennuate, not exposing hind wing along suture. Antennae 11-segmented226



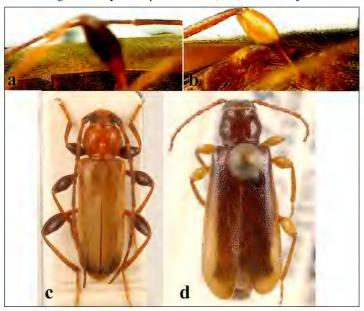


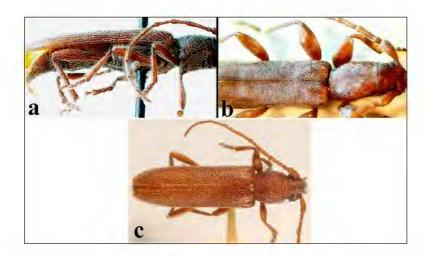
227(226).	Procoxal	process	wide	between	procoxae,	not	tapering to a	point	between	them (a);
habitu	s as in figu	ıre (c)					Hesperoph	anes p	oubescens	(Haldeman

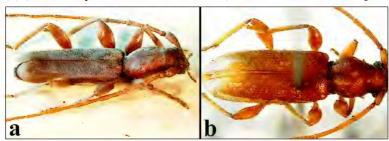
227'. Procoxal process tapering to a point and terminating between procoxae (b)......228



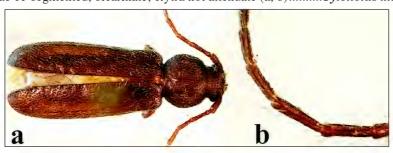
228'. Elytra covered in longer, mostly erect pubescence (b, d).............Phymatodes aereus (Newman)



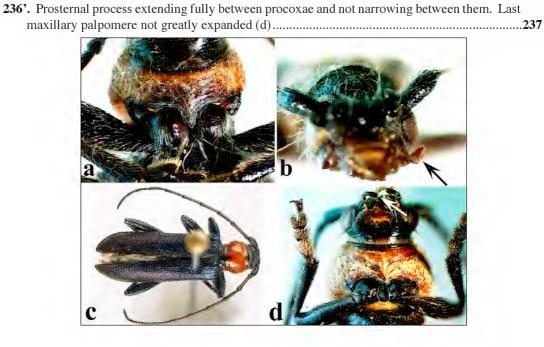




- 232'. Antennae 11-segmented, bicarinate; elytra not attenuate (a, b)...........Tylonotus masoni (Knull)



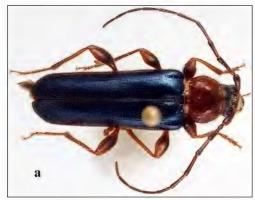
233(231'). Pronotum unicolorous red, orange, or reddish brown, elytra unicolorous dark blue or black (pronotal margins rarely partially black)
233'. Coloration of pronotum and/or elytra different
234(233). Pronotum angulate at middle of sides, much wider than long; densely punctate, semirugose, dull (a)
234'. Pronotum rounded at sides; sparsely punctate, shiny, smooth (b)235
a b
235(234'). Legs uniformly dark blue or black
235'. Legs uniformly or partially pale testaceous
236(235). Prosternal process short and narrowed to a point between procoxae (a). Last maxillary palpomere greatly expanded (b); habitus as in figure (c) Pronocera collaris (Kirby)



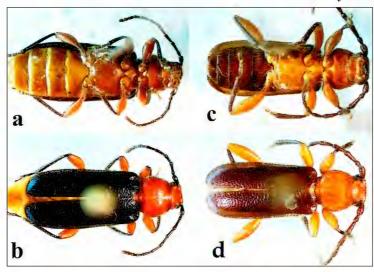
237'. Most elytral pubescence about as long as that of pronotum, erect or suberect throughout (b) (note: probably, this should *not* be recognized as a separate taxon from the nominate form)

Batyle ignicollis australis Linsley



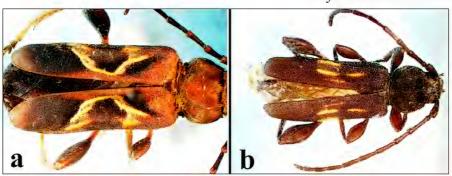


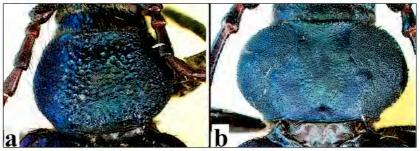
239'. Abdominal venter slightly darker, in part at least, than remaining venter which is pale reddish or testacous (c). Pronotum with weakly perceptible angle at middle of sides (d)......

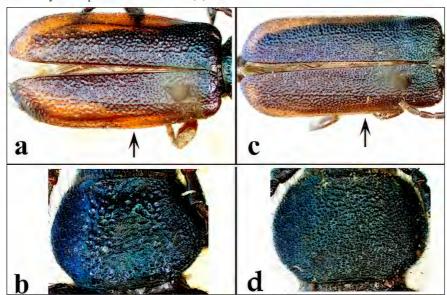


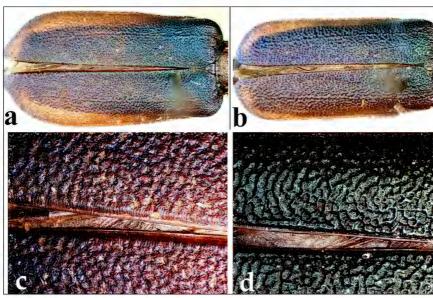
240(233').	Elytra with series of long,	, curved, raised ivory	calli (a) (Physod	cnemum)	241
240°. Flyti	ra without ivory calli				242

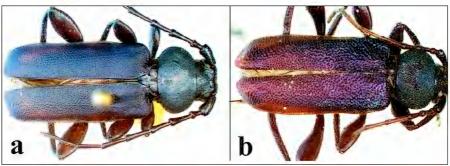












no	2'). Elytra with distinct pale orange, yellow, or white maculae (formed by integument color, t pubescence) on a darker background. These maculae form transverse bands or spots that an (or nearly span) width of elytra
	Elytra differently colored or patterned, often with white or yellow fasciae of pubescence on larker background
	7). Each elytron with a narrow antemedial and postmedial transverse white band on a ostly black background (anterior one-third of elytron sometimes red or reddish-brown) (a) **Phymatodes varius* (Fabricius)*
248'. I	Pale elytral maculae in different arrangement 249



249(248'). Each elytron with a basal transverse yellow band and a	n oblique antemedial band bordered
by a narrow black margin (see 211a)	
	Aethecerinus hornii (Lacordaire)
249'. Pale elytral maculae in different arrangement	250

250(2	49'). Anterior two thirds of elytra yellow, in-
te	errupted by a single black macula at middle,
a	ndapical one-third entirely black (a)
	Semanotus ligneus (Fabricius)
250'	Pale elytral maculae in different arrange

250'. Pale elytral maculae in different arrangement.......251



251(250'). Each elytron with an antemedial and subapid	cal oval, yellow macula on a reddish brown
background (a)	Tylonotus bimaculatus Haldeman
251 °. Pale elytral maculae in different arrangement	252



252'. Pronotum dull with numerous small asperites. Integument pale reddish brown with an antemedial and sometimes subapical pale macula on elytra (b)Penichroa fasciata (Stephens)





255(254'). Elytra with a series of at least three evenly spaced, undulating, narrow, white pubescent fasciae
255'. Elytral pubescent fasciae in different color and/or arrangement
256 (255). Pronotum densely covered in pubescence (a). Pubescent fascia of elytra diffuse. Antennae not reaching to midpoint of elytra
256'. Pronotum without white pubescence on large, circular, black region at center of disk (b). Pubescent fasciae of elytra distinct. Antennae reaching beyond midpoint of elytra
Sarosesthes fulminans (Fabricius) a b
257(255'). Pubescent fasciae of elytra in one to three very narrow white oblique or transverse bands. Small ant-like species
257'. Pubescent fasciae of elytra bold yellow, in different configuration
258(257). Elytron with a narrow, oblique, impunctate shiny band of integument at basal one third, bordered by very narrow white pubescent fasciae. Pronotum often with two arcuate, narrow longitudinal bands of white pubescence at base (a)
258'. Elytra without impunctate, shiny, oblique regions
259(258'). Elytra with one-two very narrow oblique bands of white pubescence (one postmedially and usually one antemedially). Apex of third antennomere unarmed (a)
259'. Elytra with elytral apex broadly white fasciate, with one-two narrow transverse medial white fasciae, and with one narrow, oblique white fascia extending from near scutellum to outer transverse medial fascia. Apex of third antennomere usually dentiform mesally (b)

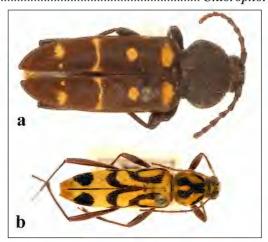
b

a

260(257').	Pronotum	without con	nspicuous	pubescence;	elytron	with four	evenly sp	aced yell	ow
pubesc	ent patches	s or bands (a)				Calloide	s nobilis	(Harris

260'. Pronotum with dense, yellow pubescence except for middle and anterolateral areas; elytron with dense yellow pubescent bands along suture, humerus, and most of apical one-third (b).....

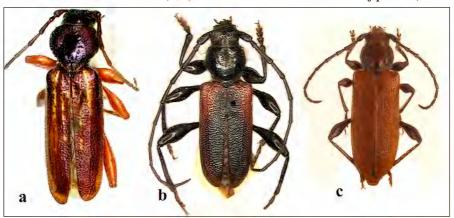
Chlorophorus annularis (Fabricius)

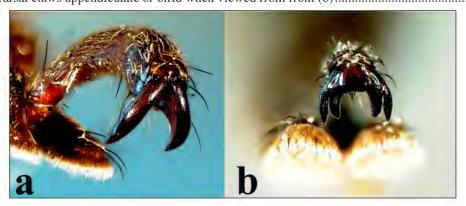


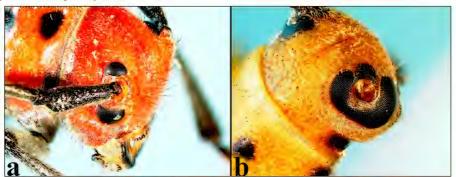




263(262'). Femora orange. Pronotum much wider anteriorly than posteriorly. Most specimens with	
subtle metallic blue, purple, or violet sheen on pronotum and sometimes elytra (a)	
	y)



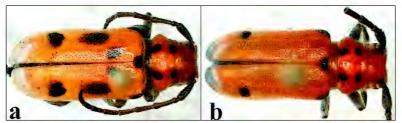


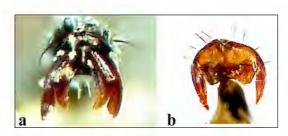


200 · Hoda and prone	otum mostly orange to reddish. Larger than 0.5 cm long	
	Danger of the control	
	a	
267(266'). Elytra con	npletely black, without maculations. Narrow bodied, about 2 mm	wide (a)
	eddish or orange with black spots or maculae. Larger and broader. (Tetraopes)	
	2 x to toll	
	a	
	onotum, and elytra covered in dense, white pubescence covering	
268 °. Integument with	h only sparse pubescence not obscuring surface	269
	-	
	a	
	· case	
269(268'). Elytra wit	h apical third and usually antemedial third with large angled black	or dark
	th apical third and usually antemedial third with large angled black	
gray macula (a)		Schoenherr
gray macula (a)	Tetraopes melanurus	Schoenherr
gray macula (a)	Tetraopes melanurus	Schoenherr
gray macula (a)	Tetraopes melanurus	Schoenherr

- **270(269').** Very long setae at apex of most antennomeres that are as long as adjacent antennomeres. Last antennomere abruptly flattened midway and slightly curved (a)......*Tetraopes texanus* Horn





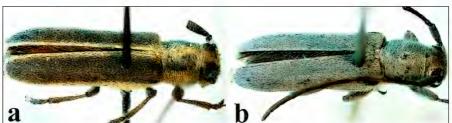


- 272(265'). Tarsal claws symmetrically bifid (a). All but one species mostly covered ventrally and dorsally with gray pubescence273
- **272'.** Tarsal claws asymmetrically appendiculate (b). Body never mostly covered ventral or dorsally with gray pubescence (*Oberea*).......**279**
- **273(272').** Third antennomere longer than pronotum. Elytra black, pronotum with broad black or dark reddish-brown fascia at center, connecting with triangular fasciae on occiput and vertex of head terminating in point between upper eye lobes (a) *Hemierana marginata* (Fabricius)

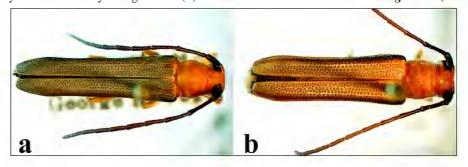


274(273'). Femora pale reddish, only sparsely covered in gray pubescence 274'. Femora dark, mostly covered in gray pubescence	270
275'. Elytral suture and outer edge without contrastingly colored pubescence (Mecas fe	
a	
276(274 ²). Pronotum with two small glabrous spots on disk (a)	27
276'. Pronotum without two small glabrous spots on disk (b)	278
a	
277(276). Suture and outer edge of elytron margined with bright white pubesce	
277'. Suture and outer edge of elytron without contrastingly colored pubescence	ee (b)

278(276').	Elytral	suture	and	outer	edge	margined	with	bright	white	or ye	ellow	pubesce	nce.
Pronotu	ım with	vague v	vhite	vitta	at mid	ddle and si	des (a)	1	Meca	s mar	ginella <mark>I</mark>	LeConte



279(272'). Pronotum without black calli or other marks. Red head, pronotum, and scutellum, most of venter, and femora (tibiae and antennae black)	80
279'. Coloration otherwise	81
280(279). Elytra above gray-brown to black (a)	us)
280'. Elytra above mostly orange to red (b)	as)

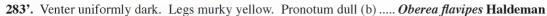


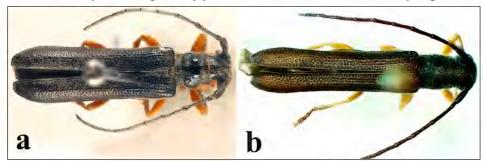
281(279'). Pronotum all or mostly dark
281'. Pronotum mostly pale yellow, red, or testaceous, with or witout black spots (if mostly dark
then two dark, discal spots are vaguely apparent)

282(281). Head mostly pale yellow or red (at least on frons). Pronotum usually with anterior margin
pale yellow or reddish. Venter and antennae mostly piceous except for legs and usually last
one-two ventrites which are pale reddish yellow (a)

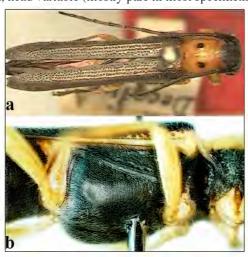
2822	Head and	1 propotum	mostly t	o entirely dark	Ventral coloration v	ariahle	283

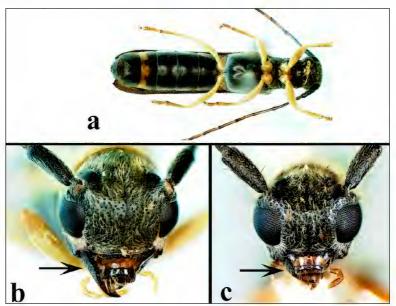






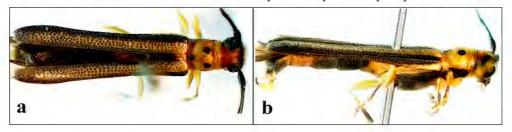






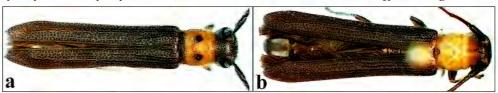
287(286'). Scutellum pale; some abdominal sternites pale; elytra with partially diffuse pale base and sides (a, b) (note: *O. deficiens* Casey is included here as there are no distinguishing characters. This should probably be synonymized with *O. praelonga* Casey)............Oberea praelonga Casey

287'. Scutellum dark. Abdominal sternites uniformly dark. Elytra mostly very dark......288



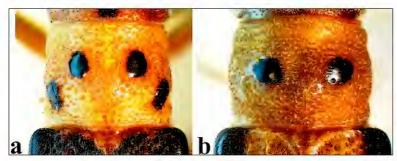
288(287'). Pronotum usually with two black calli near center. Hairs of elytra pale (a).......

Oberea perspicillata Haldeman

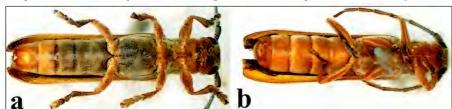




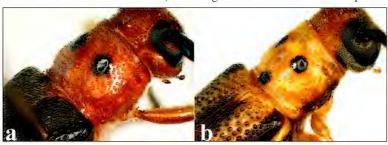
- **289(284').** Posterior margin of pronotum black or dark brown. Elytra and scutellum black or dark brown (a)..............Oberea perspicillata Haldeman



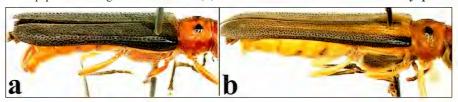
- **291'.** Abdominal sternites and head unicolorous, yellow to reddish (b). Head usually without dark markings. Posterolateral spot small, no larger than central spot *Oberea caseyi* **Plavilstshikov**



- 292'. Prothorax with four or five black calli (including one one each side above procoxa) (b)294



293'. Metepisternum mostly or all black; elytra mostly pale with vague darker infuscations along



294(292'). Head yellowish to pale reddish (never dark). Metepisternum black (a) (note: O. deficiens Casey is included here as there are no distinguishing characters. This should probably be

294'. Head dark reddish usually with dark black infuscations. Metepisternum red (see 285a) Oberea ulmicola Chittenden



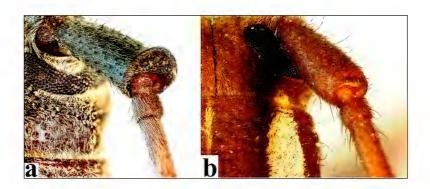
295(264). Integument shiny black with inconspicuous punctation (small, shallow punctures only apparent through high illumination and magnification). Elytra with reticulated, connected patches of very dense white pubescence. Pronotum with two longitudinal bands of white pubescence around central disk region. Pronotum with prominent, acute lateral projections. Antennal scape with

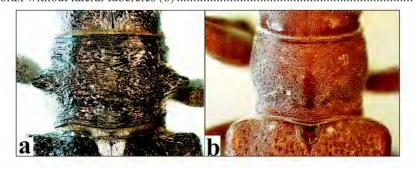
295'. Characters in different combination from above. Most species under 3 cm long296

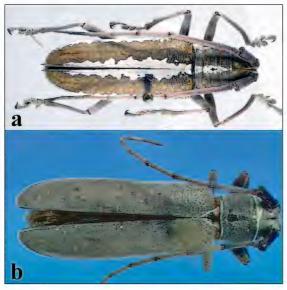


296(295'). Antennal scape with a distinct and obvious cicatrix at apex that is very exposed, somewhat flattened, and punctate or asperate and mostly free of pubescence (a) (Most Lamiini)297

296°. Antennal scape without obvious cicatrix (b) ________**313**

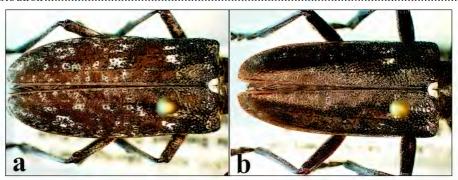










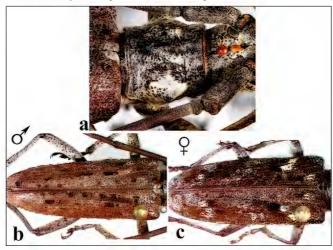


302(301').	Elytral	apex	with	suture	produced	into	weak	point	or	spine	(a,	b)	(most	Mono	chamus)
															303

302'. Elytral apex rounded, not produced into point or spine at suture (c) (most Goes)......306

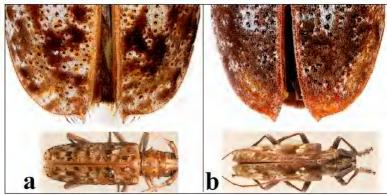






305 (304'). Suture produced into strong (but sometimes blunt) spine. light reddish brown integu-
ment with mottled patches of white, yellow, and dark brown pubescence (a)

305'. Suture produced into weak projection. Dark reddish brown integument with less distinct mottled patches of white, yellow, and dark brown pubescence (b)......



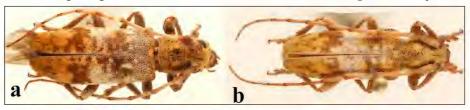
306(302'). Much of dorsum with pale orange pubescence interrupted by transverse sections on elytra of darker brown and/or white pubescence (e.g., see *307a*)**307**

306'. Dorsum with pubescence of different color and/or distribution......**309**

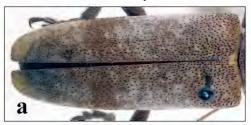
307'. Integument with white pubescence on portion or all of anterior half of elytra. Venter clothed in mixture of white and pale orange pubescence......**308**

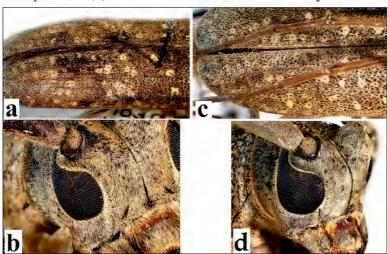


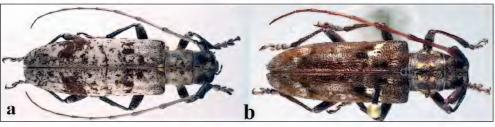
308(307'). Most of basal half of elytra with mottled white pubescence (a)......Goes debilis LeConte



309(306'). Elytral pubescence light colored only (white or very light tan) (e.g., see	ee 310a) 310
309'. Elytral pubescence includes either some small patches or extensive cove	ring of brown
pubecence, in addition to light colored pubescence	312









- 313(296'). Head with unusual shape: triangular from lateral view with antennal tubercles sticking anteriorly and front angle and mouthparts removed to extreme posterior margin ventrally; as long as pronotum from antennal tubercles to posterior margin. Eyes small, round, far removed from antennae: about half way between antennal tubercles and mouthparts (a) (Spalacopsis, included species known only from Florida in the US)...........314





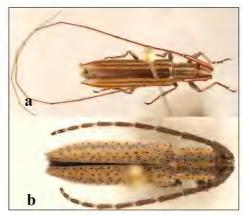


317(313'). Head somewhat triangular from lateral view, with antennal tubercles extended anteriorly,
and mouthparts posteriorly, but overall, head shorter than pronotum (a). Elytra with 2-3 longitudinal
vittae of lighter colored pubescence (golden-yellow or pale orange), that continue onto pronotum as
3-4 separate vittae (as in 318a, b). Eye emarginate around insertion of antenna318



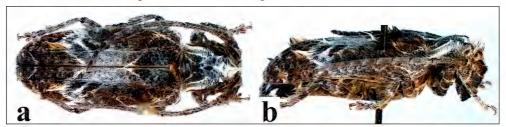
318'. Antennae short, about as long as body. Elytra with numerous long, erect black hairs (b).....

Dorcasta cinerea (Horn)



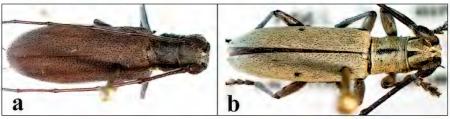


320 (319'). Integument covered with dense pubescence that is formed into long tufts projecting from
anterior of pronotum, middle of apical third of elytron, vertex of head, tibiae, etc. Elytra with
distinct arcuate longitudinal costa somewhat parallel to suture. Antennae shorter than elytra,
consecutive antennomeres shorter beyond third (a, b)





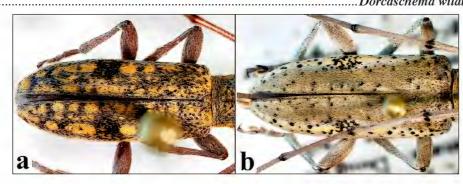
- **322**(321). Coloration mostly uniform without mottled or spotted pubescence323
- 322'. Coloration not uniform; pubescence broken in parts by mottling, glabrous spots, or vittae...324



324(322'). Numerous orange patches of pubescence over elytral surface, mottled with pale or brown	
patches. Elytral surface with only a few glabrous spots, if any (a)	
	y

324'. Dorsal surface of elytra with pale olive green pubescence. Sides of elytra with vittae of white and orange pubescence. Numerous round, glabrous spots present on elytral surface (b).......

Dorcaschema wildii Uhler

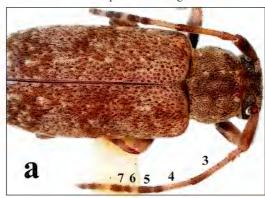


325(321'). Pronotum with sides unarmed; no spines or angled projection at side (although a rounded bulge present in some genera) (a) (note: This character is variable or easy to be misinterpreted in some species of several Acanthocinini genera including *Astylopsis*, *Leptostylopsis*, and *Leptostylus*, among others, that have lobed pronotal sides. They are, therefore, included in both sections of the key)

325'. Pronotum with acute angle to spine at side, usually around middle (b, c)......369

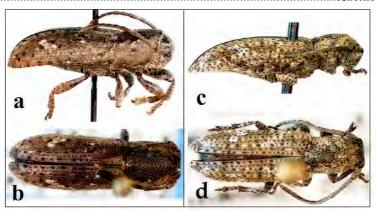


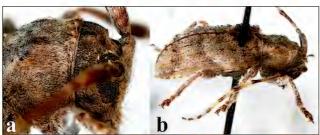




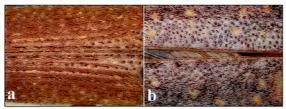
328(327'). Antennae, at most, little more than half length of body. Antennomeres after fourth very short. Eye lobes divided or very widely separated, connected by only a thin line (a)**329**

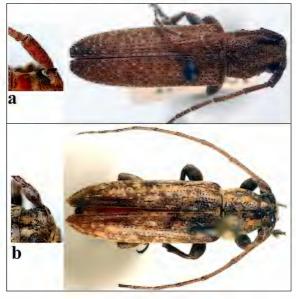






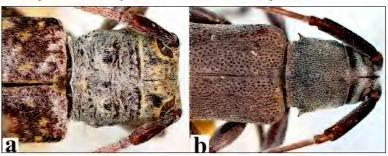
- **331'.** Elytra with punctures not confined in rows and without narrow, longitudinal rows of impunctate regions (b). Pronotum with punctation variable, usually not very conspicuous.....**333**





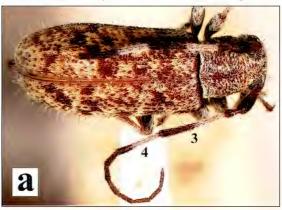
333(331'). Scape short, sometimes clavate, not extending beyond apical one-third of pronotum (a)	
	34

333'. Scape long, slender, only slightly and gradually enlarged apically; extending to about halfway point of pronotum or beyond (b) (Acanthocinini, in part).......**351**

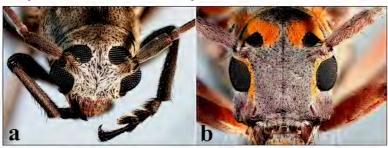




335'. Antennae without annulations as above. Long hairs usually sparse, especially on antennae. Appressed elytral pubescence uniformly colored or with distinct patterns, never mottled.......336

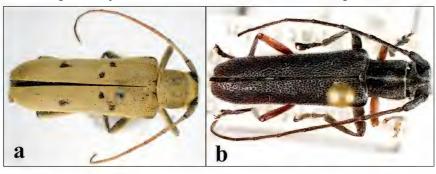


336(335'). Eyes coarse	ely faceted; lower eye lobe occ	upying nearly all of gena, extending to
near genal margin ((a). Elytra with two to three sho	rt, longitudinal white pubescent fasciae,
mostly in a row		Lypsimena fuscata Haldeman
336'. Eves finely face	eted: lower eve lobe smaller and	well removed from genal margin (b).



337(336'). Elytra with uniform pubescence, without patterns or maculae	338
337'. Elytra with pubescence forming maculae or vittae	341
338(337). Integument mostly black. Elytra with very large confluent punctation (a)	
338'. Elytra and appendages differently colored. Punctation not as prominent on elytra	339

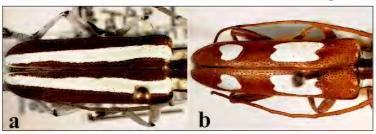




340(339). Antennae with uniform pubescence and without annulae (see <i>339a</i> above)	
	<i>tita</i> Say
340'. Antennae with annulae of pubescence on most antennomeres (a) Saperda inorn	ata Say



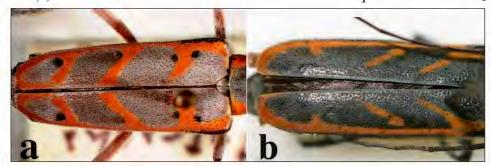
341(337'). Elytra and pronotum with large patches or longitudinal vittae of very dense, bright white pubescence	342
341'. Elytra and pronotum without patches of bright white pubescence	343
342(341). White elytral pubescence in form of broad longitudinal stripe (a)	
342'. White elytral pubescence in form of two (rarely one) large maculae (b)	

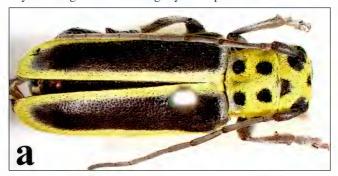


343(341'). Pronotum black or gray at center of disk, with longitudinal bands of orange pubescence on lateral margins	344
343°. Pronotum not black or gray at center of disk, and if longitudinal bands of orange pubescence are present, they are not restricted to lateral margins	346
344(343). Elytra with transverse or oblique bands of orange pubescence	345
344°. Elytra without transverse or oblique bands of orange pubescence (only a longitudinal band around margin and suture) (a)	ius



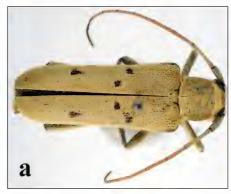
345(344).	Basal-most	orange bar	nd on ely	tra trans	verse at	least for	r half it	s length.	Black spo	ts
often	present aroui	nd base of	ransverse	e and obl	lique bar	nds (a)	5	Saperda tr	identata O	livier





347(346'). Elytra with uniform, dense green or greenish-gray pubescence except for two-three small black, glabrous spots, each. Pronotum without longitudinal pubescent vitta at middle (a)

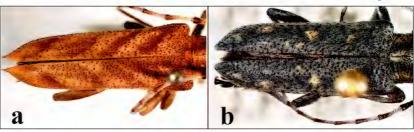
Saperda vestita Say





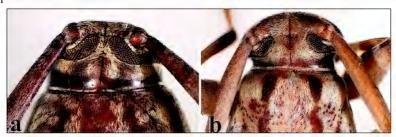


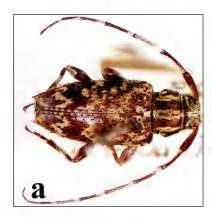
350'. Pubescence pale gray or yellow over black integument. Elytral pubescence contains irregularly sized and placed spots. Elytral apices tapering to a point without a spine (b)......



351 (333 '). Antennae with third and fourth antennomeres very long and broadly annulate at basal
half with pale gray pubescence. Remainder of antennae dark, apical antennomeres much shorter
than third and fourth. Entire body and appendages with long scattered hairs and short mottled
pubescence (see 335a)







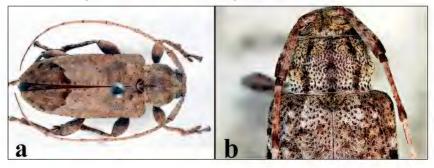
355(354' & 398'). Pronotum with numerous large, evenly scattered punctures over nearly all of surface (sometimes evidenced by regular interruptions in pubescence that otherwise hide punctures). Elytra, especially at basal half, with numerous large punctures (sometimes mostly obscured by pubescence) (a, b) (<i>Astylopsis</i>)	66
355°. Pronotum with few large punctures, limited at most to transverse row near posterior margin, and regions of unevenly scattered punctures elsewhere. Elytra with punctures smaller and less conspicuous (c, d)	1
356(355). Scutellum black, almost always darker than periscutellar region	57
356'. Scutellum gray to reddish brown, similar in color to periscutellar region	8
357(356). Elytra with large mottled gray or white, postmedial transverse macula (a)	y)
357'. Elytra with, at most, few small spots of gray or white maculae (b)	

358(356'). Elytral epipleuron black and strongly contrasting from adjacent integument (a)........



359(358'). Apex of scape and most antennomeres not distinctly darker from remaining areas (mottled but not annulate) (a). Most specimens over 1 cm in length...... Astylopsis perplexa (Haldeman)

359'. Apex of scape and most antennomeres distinctly darker from remaining areas (mottled and



360(359'). Apical one-third of elytra with narrow, angled black or brown transverse macula

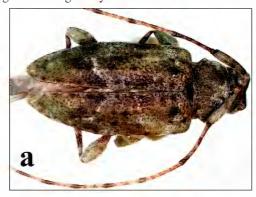
360°. Apical one-third of elytra without distinct black transverse macula (b).....





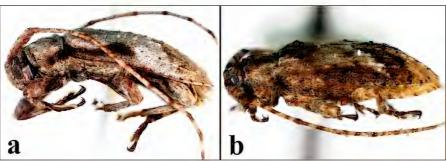
361	(355'). Integument pubescence with distinctive greenish tinge	e. Elytron with angled transverse,
	dark postmedial fascia (maybe subtle) and a raised black, p	subescent callus posterolateral to
	scutellum (a)	Astylidius parvus (LeConte)

361'. Integument without greenish tinge. Elytral calli and white fasciae variable......362

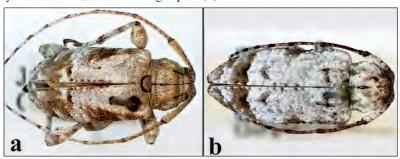


362'. Pronotum without distinct dorsal tubercles (at most 3 vaguely raised areas) (b) (*Styloleptus*)

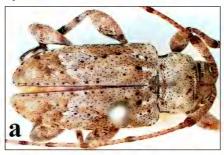




364'. Pronotum with, at most, an incomplete glabrous patch at middle. Integumental pubescence not exactly as described, without orange spots (b).......**365**

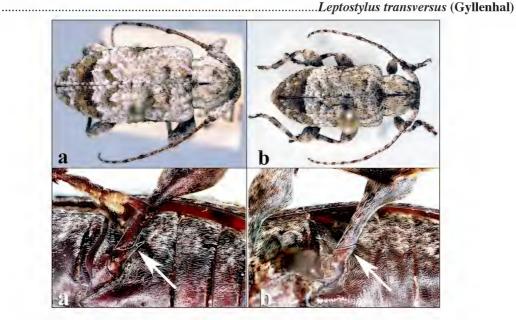


365'. Elytra with, at most, thin postmedial transverse white fascia (a)......**367**



366(365). Metatrochanter long and with apex extending out away from metafemur (a, bottom). Dorsal habitus somewhat variable, commonly as in a, top......*Leptostylus asperatus* (Haldeman)

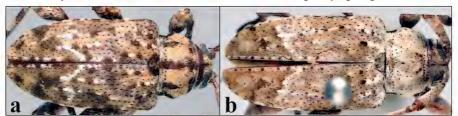
366'. Metatrochanter normal, flush with surface of metafemur (b). Dorsal habitus somewhat variable, commonly as in b, top (note: *Leptostylus hispidulus* Bates would key here, but is excluded since its current presence in the U. S. cannot be confirmed.)......

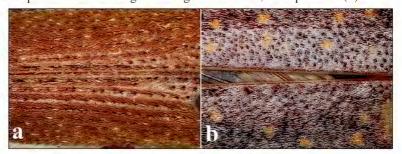


367	(365').	Pubescence mostly uniform brownish or tawny with exception of a partial transverse,
	narrow.	black postmedial fascia on elytron. Pronotum with small patches of denser punctures
	around	central disk (a)

367'. Pubescence not uniform brownish or tawny, interrupted by white postmedial elytral fascia and black spots or small maculae. Pronotal punctures not in above arrangement**368**



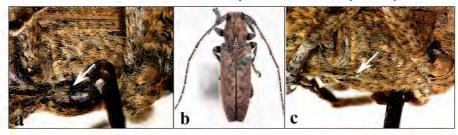


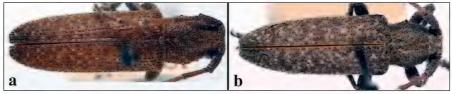


370(36	9). Outer elytral apex with long acute point or spine (a). Pronotum often with conspicuous	
wl	ite pubescence at sides (b) (known only from Florida and the West Indies)	
		el



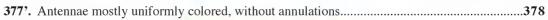
371'. Metasternum with shallow notch that is mostly covered in normally dense pubescence (c)..372







2742 Floren	nicauda Schaeffer
374°. Elytral apex rounded, obliquely truncate, or weakly bidentate (b)	575
375(374'). Elytral apices truncate to weakly bidentate (a). Elytra with a broad transvorantemedially (<i>Pogonocherus</i>) (b) (note: <i>Pogonocherus fasciculatus</i> (DeGeer), verificated at least once in New Jersey, is not included in the key since its establist be confirmed)	which has been shment cannot
375'. Elytral apices rounded (c). Elytra without a broad transverse pale fascia (Eupogonius)	
a b	
376(375). Three to five tufts of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along middle costal ridge of elytrometric and the state of long black hairs along the state of long black hairs	
376'. Elytra lacking tufts of long black hairs (b)	<i>mixtus</i> Haldeman
a b	
377(375'). Antennae with third and fourth antennomeres very long and broadly an	nulate at basal

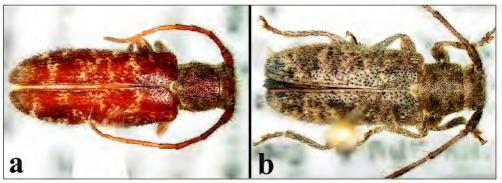


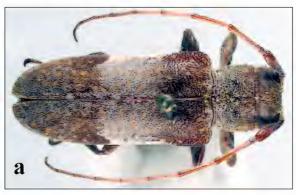


378(377').	Integument	mostly blac	ck to very	dark redo	dish brown.	Pronotum	with a longit	udinal
fascia	of red or ora	nge pubesce	ence on ea	.ch side (a	ı) <i>Eu</i>	pogonius .	subarmatus (LeConte

378'.	Integument	reddish to	gray	or light	brown.	Pronotum	without	red or	orange	pubescent	
fa	sciae										37



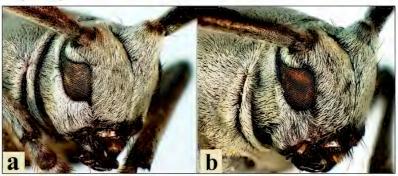




381(380'). Dorsally, integument uniformly light gray or light brown, without mottling or fasciae. Acute spine at posterior fourth of pronotum which is directed posterolaterally (a) (*Dectes*)**382**

381'. Dorsally, integument almost always mottled or patterned with several colors of pubescence (if uniformly pubescent, then very dark brown to black). Spine on pronotum variable......**383**

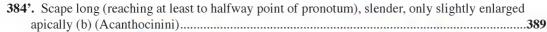


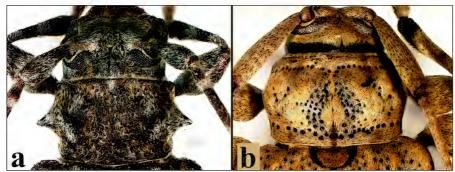


383'. Pronotum usually with only 1 lateral tubercle or projection. Pronotal disk without longitudinal crests. Elytron without prominent middle carina. Size variable......**384**

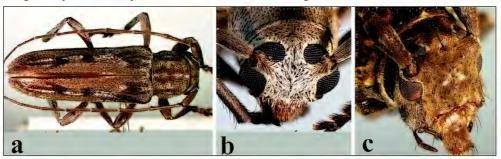


384(383'). Scape short (not reaching halfway point of pronotum), usually distinctly clavate (a) ...385





385'. Scape distinctly clavate and short. Lower eye lobe small and elongate, far removed from genal margin. Elytra without pubescent fasciae in above arrangment (c) (Acanthoderini)......386



386(385'). Elytra with a well defined, undulating, antemedial, transverse white macula (a).......

Aegomorphus quadrigibbus (Say)

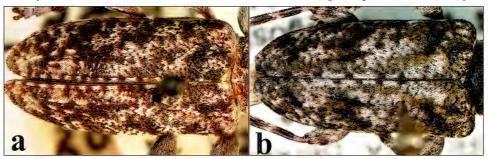


387(386').	Base of	elytra wi	th scattered	granulae	and few	punctures.	A distinct	"N"	or "M"
shape	d black ma	icula pos	t-medially,	otherwise	pubescer	nce mostly	uniformly	gray	(a)
						Ae,	gomorphus	mori	risii (Uhler

387'. Base of elytra without granulae and with many punctures. Black maculae, if present, not very distinct. Pubescence somewhat light and dark mottled over elytron......**388**

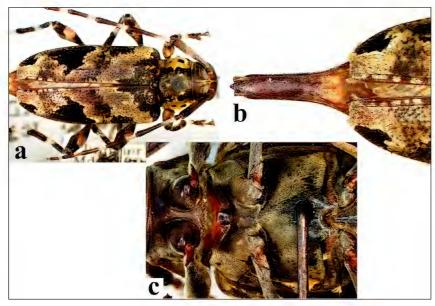


388(387'). Elytral apices rounded. At most, a very ill-defined dark macula postmedially (a)....... *Oplosia nubila* (LeConte)

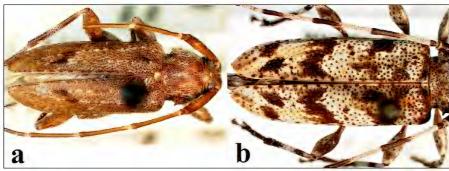




390(389'). If less than 1 cm, then go to 397. If more than 1 cm, then with following characters in
combination: Elytra with dark patch postmedially (a). Mesocoxal process about half as wide
as mesocoxa or less (c). Lateral pronotal tubercles acute. Females with very long protruding
ovipositor (modified final tergite/sternite) (b)



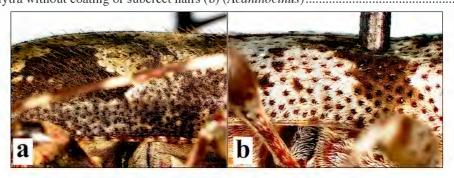
392'. Antennae with distinct pale annulations occupying basal one-third or more of most antennomeres. Elytra with more distinct black maculae and mottling of light and dark pubescence throughout (b). Males with terminal tergite and sternites strongly notched393



393(392'). Elytral epipleural margin and sides of pronotum with bold, black pubescence. Apical half
of elytron with notched, triangular, black macula as in (a). Length over 2 cm

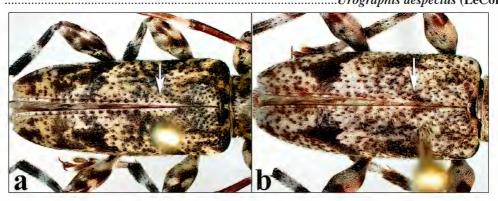
393'. Elytral maculations not as in (a). Most specimens less than 2 cm394



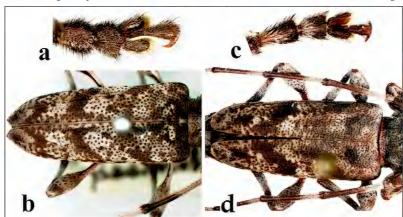


395'. Anterior two-thirds of elytra without large patches or coalesced spots of black. Small region along suture, at anterior one-third, darker than surrounding sutural area. Gray or light reddish brown appearance with less mottling at anterior two-thirds of elytra as in previous species (b)...

**Urographis despectus* (LeConte)*



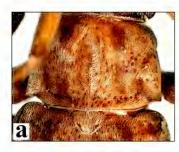
396(394').	Tarsi	uniformly	dark	(a).	Males	with	dense	fringe	of s	short	white	pubesceno	ce on
inner-v	entral	margin of b	oasal	anten	nomere	s (b).			.Ace	antho	cinus	obsoletus	(Olivier



397(390'). Upper eye lobes very close together, separated less than the width of the upper eye lobe (see *353a*). Pronotum with yellow-brown pubescent bands or elongated spots, remainder darker. Elytra with postmedial and antemedial dark maculae bordered by lighter maculae (a)...

**Nyssodrysina haldemani* (LeConte)

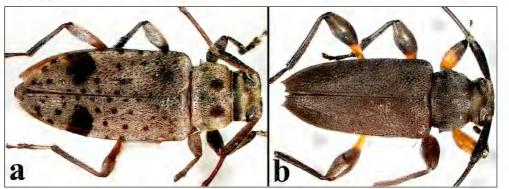




399(398'). Pronotal spines acute, posteriorly positioned at sides, usually with abrupt constriction of pronotum posterior to spine (a) (see also 325c)......400

399'. Pronotal tubercles rounded or without abrupt constriction at posterior base of tubercle, usually closer to middle of sides (see 355a-d).......355

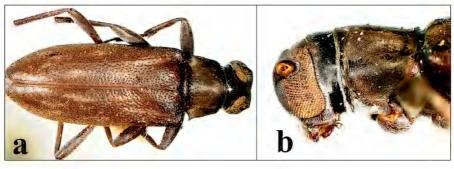
400(399). Antennae with bold, white pubescent annulae restricted to extreme base of each joint.
Uniformly reddish brown except for subtle, angled black macula postmedially and subtle white
highlights along portions of costae of elytra. Females with long, protruding ovipositor formed by
terminal tergite and sternite (see 392a)



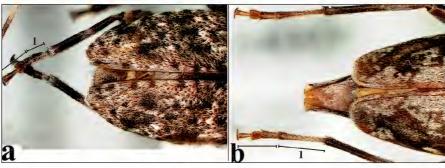


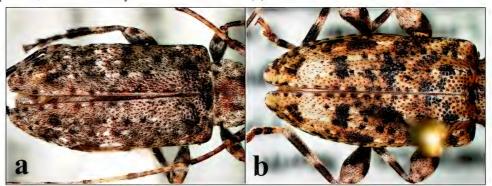
403(402).	Epipleuron of elytron dark (a)	Hyperplatys maculata Haldeman
403 °. Epi	ipleuron of elytron without dark markings (b)	Hyperplatys aspersa (Say)



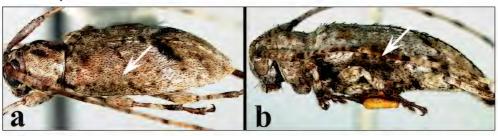


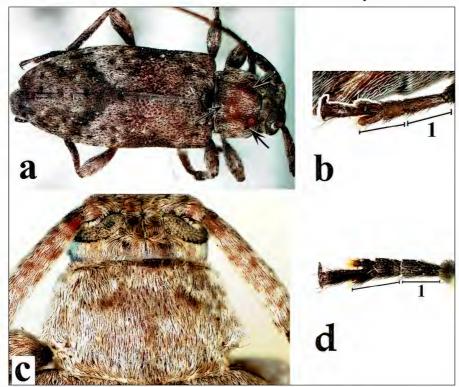
406(**405**'). Part or all of elytral suture with alternating light and dark maculations. First metatarsomere shorter than remaining metatarsomeres combined (a) (*Liopinus & Sternidius*)......**407**



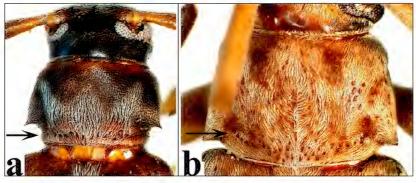




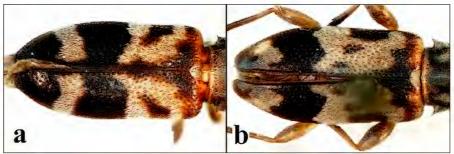




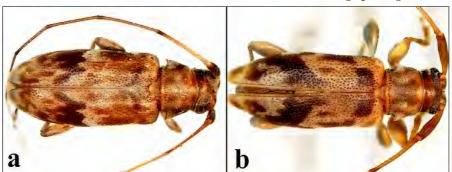
411'. Pronotum with row of large punctures posteriorly that terminate before reaching lateral spine and meet scattered large punctures on inside of spine base. Large punctures also present around base of lateral spine and elsewhere on pronotal disk (b) (*Lepturges*)......**415**

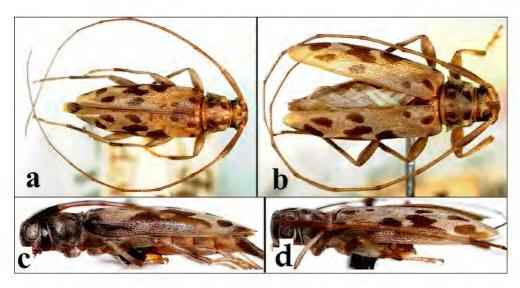


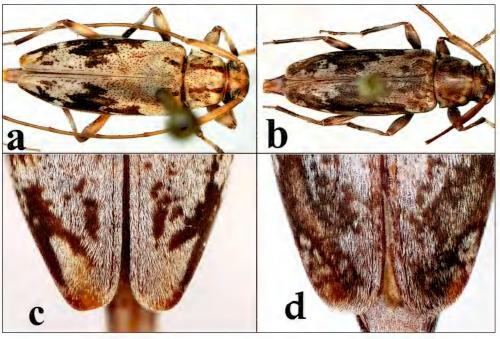
412(411). Ely	a with apices behind transverse pale macula completely dark. Apical pale	
macula bo	ered anteriorly by a broader transverse dark macula that reaches suture. Base of	
elytra arou	d scutellum completely dark, not interrupted by paler fasciae (a)	
		7)
		_



- **414'.** Anterior border of postmedial dark macula oblique, with, at most, subtle indentations (b)... *Urgleptes signatus* (LeConte)







Species Accounts and Notes

All the species treated in the key are listed below in alphabetical order within each subfamily. The current tribal placement and a brief overview of distribution, seasonality, and host plants are listed for each species. The sizes listed are based on the photographed specimens (or average if more than one specimen was photographed). Appendix 1 is a list of scientific and common names of host plants alphabetically arranged by scientific name. Plant names are presented in their currently accepted scientific combination with standardized botanical author abbreviations. These names are based on the USDA Natural Resources Conservation Service website (USDA NRCS 2007). Host plant information is not exhaustive and primary literature should be examined for more details. Biological information is summarized mostly from Yanega (1996) and Chemsak & Linsley (1997), but also Thomas, et al. (2006); Linsley (1962a, 1962b, 1963, 1964); Linsley & Chemsak (1972, 1976, 1984, 1995); Bond & Philips (1999); Browne, et al. (1993); Brown & Peck (1996); Giesbert (1993); Korotyaev, et al. (2005); Lingafelter & Chemsak (2002); Lingafelter & Nearns (2006); Lingafelter & Hoebeke (2002); Lingafelter & Horner (1993); Lingafelter & Ivie (2005); Hanley (2005); Hoffman (2003); MacRae (1993); MacRae & Rice (2007); Monné (2001a-c; 2002a-b; 2005a-b; 2006); Monné & Hovore (2005); Morris (2002); Rice & Veal (2006); Hoffman, et al. (2002); Vogt (1949) and Schiefer (1998, 2000, 2001). To this list are important unpublished observations from Roy Morris and Eugenio Nearns, along with my own observations from fieldwork and examination of label data from the Smithsonian Institution collections. Plates of nearly every species follow, arranged based on their similarity by key characters. Using these plates as the primary identification tool is not encouraged as errors can result since key characters may not be visible. The plates are provided as a final confirmation reference to be consulted after the beetle is keyed.

DISTENIINAE

Distenia undata (Fabricius) (Disteniini, Plate 1a, 24 mm). Known throughout the eastern and central U. S., adults are active from June–September. Larvae feed in various hardwoods including *Cercis canadensis* and also *Pinus*.

PARANDRINAE

- *Hesperandra polita* (Say) (Parandrini, Plate 1c, 15 mm). Until recently, this species was placed in the genus *Parandra*. Present throughout eastern U. S., adults are active from June–August. Larvae feed in decaying heartwood of *Carya*, *Liriodendron*, and *Fagus*.
- *Neandra brunnea* (Fabricius) (Parandrini, Plate 1d, 14 mm). Until recently, this species was placed in the genus *Parandra*. Known throughout eastern North America, adults are active from March–November. Larvae feed in moist, decaying heartwood of most eastern trees.

PRIONINAE

- Archodontes melanopus (Linnaeus) (Macrotomini, Plate 2j, 45 mm). Known throughout the southeastern U. S. north into Virginia and west to Texas, adults are active from June–September. Larvae develop and form galls in roots of *Quercus virginiana* and *Q. nigra*, primarily, and other hardwoods.
- **Derobrachus brevicollis Audinet–Serville** (Prionini, Plate 2a, female on left, male on right, 36 mm). Known from the southeastern U. S. to Texas, adults are active from June–August. Larvae develop in the soil on roots of *Paspalum notatum* and *Quercus*.
- *Elateropsis rugosus* Gahan (Solenopterini, Plate 1f, two variants shown, 24 mm). Known from southern Florida, Bahamas, and possibly Cuba, adults have been collected from June–August. Larvae are known to develop in *Metopium toxiferum*, *Bursera simaruba*, and *Coccaloba diversifolia*.

- *Elateropsis scabrosus* Gahan (Solenopterini, Plate 1g, 23 mm). Occurring in southern Florida, Bahamas, and Cuba, adults have been collected from May–August. Larvae are known to develop in *Bursera simaruba*.
- Mallodon dasystomus (Say) (Macrotomini, Plate 2k, 40 mm). Known primarily from the southern U. S. and into Mexico, but with records from Illinois and Virginia, adults are active from May–October. Larvae feed in many hardwoods including Acer negundo, A. pseudoplatanus, Alnus, Bursera simaruba, Quercus, Liquidambar styraciflua, Carya, Platanus occidentalis, P. wrightii, Salix, Sideroxylon lanuginosa, Celtis laevigata, and Ulmus crassifolia.
- Orthosoma brunneum (Forster) (Prionini, Plate 2b, 32 mm). Adults of this widespread eastern U. S. species are active from May–November. Larvae feed in hardwoods and conifers including Juglans, Carya, Castanea, Quercus, Tilia, Acer, Abies, Pinus, and Tsuga.
- *Prionus debilis* Casey (Prionini, Plate 2e, 22 mm). Adults of this central U. S. species are active from May–November. Larvae may feed in living roots of primarily *Quercus* and *Castanea*, but also *Vitis*, *Pyrus*, and *Zea mays*.
- **Priorus fissicornis Haldeman** (Prionini, Plate 2c, 25 mm). Adults of this central U. S. species are active from May–July. Larvae are root feeders on grasses (also reported from roots of *Zea mays*).
- *Prionus imbricornis* (Linnaeus) (Prionini, Plate 2f, 25 mm). Adults of this widespread eastern and central U. S. species are active from April–November. Larvae feed on living roots, primarily *Quercus*, *Castanea*, *Carya illinoinensis*, *Vitis*, *Pyrus*, and *Zea mays*.
- **Prionus laticollis** (**Drury**) (Prionini, Plate 2g, 26 mm). Adults of this widespread eastern and central North American species are active from June–September. Larvae feed on living roots of various trees and shrubs, including *Quercus*, *Populus*, *Tilia*, *Castanea*, *Malus*, and *Carya illinoinensis*, among others.
- *Prionus palparis* Say (Prionini, Plate 2d, 29 mm). Adults of this central and west–central U. S. species are active from April–August. Larval hosts are unknown.
- *Prionus pocularis* Dalman (Prionini, Plate 2h, 26 mm). Adults of this widespread eastern U. S. species are active from May–August. Larvae feed in dead *Pinus*.
- *Sphenostethus taslei* (**Buquet**) (Solenopterini, Plate 1e, two variants shown, 21 mm). Known from central and eastern U. S., but much more common in Florida, adults are active from June–August. Larvae feed in *Quercus*, *Castanea*, *Fagus*, and *Cercis*.
- Stenodontes chevrolati Gahan (Macrotomini, Plate 2i, 55 mm). Known from southern Florida, Bahamas, and Cuba, adults have been collected in many months of the year, especially from May–September. Larvae develop in many tropical hardwoods including *Spondias purpurea* and *Bursera simaruba*.
- Strongylaspis corticarius (Erichson) (Macrotomini, Plate 1i, 30 mm). Known from Mexico, Central America, and northern South America, with one unconfirmed record in Florida, adults have been collected from May–August. Hosts are not reported for this species although other Neotropical species are known from *Scalesia*.
- *Tragosoma depsarius* (Linnaeus) (Prionini, Plate 1h, male on left and female on right, 24 mm). Adults of this widespread U. S. species are active from June–September. Larvae feed in sapwood of decaying *Pinus*.

SPONDYLIDINAE

Scaphinus muticus (Fabricius) (Spondylidini, Plate 1b, 14 mm). Adults of this uncommon southeastern and east-central U. S. species are active from June–July. Larvae develop in *Pinus*.

ASEMINAE

Arhopalus foveicollis (Haldeman) (Asemini, Plate 3d, 20 mm). Adults of this widespread eastern and central North American species are active from June–August. Larvae feed in dead *Pinus* and *Picea*.

- *Arhopalus rusticus* (Linnaeus) (Asemini, Plate 3e, 19 mm). Adults of this southeastern U. S. and Bahamian species are active from March–August. Larvae develop in *Pinus*.
- **Asemum australe LeConte** (Asemini, no figure). Adults of this rare eastern U. S. species are active from June–July. The only known larval host is *Pinus strobus*.
- **Asemum striatum** (Linnaeus) (Asemini, Plate 3c, 13 mm). Adults of this widespread North American species are active from April–July. Larvae feed in recently dead conifers, especially *Pinus*, but also *Picea*, *Larix*, *Abies*, and *Pseudotsuga menziesii*.
- Atimia confusa (Say) (Atimiini, Plate 18e, 9 mm). Adults of this widespread central and eastern U. S. species are active in spring and fall. Larvae feed under bark of *Juniperus*, *Cupressus*, *Taxodium distichum*, *Thuja*, and *Chamaecyparis*.
- **Tetropium cinnamopterum Kirby** (Asemini, Plate 3a, 12 mm). Adults of this southeast Canada and northeast U. S. species are active from May–July. Larvae develop beneath bark of *Larix*, *Abies*, *Pinus*, and *Picea*.
- **Tetropium schwarzianum** Casey (Asemini, Plate 3b, 12 mm). Adults of this southeast Canada and northeast U. S. species are active from June–July. Larvae feed in *Picea* and *Pinus strobus*.

LEPTURINAE

- **Acmaeops discoideus (Haldeman)** (Lepturini, Plate 7b, 8 mm). Adults of this eastern U. S. species are active from May–June. Larvae develop in *Pinus*.
- Acmaeops proteus (Kirby) (Lepturini, Plate 5i, two variants shown, 7 mm). Adults of this wide-spread eastern and northern North American species are active from May–August. Larvae feed on the inner bark of conifers, especially *Abies*, *Picea*, *Tsuga*, and *Pinus*.
- **Alosternida chalybaea** (Haldeman) (Lepturini, Plate 7e, 6 mm). Adults of this eastern U. S. species are active from May–July, occasionally attracted to flowers of *Cornus*. Larval hosts are unknown.
- Analeptura lineola (Say) (Lepturini, Plate 7j, 10 mm). Adults of this common eastern U. S. species are active from May–August. Larvae feed in various hardwoods, especially *Betula*, *Carpinus caroliniana*, *Ostrya virginiana*, and also *Pinus*. Adults are attracted to many flowers, especially *Aruncus dioicus*, *Hydrangea arborescens*, *Vitis* and *Smilacina racemosa*.
- Anoplodera pubera (Say) (Lepturini, Plate 7i, 10 mm). Adults of this widespread eastern U. S. species are active from May–July. Larvae feed in *Ulmus*, *Juglans*, and *Pinus*. Adults are attracted to many wildflowers, especially *Viburnum*, *Spiraea*, *Heracleum*, *Achillea*, *Tragopogon* and *Hydrangea arborescens*.
- Anthophylax attenuatus (Haldeman) (Lepturini, Plate 4g, 14 mm). Adults of this southeastern Canada and northeastern U. S. species are active from May–August. Larvae feed in decaying hardwood logs, including Acer, Fagus, Ostrya virginiana, and Populus.
- Anthophylax cyaneus (Haldeman) (Lepturini, Plate 3f, two variants shown, 13 mm). Adults of this eastern North American species are active from May–July. Larvae feed in various hardwoods including Acer, Betula, Amelanchier arborea, Castanea, Fagus, and may also feed in conifers. Adults are often collected on Acer spicatum.
- Anthophylax hoffmani Beutenmüller (Lepturini, no figure). Adults of this localized U. S. species are rarely collected. They are known from high elevations in Virginia, above 3,500 feet from June–July. Abies fraseri is a probable larval host, Picea rubens may also be a host.
- Anthophylax viridis LeConte (Lepturini, Plate 3g, two variants shown, 14 mm). Adults of this eastern U. S. species are active from June–July. Larvae develop in *Betula*, *Fagus*, and *Acer*.
- Bellamira scalaris (Say) (Lepturini, Plate 8c, two variants shown, 22 mm). Adults of this eastern U. S. species are active from May–September. Larvae feed in decayed hardwoods including *Populus*, Carya, Salix, Acer, Liriodendron, and Betula, as well as Pinus. Adults have been collected on flowers of Rhus, Spiraea, Cirsium, and Phytolacca.
- *Brachyleptura champlaini* Casey (Lepturini, Plate 5g, 10 mm). Adults of this eastern U. S. species are active from June–August. Larvae feed in *Pinus*. Adults are attracted to many wildflowers, especially *Tragopogon*, *Hydrangea arborescens*, and *Achillea*.

- **Brachyleptura circumdata** (Olivier) (Lepturini, Plate 5f, 8 mm). Adults of this eastern U. S. species are active from June–July. Larvae feed in *Picea* and possibly *Pinus*.
- Brachyleptura rubrica (Say) (Lepturini, Plate 5e, 13 mm). Adults of this eastern North American species are active from May–August. Larvae feed in decayed hardwoods including Carya, Fagus, Quercus, Prunus, and Platanus. Adults are attracted to many flowers, especially Tragopogon, Sambucus nigra canadensis, Hydrangea arborescens, and Achillea.
- **Brachyleptura vagans** (Olivier) (Lepturini, Plate 5h, two variants shown, 10 mm). Adults of this eastern North American species are active from April–August. Larvae feed in decaying conifers and hardwoods including *Betula*, *Carya*, *Juglans nigra*, *Pinus*, and *Tsuga*. Adults are attracted to many flowers, especially *Rhus glabra* and *Daucus carota*.
- **Brachysomida bivittata** (Say) (Lepturini, Plate 4d, two variants shown, 9 mm). Adults of this eastern North American species are active from April–July. Larvae develop in *Quercus*, *Carya*, and *Cornus*. Adults are attracted to many wildflowers, in particular *Geranium maculatum*.
- Centrodera decolorata (Harris) (Lepturini, Plate 3j, 21 mm). Adults of this eastern North American species are active from May–July. Larvae develop in many trees such as Acer, Castanea, Fagus, and Quercus, among others.
- *Centrodera quadrimaculata* (Champlain & Knull) (Lepturini, no figure). Adults of this rare northeastern U. S. species are active from May–June. Larval feeding habits are unknown.
- *Centrodera sublineata* LeConte (Lepturini, Plate 4f, 12 mm). Adults of this central and eastern U. S. species are active from March–May. No larval hosts are known.
- *Charisalia americana* (Haldeman) (Lepturini, Plate 8h, 8 mm). Adults of this uncommon eastern U. S. species are active from April–July. Larvae feed in decayed *Liriodendron* and *Nyssa*. Adults are attracted to wildflowers, notably *Heracleum*.
- **Desmocerus palliatus** (Forster) (Desmocerini, Plate 6b, 22 mm). Adults of this widespread, but generally uncommon eastern and central U. S. species are active from April–August. Larvae feed in the living roots of *Sambucus nigra canadensis*, and adults are often found on the blossoms.
- *Encyclops caerulea* (Say) (Lepturini, Plate 3h, 8 mm). Adults of this eastern North American species are active from May–July. Larvae feed in living hardwoods, especially *Quercus* and *Acer*, and rarely *Pinus*.
- *Evodinus monticola* (Randall) (Lepturini, Plate 4h, two variants shown, 8 mm). Adults of this eastern and northern North American species are active from May–July. Larvae develop in various conifers.
- *Gaurotes cyanipennis* (Say) (Lepturini, Plate 3i, 10 mm). Adults of this eastern North American species are active from May–August. Larvae develop in numerous hardwoods and shrubs. Adults are attracted to *Hydrangea arborescens*, among other wildflowers.
- *Gaurotes thoracica* (Haldeman) (Lepturini, Plate 4c, 8 mm). Adults of this typically uncommon (but occasionally, locally abundant) eastern U. S. species are active from April–July. Larval feeding habits are unknown.
- *Grammoptera haematites* (Newman) (Lepturini, Plate 8d, 5 mm). Adults of this eastern North American species are active from April–July. Larvae feed in various shrubs. Adults are attracted to flowers of genera *Viburnum*, *Cornus*, *Tragopogon* and *Hydrangea arborescens*, among others.
- Grammoptera subargentata (Kirby) (Lepturini, Plate 8e, 7 mm). Adults of this boreal North American species are active from April—August. Larvae feed in various hardwoods, including *Rhus*, *Populus*, and *Quercus*. Adults are attracted to many wildflowers, especially *Heracleum*, *Tragopogon* and *Hydrangea arborescens*.
- *Idiopidonia pedalis* (LeConte) (Lepturini, Plate 7h, 8 mm). Adults of this uncommon eastern North American species are active from June–July. Larval feeding habits are unknown. Adults have been collected on flowers of *Rhododendron*, *Viburnum*, *Spiraea*, and *Pyrus*.
- Judolia cordifera (Olivier) (Lepturini, Plate 9h, 10 mm). Adults of this eastern U. S. species are active from May–August. Larvae feed in *Castanea*. Adults are attracted to many wildflowers, especially *Tragopogon* and *Hydrangea arborescens*, *Daucus carota*, *Achillea*, and also *Sambucus nigra* and *Cornus*.

- *Judolia montivigans* (Couper) (Lepturini, Plate 6k, two variants shown, 9 mm). Adults of this boreal North American species are active from June–August. Larvae feed in conifers as well as *Salix* and *Populus*.
- *Leptorhabdium pictum* (Haldeman) (Lepturini, Plate 5c, 13 mm). Adults of this eastern U. S. species are active from May–June. Larvae feed in various hardwoods including *Betula*, *Quercus*, and *Carya*.
- *Leptura abdominalis* (Haldeman) (Lepturini, Plate 7k, 12 mm). Adults of this eastern U. S. species are active from May–July. Larvae feed in *Taxodium distichum* and *Juniperus*.
- *Leptura obliterata deleta* (LeConte) (Lepturini, Plate 8b, 14 mm). Adults of this rare northeastern U. S. subspecies are active from July–August. Larval hosts are unknown. Adults have been taken on flowers of *Spiraea* and *Solidago*.
- Leptura subhamata Randall (Lepturini, Plate 8a, two variants shown, 13 mm). Adults of this southeastern Canada and northeastern U. S. species are active from June–August. Larvae feed in decaying *Tsuga* and *Pinus*. Adults are attracted to many wildflowers, especially *Spiraea*, *Achillea*, *Ceanothus*, *Heracleum*, *Tragopogon*, and *Hydrangea arborescens*.
- *Lepturopsis biforis* (Newman) (Lepturini, Plate 7f, 14 mm). Adults of this eastern North American species are active from June–August. Larvae feed in many decaying hardwoods and conifers.
- *Lycochoriolaus lateralis* (Olivier) (Lepturini, Plate 8i, two variants shown, 11 mm). Adults of this uncommon southeastern U. S. species are active from March–June. Larval feeding habits are unknown.
- *Metacmaeops vittata* (Swederus) (Lepturini, Plate 6a, 8 mm). Adults of this very common eastern North American species are active from May–July. Larvae feed in *Liriodendron* and *Castanea*. Adults have been collected on many flowers, in particular those of *Vitis*.
- **Necydalis mellita** (Say) (Necydalini, Plate 10g, 13 mm). Adults of this central and eastern U. S. species are active from May–August. Larvae develop in the heartwood of *Quercus* and *Castanea*, primarily, but also rarely in *Pinus*.
- **Neoalosterna capitata** (Newman) (Lepturini, Plate 6d, 8 mm). Adults of this eastern U. S. species are active from May–July. Larvae develop in *Betula*. Adults have been collected on flowers of *Cornus*, *Aruncus*, *Viburnum*, *Pyrus*, and *Hydrangea*.
- *Pidonia aurata* (Horn) (Lepturini, Plate 6i, 10 mm). Adults of this eastern U. S. species are active from May–July. Larval feeding habits are unknown. Adults are attracted to many wildflowers, especially *Viburnum*, *Spiraea*, *Tragopogon* and *Hydrangea arborescens* and flowering trees such as *Rhododendron* and *Cornus*.
- *Pidonia densicollis* (Casey) (Lepturini, Plate 6j, 9 mm). Adults of this eastern U. S. species are active from May–June. Larval feeding habits are unknown. Adults are attracted to flowers of *Rhododendron*, *Hydrangea*, *Cornus*, *Geranium*, *Aruncus*, and *Rubus*.
- *Pidonia ruficollis* (Say) (Lepturini, Plate 7c, 9 mm). Adults of this eastern U. S. species are active from May–July. Larvae feed in numerous hardwoods. Adults are attracted to flowers of many genera including *Viburnum*, *Spiraea*, *Geranium*, *Tragopogon*, *Hydrangea arborescens* and flowering trees and shrubs such as *Rhododendron*, *Ceanothus*, and *Cornus*.
- **Pseudogaurotina abdominalis** (**Bland**) (Lepturini, Plate 6c, 8 mm). Adults of this northeastern U. S. species are active from May–July. Larval feeding habits are unknown.
- **Pseudostrangalia cruentata** (Haldeman) (Lepturini, Plate 9a, 7 mm). Adults of this uncommon central and eastern U. S. species are active from May–June. Larval feeding habits are unknown. Adults are attracted to flowers of *Cornus*, *Aesculus*, *Rosa*, *Heracleum*, and *Tragopogon*.
- **Rhagium inquisitor** (Linnaeus) (Lepturini, Plate 4e, 12 mm). Adults of this widespread North American species are active from February–July. Larvae bore in various conifers including *Abies, Larix, Picea, Pinus, Pseudotsuga*, and *Tsuga*.
- Stenelytrana emarginata (Fabricius) (Lepturini, Plate 8j, 31 mm). This widespread eastern and central U. S. species was, until recently, known under the genus *Leptura*. Adults are active from April–August. Larvae develop in decaying hardwoods including *Fagus*, *Ulmus*, *Liriodendron*, *Nyssa*, *Castanea*, and *Acer*. Adults have been collected on *Rosa* flowers and sapflows.
- Stenocorus cinnamopterus (Randall) (Lepturini, Plate 4b, 12 mm). Adults of this central and eastern U. S. species are active from April–July. Larvae feed in *Hydrangea* and *Prunus serotina*.

- Stenocorus cylindricollis (Say) (Lepturini, Plate 3k, 15 mm). Adults of this eastern U. S. species are active from May–August. Larvae feed in *Carya* and *Rhus*.
- Stenocorus schaumii (LeConte) (Lepturini, Plate 4a, 23 mm). Adults of this central and eastern U. S. species are active from May–August. Larvae feed in various hardwoods including Fraxinus, Acer, Fagus, Juglans nigra, and Amelanchier.
- Stenocorus trivittatus (Say) (Lepturini, Plate 5b, 15 mm). Adults of this uncommon North American species are active from June–July. Larval feeding habits are unknown.
- *Stenocorus vittiger* (Randall) (Lepturini, Plate 5a, 14 mm). Adults of this southern Canada and eastern U. S. species are active from May–July. Larvae develop in various shrubs.
- Stictoleptura canadensis (Olivier) (Lepturini, Plate 5d, 14 mm). Adults of this southern Canada and eastern U. S. species are active from May–August. Larvae feed in various conifers and occasionally hardwoods. Adults are attracted to flowers of many genera including Melilotus, Cirsium, Spiraea, Achillea, Solidago, Eupatorium, Rosa, Rhus, and Daucus.
- Strangalepta abbreviata (Germar) (Lepturini, Plate 7d, 13 mm). Adults of this widespread eastern North American species are active from May–August. Larvae feed in various decaying conifers and hardwoods including Abies, Pinus, Picea, Juniperus, Tsuga, Larix, Populus, Acer, Betula, and Castanea. Adults are attracted to many wildflowers, especially Heracleum, Tragopogon, Daucus carota, Vitis, and Hydrangea arborescens.
- Strangalia acuminata (Olivier) (Lepturini, Plate 10a, 12 mm). Adults of this common eastern North American species are active from June–July. Larvae develop in various shrubs and hardwoods including Alnus and Ostrya virginiana. Adults are attracted to many wildflowers, especially of genera Rhus, Viburnum, Prunus, Tragopogon, Sambucus nigra canadensis, and Hydrangea arborescens.
- Strangalia bicolor (Swederus) (Lepturini, Plate 8g, 12 mm). Adults of this eastern U. S. species are active from May–July. Larvae feed in *Acer* and *Quercus*. Adults are attracted to *Hydrangea* arborescens, among other wildflowers.
- Strangalia famelica famelica Newman (Lepturini, Plate 9f, 13 mm). Adults of this widespread eastern U. S. species are active from May–July. Larvae develop in Castanea, Quercus, and Betula.
- Strangalia famelica solitaria Haldeman (Lepturini, Plate 9g, 12 mm). Adults of this southeastern U. S. species are active from May–September. Larvae develop in *Quercus* and *Betula*.
- Strangalia luteicornis (Fabricius) (Lepturini, Plate 9e, 12 mm). Adults of this common central and eastern U. S. species are active from May–August. Larvae feed in various hardwoods and shrubs including *Ulmus*, *Quercus*, and *Vitis*. Adults are attracted to many wildflowers, especially *Tragopogon*, *Daucus carota*, and *Hydrangea arborescens*, and occasionally on *Asclepias syriaca*.
- *Strangalia sexnotata* **Haldeman** (Lepturini, Plate 10b, 12 mm). Adults of this common central and eastern U. S. species are active from May–August. Larval feeding habits are unknown.
- Strangalia strigosa Newman (Lepturini, Plate 9d, 13 mm). Adults of this Florida species are active from March–June. Larval hosts are unknown for this species. Adults have been collected on many flowers including Rhamnus, Castanea, Persea, Erigeron, Rubus, Ilex, Vaccinium, Oxypolis, and Asparagus.
- Strophiona nitens (Forster) (Lepturini, Plate 6e, 11 mm). Adults of this widespread eastern and central U. S. species are active from May–July. Larvae feed in living or dead hardwoods, including Castanea, Quercus, Carya, Juglans, Fagus, and Acer. Adults are attracted to Hydrangea, Spiraea, Tragopogon, and other flowers.
- *Trachysida mutabilis* (Newman) (Lepturini, Plate 7g, two variants shown, 12 mm). Adults of this eastern North American species are active from April–July. Larvae feed in decaying hardwoods. Adults are attracted to flowers of *Cornus*, *Viburnum*, *Crataegus*, *Achillea*, *Spiraea*, *Aruncus*, *Ceanothus*, and *Prunus*, among many others.
- *Trigonarthris atrata* (LeConte) (Lepturini, Plate 6g, 15 mm). Adults of this uncommon central and eastern U. S. species are active from May–June. Larvae may feed in *Ulmus*. Adults have been collected on flowers of *Cornus* and *Castanea*.
- *Trigonarthris minnesotana* (Casey) (Lepturini, Plate 6f, 14 mm). Adults of this eastern U. S. species are active from May–August. Larvae feed in various hardwoods and *Pinus*. Adults have

- been collected on many flowers including Cirsium, Hydrangea, Achillea, Viburnum, Spiraea, Cornus, and others.
- *Trigonarthris proxima* (Say) (Lepturini, Plate 6h, 14 mm). Adults of this eastern North American species are active from May–August. Larvae feed in various decaying hardwoods, including *Acer* and *Carya*. Adults are attracted to wildflowers, especially *Viburnum*, *Sambucus nigra canadensis*, *Achillea*, *Cornus*, *Tragopogon*, and *Hydrangea arborescens*.
- *Typocerus acuticauda* Casey (Lepturini, Plate 10d, 12 mm). Adults of this common eastern North American species are active from May–September. Larval feeding habits are unknown. Adults have been taken on *Spiraea*, among other flowers.
- *Typocerus badius* (Newman (Lepturini, Plate 9l, 13 mm). This species is known only from Georgia and Florida. Larval hosts are unknown. Adults are active from April–June and can be collected on flowers of *Ilex*, *Vaccinium arboreum*, and *Cornus asperifolia*. Larval hosts include *Quercus*.
- *Typocerus deceptus* Knull (Lepturini, Plate 10c, 12 mm). Adults of this eastern U. S. species are active from June–July. Larval feeding habits are unknown. Adults are attracted to *Hydrangea arborescens* and *Rhus glabra*.
- *Typocerus fulvocinctus* **Knull** (Lepturini, Plate 9c, 9 mm). Known only from Florida, adults are active from April to June and have been collected on flowers of *Ilex glabra*. Larval hosts are unknown.
- *Typocerus lugubris* (Say) (Lepturini, Plate 8f, 10 mm). Adults of this eastern U. S. species are active from May–August. Larvae feed in *Pinus*. Adults are attracted to many wildflowers, especially *Tragopogon*, *Rhus glabra*, and *Hydrangea arborescens*.
- *Typocerus lunulatus* (Swederus) (Lepturini, Plate 9b, 9 mm). Adults of this east-central and southeast U. S. species are active from May–August. Larvae develop in *Pinus*.
- Typocerus octonotatus (Haldeman) (Lepturini, Plate 9j, 11 mm). Adults of this common central and eastern U. S. species are active from May–August. Larvae are borers in native grasses including Andropogon, Sporobolus, Sorghastrum, and Agropyron. Adults are on many flowers including Rudbeckia, Aster, Cirsium, Rhus, Helianthus, Solidago, and Coreopsis.
- *Typocerus sinuatus* (Newman) (Lepturini, Plate 9i, 12 mm). Adults of this southern U. S. species are active from March–August and can be collected on flowers of *Asclepias*, *Ceanothus americanus*, *Oenothera*, *Marshallia*, and *Opuntia*. Larval feeding habits are unknown.
- *Typocerus velutinus* (Olivier) (Lepturini, Plate 10e, 12 mm). Adults of this eastern North American species are active from May–September. Larvae feed in various decaying hardwoods including *Betula, Populus, Quercus*, and *Carya*. Adults are attracted to many wildflowers, especially *Spiraea, Viburnum, Sambucus nigra, Asclepias, Achillea, Daucus carota*, and *Hydrangea arborescens*.
- *Typocerus zebra* (Olivier) (Lepturini, Plate 9k, 12 mm). Adults of this east central and southern U. S. species are active from March–August. Larvae feed in *Pinus*. Adults have been taken on many flowers including *Cornus*, *Rudbeckia*, *Viburnum*, *Cirsium*, *Rhus*, *Hydrangea*, *Solidago*, and *Achillea*.
- Xestoleptura octonotata (Say) (Lepturini, Plate 7a, 12 mm). Adults of this eastern North American species are active from May–July. Larvae feed in *Quercus*. Adults are attracted to many wild-flowers and flowering trees, especially *Cornus*, *Ceanothus*, *Viburnum*, *Tragopogon*, and *Hydrangea arborescens*.

CERAMBYCINAE

- Achryson surinamum (Linnaeus) (Achrysonini, Plate 21k, 18 mm). Adults are active from April—September. Larvae feed in numerous woody plants, mostly legumes, including Acacia, Prosopis, Robinia, but also Ulmus and Celtis.
- Aethecerinus hornii (Lacordaire) (Trachyderini, Plate 171, 14 mm). Adults of this uncommon southeastern U. S. species are active from March–July. Larvae are known from *Quercus inopina*, Carya floridana, and Persea bourboni in Florida.
- Agallissus lepturoides (Chevrolat) (Agallissini, Plate 18b, 14 mm). This species ranges from Central America into Texas and may occur into coastal Arkansas and Louisiana. Adults are active from April–June. No larval hosts have been reported in the literature.

- Ancylocera bicolor (Olivier) (Trachyderini, Plate 17k, 12 mm). Adults of this southern U. S. species are active from April–July. Larvae feed in Carya, Quercus, and Acacia farnesiana.
- Aneflomorpha delongi (Champlain & Knull) (Elaphidiini, Plate 12k, 13 mm). Adults of this rare Florida and Georgia endemic are active from April—September. Larvae are known to develop in *Quercus laevis*.
- Aneflomorpha subpubescens (LeConte) (Elaphidiini, Plate 12l, 14 mm). Adults of this widespread eastern U. S. species are active from May–August. Larvae feed in live saplings of *Quercus* and *Castanea*.
- Anelaphus cinereus (Olivier) (Elaphidiini, Plate 13f, 9 mm). Adults of this widespread Caribbean and southeastern U. S. species are active from May–December. Larval hosts include Conocarpus erecta, Guaiacum officinale, Casuarina equisetifolia, and Zanthoxylum.
- Anelaphus inermis (Newman) (Elaphidiini, Plate 13i, 15 mm). Adults of this southern U. S., Bahamian, and Central American species are known from April–August. Larval hosts include Citrus, Quercus virginiana, Carya, Sideroxylon tenax, Avicennia germinans, and Piscidia piscipula.
- Anelaphus moestus (LeConte) (Elaphidiini, Plate 12i, 11 mm). Adults of this widespread central and eastern U. S. species are active from June–August. Larval hosts include *Juglans*, *Quercus* (including *Q. laurifolia*), *Celtis*, and *Rhus*.
- **Anelaphus mutatum** (Gahan) (Elaphidiini, Plate 13g, 17 mm). This species, formerly in the genus *Parelaphidion*, is known from Florida and the northern Caribbean. Adults are active in July–December. Larval hosts are unknown.
- Anelaphus parallelus (Newman) (Elaphidiini, Plate 13m, 12 mm). Adults of this widespread eastern North American species are active from May–August. Larvae are live twig pruners of most eastern hardwoods and shrubs, especially *Quercus*, but also *Crataegus viridis*, *Celtis tenuifolia*, and *Betula nigra*.
- Anelaphus pumilus (Newman) (Elaphidiini, Plate 13h, 11 mm). Adults of this eastern and central U. S. species are active from March–July. Larvae feed in *Quercus* (including *Q. phellos*), *Carya*, *Castanea*, *Ulmus*, *Betula nigra*, and *Tilia*.
- Anelaphus villosus (Fabricius) (Elaphidiini, Plate 131, 13 mm). Adults of this widespread eastern North American species are active from April–September. Larvae are live twig pruners of most eastern hardwoods and shrubs, especially *Carya* and *Tilia*.
- **Batyle ignicollis australis Linsley** (Trachyderini, Plate 19m, 10 mm). Adults of this southeastern U. S. subspecies are active from April–June. Larval hosts include *Pinus* and *Ouercus inopina* Ashe.
- *Batyle ignicollis ignicollis* (Say) (Trachyderini, Plate 19l, 11 mm). Adults of this common central and southern U. S. species are active from April—August. Larvae are stem borers in herbaceous plants and shrubs including *Rhus*. Adults can be collected on many wildflowers.
- **Batyle suturalis** (Say) (Trachyderini, Plate 21j, 9 mm). Adults of this widespread common central and southern U. S. species are active from April–September. Larvae develop in small dead branches of *Quercus*, *Carya*, *Celtis occidentalis*, *C. tenuifolia*, *C. laevigata*, *Castanea*, *Crataegus viridis*, and *Sapindus saponaria*. Adults are attracted to many wildflowers.
- Callidiellum rufipenne (Motschulsky) (Callidiini, Plate 211, two variants shown, 9 mm). In 1997, a population of this Japanese species was discovered in North Carolina. In 1998, infestations were discovered on live *Thuja occidentalis* in southwestern Connecticut. Native hosts in Japan include *Chamaecyparis*, Cryptomeria japonica, and Cupressus. Adults are active in spring and early summer.
- Callidium antennatum Newman (Callidiini, Plate 20h, 11 mm). Adults of this widespread northern and eastern U. S. species are active from April–July. Larvae feed under bark of recently dead or dying *Pinus* and rarely *Picea*.
- *Callidium frigidum* Casey (Callidiini, Plate 20e, 8 mm). Adults of this uncommon eastern North American species are active from April–June. Larvae feed under bark of *Juniperus* and *Chamaecyparis*.
- *Callidium schotti* Schaeffer (Callidiini, Plate 20g, 10 mm). Adults of this northeastern U. S. and southern Canada species are active from March–July. Larvae feed under *Pinus* bark and possibly also in *Chamaecyparis*.

- *Callidium texanum* Schaeffer (Callidiini, Plate 20f, 10 mm). Adults of this widespread U. S. species are active from May–July. Larvae feed under bark in *Juniperus virginiana* and *J. ashei*.
- *Callidium violaceum* (Linnaeus) (Callidiini, Plate 20i, 11 mm). This species, introduced from Europe, is widespread in southeast Canada and northeast United States. Adults are active from May–August. Larvae feed in dead conifers including *Pinus*, *Larix*, and *Picea*.
- *Callimoxys sanguinicollis* (Olivier) (Stenopterini, Plate 10f, two variants shown, 9 mm). Adults are active from April–July. Larvae feed in *Carya*, but also reportedly mine in *Ceanothus* and related plants.
- *Calloides nobilis* (Harris) (Clytini, Plate 21i, 20 mm). Adults of this rare eastern U. S. species are active from May–October. Larvae develop in dead or dying hardwoods including *Quercus* (including *Q. velutina*), *Fraxinus*, and *Castanea*.
- Chlorida festiva (Linnaeus) (Bothriospilini, Plate 16h, 21 mm). This species is widespread throughout the Neotropics and southern U. S. and is broadly polyphagous. Larval hosts include Mangifera indica, Schinopsis balansae, Delonix regia, Hymenaea courbaril, Casuarina equisetifolia, Gossypium, Acacia, Citrus, and Solanum, among others.
- Chlorophorus annularis (Fabricius) (Clytini, Plate 21e, 14 mm). There have been over 20 interceptions of this Asian species at ports and nurseries around North America in the last 10 years. There are no confirmed established populations in the U. S., however. Although Bambusa is the primary host, many others include Sinobambusa gibbosa, Spondias, Tectona, Citrus, Pyrus, Acer, and Gossypium.
- Clytoleptus albofasciatus (Castelnau & Gory) (Clytini, Plate 21h, 7 mm). Adults of this uncommon eastern U. S. species are active from May–August. Larvae mine in dead or dying Vitis and rarely Carya.
- *Clytus marginicollis* Castelnau & Gory (Clytini, Plate 14g, 7 mm). Adults of this eastern North American species are active from April–July. Larvae feed under bark of recently dead *Pinus* branches.
- *Clytus ruricola* (Olivier) (Clytini, Plate 14f, 8 mm). Adults of this eastern U. S. species are active from May–August. Larvae develop in *Acer*, primarily, but also *Betula*, *Carya*, *Fagus*, *Quercus*, *Tilia*, and others.
- *Curius dentatus* **Newman** (Curiini, Plate 17f, 7 mm). Adults of this uncommon eastern U. S. species are active from May–July. Larvae feed beneath the bark of small branches of both conifers and hardwoods, including *Carya floridana*.
- *Curtomerus fasciatus* (Fisher) (Elaphidiini, Plate 21b, 10 mm). Adults of this Florida and Cuban species have been collected in June. Larval hosts are unknown.
- Curtomerus flavus (Fabricius) (Elaphidiini, Plate 19g, 10 mm). Adults of this common and widespread southern U. S, Caribbean, and Pacific Island species are known from all months of the year. Larval hosts include Acacia farnesiana, Nicotiana, Casuarina equisetifolia, Coccolobis, and other genera.
- Cyrtophorus verrucosus (Olivier) (Anaglyptini, Plate 11d, two variants shown, 9 mm). Adults of this very common and widespread eastern U. S. species are active from March–July, and this species is often among the first encountered each year. Larvae feed in genera that include Acer, Betula, Carya, Castanea, Cercis, Cornus, Fagus, Quercus, Ulmus, and Pinus.
- Dryobius sexnotatus Linsley (Dryobiini, Plate 11i, 18 mm). Adults of this rare eastern and central U. S. species are active from March–September. Larvae feed in *Acer*, *Ulmus*, *Fagus*, and *Tilia* (*Acer saccharum* is the primary host).
- *Eburia cinereopilosa* Fisher (Eburiini, Plate 16a, 21 mm). Adults of this Cuban and Florida species are active from April–June. Larval hosts are unknown.
- *Eburia distincta* **Haldeman** (Eburiini, Plate 16c, 18 mm). Adults of this southeastern U. S. and Bahamian species are active in May–August. This species has been reared from *Baccharis halimifolia*.
- *Eburia haldemani* LeConte (Eburiini, Plate 16f, 22 mm). Adults of this common central U. S. species are active from May–July. Larvae feed in hardwoods, including *Celtis*, *Ulmus*, and possibly *Salix*, and also *Juniperus ashei*.

- *Eburia quadrigeminata* (Say) (Eburiini, Plate 16e, 21 mm). Adults of this common eastern U. S. species are active from April–September. Larvae develop in numerous hardwoods.
- **Eburia stigma** (Olivier) (Eburiini, Plate 16b, 21 mm). Adults of this southern U. S. and Caribbean species are active in May–August. Larvae are known from *Pinus caribaea*, *Lysiloma*, *Sideroxylon* (including *S. foetidissimum*) and *Avicennia germinans*.
- *Eburia stroheckeri* Knull (Eburiini, Plate 16d, 24 mm). Adults of this southern Florida endemic have been collected in early June. Host plants are unknown.
- *Elaphidion cryptum* Linsley (Elaphidiini, Plate 12e, 15 mm). Adults of this uncommon south Florida species have been collected on cut branches of *Cojoba arborea* and reared from *Metopium* and *Rhizophora mangle*.
- *Elaphidion irroratum* (Linnaeus) (Elaphidiini, Plate 12f, 16 mm). Adults of this widespread Caribbean and southern U. S. species are active from April–December. Larval hosts include *Avicennia germinans*, *Laguncularia racemosa*, *Spondias purpurea*, and *Albizia*.
- *Elaphidion knulli* Linsley (Elaphidiini, no figure). Adults of this rare Florida species have been reared from *Avicennia germinans* from September–June.
- *Elaphidion mucronatum* (Say) (Elaphidiini, Plate 12c, 14 mm). Adults of this widespread and common eastern North American species are active from April–October. Larvae develop in most eastern hardwoods and shrubs, as well as *Taxodium distichum*.
- **Elaphidion tectum LeConte** (Elaphidiini, Plate 12d, 16 mm). Adults of this south Florida species are active from June–August, and have been reared from *Avicennia germinans*. This species treatment includes *Elaphidion clavis* Linsley that will be synonymized with *E. tectum* in a future work.
- *Elytroleptus floridanus* (LeConte) (Trachyderini, Plate 18f, 9 mm). Adults of this uncommon southeastern U. S. species are active from May–June. Larvae are known to develop in *Quercus laevis*, *Q. inopina*, and *Q. phellos*.
- **Enaphalodes archboldi** Lingafelter & Chemsak (Elaphidiini, Plate 13d, 24 mm). Adults of this rare and endemic central Florida species are active mostly in September, but also records from June and August have been reported. Larval hosts are unknown, but probably include *Quercus*.
- *Enaphalodes atomarius* (**Drury**) (Elaphidiini, Plate 14b, 23 mm). Adults of this widespread central and eastern U. S. species are active from May–October. Larvae feed in various hardwood genera including *Quercus* and *Carya*.
- *Enaphalodes cortiphagus* (Craighead) (Elaphidiini, Plate 14c, 21 mm). Adults of this widespread, but not generally common eastern and central U. S. species are active from July–September. Larvae feed in living *Ouercus*.
- *Enaphalodes hispicornis* (Linnaeus) (Elaphidiini, Plate 13e, 31 mm). Adults of this widespread central and eastern U. S. species are active from May–October. Larvae feed in *Quercus*.
- *Enaphalodes rufulus* (Haldeman) (Elaphidiini, Plate 14a, 23 mm). Adults of this widespread eastern and central U. S. species are active from May–October. Larvae feed in living *Quercus* and *Acer*.
- *Euderces picipes* (Fabricius) (Tillomorphini, Plate 11g, two variants shown, 6 mm). Adults are active from April–August. Larvae feed in branches of numerous hardwoods, especially *Cercis canadensis*, *Prunus serotina*, *Quercus muhlenbergii*, *Salix*, and *Carya*.
- Euderces pini (Olivier) (Tillomorphini, Plate 11f, 8 mm). Adults are active from March–June. Larvae feed in various species including *Cornus florida*, *Ulmus alata*, *Carya illinoinensis*, and *Fagus grandifolia*
- Euderces reichei reichei LeConte (Tillomorphini, Plate 11e, 4 mm). Adults of this widespread eastern U. S. species are active from March–June. Larvae feed in various hardwoods including Diospyros virginiana, D. texana, Prosopis glandulosa, Crataegus viridis, Celtis tenuifolia, C. laevigata, and Parkinsonia aculeata. Adults have been collected on Salix, Zanthoxylum, and Cornus.
- Euryscelis suturalis (Olivier) (Clytini, Plate 17a, male on left, female on right, 17 mm). This wide-spread Caribbean species that also occurs in southern Florida is active throughout the year, but in the summer months for U. S. populations. Hosts include Metopium toxiferum, Prosopis juliflora, and Sideroxylon foetidissimum.

- *Glycobius speciosus* (Say) (Clytini, Plate 11h, 23 mm). Adults of this rare eastern U. S. species are active from June–August. Larvae develop in living *Acer saccharum*.
- *Gracilia minuta* (Fabricius) (Graciliini, Plate 19h, 5 mm). This widespread European species has been reported in the eastern United States, but existing populations are unconfirmed. Adults are active from May–July. Larvae feed in dead branches of numerous shrubs and hardwoods, including *Quercus*, *Carya*, and *Salix*.
- *Hesperophanes pubescens* (Haldeman) (Hesperophanini, Plate 19b, 16 mm). Adults of this uncommon eastern and central U. S. species are active from July–August. Larval feeding habits are unknown.
- *Heterachthes ebenus* Newman (Ibidionini, Plate 18h, 9 mm). Adults of this uncommon eastern and central U. S. species are active from April–August. Larval hosts include *Pinus*, *Acacia farnesiana*, and *Quercus inopina*.
- *Heterachthes quadrimaculatus* **Haldeman** (Ibidionini, Plate 18g, 10 mm). Adults of this eastern U. S. species are active from June–August. Larval hosts include *Carya* and *Liriodendron*.
- *Heterachthes sablensis* Blatchley (Ibidionini, Plate 18i, 9 mm). Adults of this rare endemic Florida species are active in February–March. The only known larval host is *Avicennia germinans*.
- *Heterops dimidiatus* (Chevrolat) (Heteropsini, Plate 15g, 11 mm). This species occurs in the West Indies and has been collected rarely in Florida in May. One known host is *Pithecellobium dulce* for Cuban specimens. It has also been collected by beating *Bursera simaruba*.
- *Hylotrupes bajulus* (Linnaeus) (Callidiini, Plate 21c, 15 mm). This species, introduced from Europe, is widespread in the eastern United States. Adults are active from June–August. Larvae bore in dry conifer wood including *Pinus*, *Abies*, and *Picea* commonly used in building construction.
- *Knulliana cincta cincta* (**Drury**) (Bothriospilini, Plate 16k, 17 mm). Adults are active from March–September. Larvae feed in dry branches of hardwoods including *Quercus*, *Carya*, and *Salix*.
- *Knulliana cincta spinifera* (Fabricius) (Bothriospilini, Plate 16l, 18 mm). Adults of this widespread southeastern U. S., Caribbean, and Mexican species are active from July–August. Larval hosts include *Carya illinoinensis* and *Ostrya virginiana*.
- *Linsleyonides albomaculatus* (Champlain & Knull) (Elaphidiini, Plate 12g, 8 mm). Adults of this uncommon Caribbean species are occasionally collected in Florida from May–June. Larval hosts are unknown.
- Megacyllene caryae (Gahan) (Clytini, Plate 12b, 16 mm). Adults are active from March–June. Larvae develop in recently dead Carya, and sometimes in other hardwoods including Cercis canadensis, Gleditsia, Prosopis juliflora, Maclura pomifera, Fraxinus americanus, and others. Adults can be collected on Solidago and Grindelia, among other flowers.
- *Megacyllene decora* (Olivier) (Clytini, Plate 11j, two variants shown, 18 mm). Adults are active from June–October. Larvae feed in stems of *Amorpha fruticosa*.
- *Megacyllene robiniae* (Forster) (Clytini, Plate 12a, 16 mm). Adults of this widespread North American species are active from June–November. Larvae mine in *Robinia pseudoacacia* and other locust species. Adults are often collected on *Solidago*.
- *Meriellum proteus* (Kirby) (Callidiini, Plate 21m, 14 mm). Adults of this uncommon northern U. S. species are active from June–July. Larvae develop in dead *Picea* and *Pinus*.
- *Methia necydalea* (Fabricius) (Methiini, Plate 10h, 4 mm). This species is widespread in the Caribbean and southern U. S. Adults are active from May–September. Larvae have been reared from *Quercus virginiana* and *O. phellos*.
- *Michthisoma heterodoxum* LeConte (Saphanini, Plate 16g, 6 mm). Adults of this uncommon southeastern U. S. species are active from April–July. Larvae feed in *Carya* and *Quercus*.
- *Micranoplium unicolor* (Haldeman) (Elaphidiini, Plate 19f, 7 mm). Adults of this widespread but uncommon eastern and central U. S. species are active from May–July. Larval feeding habits are unknown.
- Microclytus gazellula (Haldeman) (Anaglyptini, Plate 21g, 7 mm). Adults are active from May–June. Larvae feed in living Quercus and Carya glabra. Note that the similar species, M. compressicollis (Castelnau & Gory) is not included in the key since it is restricted to extreme northeast U. S. and Canada. It can be recognized by having the second antennomere less than half length of fourth (in M. gazellula, the second antennomere is about as long as fourth).

- Molorchus bimaculatus bimaculatus Say (Molorchini, Plate 11a, 6 mm). Adults of this common eastern and central U. S. species are active from March–July. Larvae develop in dead branches of numerous hardwoods including Acer, Carya, Quercus muhlenbergii, Crataegus viridis, Rhus glabra, Cercis canadensis, Juglans nigra, and Celtis tenuifolia.
- *Molorchus bimaculatus corni* Haldeman (Molorchini, Plate 11b, 6 mm). Adults of this southeastern U. S. subspecies are active from April–August. Larvae develop in *Cornus floridanus* and *Quercus geminata*.
- *Molorchus bimaculatus semiustus* (Newman) (Molorchini, Plate 11c, 6 mm). Adults of this subspecies are active from April–May. Larvae develop in *Crataegus viridis*, *Carya illinoinensis*, and *Vitis*. Adults are often collected on flowers of *Cornus*.
- Neoclytus acuminatus (Fabricius) (Clytini, Plate 17b, 11 mm). Adults of this widespread eastern North American species are active from March–October. Larvae develop in most eastern hardwoods, especially Fraxinus (including F. quadrangulata), Quercus (including Q. velutina, Q. phellos, Q. muhlenbergii, and Q. macrocarpa), Carya (including C. ovata and C. illinoinensis), Diospyros virginiana, Celtis (including C. tenuifolia), Crataegus viridis, Salix nigra, and Prunus serotina, among others.
- **Neoclytus caprea** (Say) (Clytini, Plate 14e, 15 mm). Adults are active from March–June. Larvae develop in *Fraxinus*, *Quercus* (including *Q. vaseyana* var. *vaseyana* and *Q. emoryi*), *Acer*, *Diospyros virginiana*, and *Celtis laevigata* var. *reticulata*.
- Neoclytus cordifer (Klug) (Clytini, Plate 16j, 12 mm). This species occurs in the Caribbean and southeastern U. S. Adults are active from April through July. Larvae develop in Avicennia germinans, Rhizophora mangle, Citrus, Mangifera indica, Punica granatum, Quercus geminata, Carya floridana, and Ximenia americana.
- *Neoclytus horridus* (LeConte) (Clytini, Plate 17e, 8 mm). Adults of this uncommon eastern U. S. species are active from April–July. Larvae feed in *Quercus*.
- *Neoclytus jouteli jouteli Davis* (Clytini, Plate 17c, 7 mm). Adults of this uncommon eastern U. S. subspecies are active from May–September. Larvae feed in *Carya*, *Quercus laevis*, *Q. velutina*, *Q. geminata*, and *Q. alba*.
- *Neoclytus jouteli simplarius* **Blatchley** (Clytini, Plate 17d, 7 mm). Adults of this uncommon southern U. S. subspecies have been collected in March and April. Larval hosts include *Ampelopsis arborea*, *Quercus nigra*, and *Q. laevis*.
- **Neoclytus longipes** (**Drury**) (Clytini, Plate 16i, 12 mm). This species is evidently quite rare in the U. S., known only from a few specimens in Florida and Georgia, but is more widespread in the Greater Antilles. Adults are active from April through July. Larvae are known to develop in *Pimenta dioica*.
- Neoclytus mucronatus (Fabricius) (Clytini, Plate 14d, 14 mm). Adults of this common eastern U. S. species are active from April—October. Larvae develop in Carya (including Carya illinoinensis), Diospyros virginiana, Celtis laevigata and rarely Pinus.
- **Neoclytus scutellaris** (Olivier) (Clytini, Plate 14h, 11 mm). Adults of this common U. S. species are active from May–October. Larvae typically develop in *Quercus* (including *Q. lyrata*, *Q. palustris*, *Q. phellos*, *Q. velutina*, and *Q. stellata*) and *Carya*.
- Obrium maculatum (Olivier) (Obriini, Plate 17h, 5 mm). Adults of this common eastern U. S. species are active from April–October. Larvae feed in numerous hardwoods and shrubs, including Quercus (e.g., Q. phellos), Carya (including C. illinoinensis), Crataegus viridis, Betula nigra, Prunus serotina, Celtis occidentalis, and C. laevigata var. reticulata.
- *Obrium rubidum* LeConte (Obriini, Plate 17j, 8 mm). Adults of this eastern U. S. species are active in May. Larval feeding habits are unknown.
- *Obrium rufulum* Gahan (Obriini, Plate 17i, 5 mm). Adults of this eastern U. S. species are active from May–August. Larvae feed in dead branches of *Fraxinus*, *Tilia* and *Quercus*.
- *Oeme rigida* (Say) (Oemini, Plate 18j, 18 mm). Adults of this eastern U. S. species are active from June–September. Larvae feed primarily in *Juniperus* and *Taxodium distichum*, but rarely also *Pinus*.
- *Osmopleura chamaeropis* (Horn) (Agallissini, Plate 18a, 19 mm). Adults of this uncommon Florida species are active from March through May. Larvae develop in *Sabal palmetto*.

- **Parelaphidion aspersum** (Haldeman) (Elaphidiini, Plate 13k, 16 mm). Adults of this widespread eastern and central U. S. species are active from June–October. Larvae develop in *Quercus* (including *Q. velutina*), Carya, Betula nigra, and Celtis laevigata.
- Parelaphidion incertum (Newman) (Elaphidiini, Plate 13j, 16 mm). Adults of this widespread eastern and central U. S. species are active from April–September. Larvae feed primarily in living Morus, but also Quercus and Carya.
- **Penichroa fasciata** (**Stephens**) (Hesperophanini, Plate 21a, 5 mm). Adults are active from June–August. This widespread European species potentially has isolated established populations in the United States, but these are unconfirmed. Larvae feed in twigs of *Carya* and other hardwoods.
- **Phymatodes aereus** (Newman) (Callidiini, Plate 19e, 7 mm). Adults of this uncommon eastern U. S. species are active from April–July. Larvae develop in dead *Quercus* and *Castanea*.
- **Phymatodes amoenus** (Say) (Callidiini, Plate 20a, 7 mm). Adults of this locally common eastern U. S. species are active from April–August. Larvae mine in dead *Vitis*.
- **Phymatodes lengi Joutel** (Callidiini, Plate 20b, 5 mm). Adults of this rare eastern U. S. species are active from June–July. Larval feeding habits are unknown.
- Phymatodes testaceus (Linnaeus) (Callidiini, Plate 19d, two variants shown, 13 mm). This species, introduced from Europe, is widely established in the north-central, eastern and southeastern U.
 S. Adults are active from April–July. Larvae feed under bark of Quercus, Castanea, Fagus, Malus, Prunus, and Carya.
- **Phymatodes varius** (**Fabricius**) (Callidiini, Plate 20j, 8 mm). Adults of this locally common eastern U. S. species are active from April–July. Larvae feed under bark of *Quercus* (including *Q. stellata* and *Q. vaseyana* var. *vaseyana*) and *Carya*.
- Physocnemum andreae (Haldeman) (Callidiini, Plate 20d, 18 mm). This species is uncommon and occurs along the Atlantic coast. Adults are active from June–July. Larvae feed under Cupressus bark
- **Physocnemum brevilineum** (Say) (Callidiini, Plate 20c, 16 mm). Adults of this uncommon eastern and central U. S. species are active from April–August. Larvae feed in living *Ulmus*.
- Placosternus difficilis (Chevrolat) (Clytini, Plate 14i, 15 mm). Adults of this widespread southern U. S., Caribbean, and Mexican species are active from March–October. Larval hosts include Prosopis juliflora, Acacia, Celtis, and Platanus.
- Plectromerus dentipes (Olivier) (Curiini, Plate 17g, 6 mm). Adults of this southern U. S. and Caribbean species are active in many months of the year and hosts include Quercus, Carya illinoinensis, Cercis canadensis, Conocarpus erectus, Crossopetalum rhacoma, Conocarpus erectus, Cojoba arborea, and Taxodium distichum.
- *Plesioclytus relictus* Giesbert (Clytini, Plate 19c, 9 mm). Adults of this rare south-central Florida endemic are active from April–August. Larval hosts are unknown.
- **Plinthocoelium suaveolens suaveolens (Linnaeus)** (Callichromatini, Plate 15e, 26 mm). Adults of this southeastern U. S. species are active from May–September. Hosts include *Sideroxylon* and possibly *Carya*.
- *Pronocera collaris* (Kirby) (Callidiini, Plate 19k, 10 mm). Adults of this widespread northern U. S. species are active from June–August. Larvae feed in *Pinus* and *Picea*.
- **Psyrassa pertenuis** (Casey) (Elaphidiini, Plate 13b, 10 mm). Adults of this eastern U. S. species are active from the June–July. Larvae feed in various hardwoods including *Carya* and *Prunus serotina* and also *Magnolia* and *Conocarpus erectus*.
- Psyrassa unicolor (Randall) (Elaphidiini, Plate 13a, 11 mm). Adults of this widespread eastern U. S. species are active from May–August. Larvae girdle twigs of numerous hardwoods, especially Quercus, Carya (including C. illinoinensis), Cercis canadensis and Fagus.
- **Purpuricenus axillaris Haldeman** (Trachyderini, Plate 15i, 13 mm). Adults of this widespread eastern U. S. species are active from May–August. Larvae develop in branches of *Quercus*, *Carya*, and *Castanea*.
- *Purpuricenus humeralis* (Fabricius) (Trachyderini, Plate 15h, 15 mm). Adults of this widespread, but generally uncommon eastern species are active from June–August. Larvae mine in dead branches of numerous hardwoods.

- *Purpuricenus paraxillaris* MacRae (Trachyderini, Plate 15j, 16 mm). Adults of this eastern U. S. species are active in June–August. Larval hosts include *Castanea dentatus* and *Quercus velutina*.
- **Rhopalophora longipes** (Say) (Rhopalophorini, Plate 18d, 9 mm). Adults of this common central and southern U. S. species are active from May–August. Larvae feed in small branches of *Cercis canadensis* and *Cornus*. Adults have been collected on blossoms of *Crataegus*, *Rhus*, and *Cercis*.
- **Romulus globosus Knull** (Elaphidiini, Plate 12h, 33 mm). Adults of this rare Florida endemic are active from May–August. Larval hosts are unknown, but probably include *Quercus*.
- **Ropalopus sanguinicollis** (Horn) (Callidiini, Plate 19j, 14 mm). Adults of this northeast U. S. species are active from June–July. Larvae feed under bark of living *Prunus serotina*.
- Sarosesthes fulminans (Fabricius) (Clytini, Plate 21d, 15 mm). Adults of this widespread but generally uncommon eastern U. S. species are active from May–August. Larvae develop primarily in *Castanea*, *Quercus*, and *Juglans*.
- Semanotus ligneus (Fabricius) (Callidiini, Plate 20k, 10 mm). Adults of this northern U. S. and southern Canada species are active from March–August. Larvae develop in *Chamaecyparis*, *Juniperus*, and *Thuja*.
- Smodicum cucujiforme (Say) (Smodicini, Plate 18k, 9 mm). Adults of this common eastern and central U. S. species are active from June–September. Larvae feed in various hardwoods including Carya, Acer, Quercus, and Ulmus rubra.
- Stenosphenus notatus (Olivier) (Elaphidiini, Plate 13c, 12 mm). Adults of this central and eastern U. S. species are active from March–September. Larvae feed in dead limbs of *Carya* (including *C. aquatica* and *C. laciniosa*) and *Celtis*.
- *Stizocera floridana* Linsley (Elaphidiini, Plate 12j, 10 mm). Adults of this Florida endemic are active in April–May. This species has been reared from *Forestiera segregata* var. *segregata*.
- Stromatium fulvum (Villers) (Hesperophanini, Plate 19a, 19 mm). There are a few records of this European species having been introduced into eastern U. S., although current established populations cannot be confirmed. Adults there are active from May through September. Larval hosts include Acacia, Carya, Fagus, Juglans nigra, Morus, Platanus, Prunus, Quercus, Ulmus, and others.
- **Tessaropa tenuipes** (Haldeman) (Methiini, Plate 10i, two variants shown, 6 mm). Adults of this uncommon central and eastern U. S. species are active from April–May. Larvae develop in small dead branches of various hardwoods including *Quercus*, *Carya*, *Juglans*, and *Betula*. Adults have been collected by beating *Vitis*.
- *Tilloclytus geminatus* (Haldeman) (Anaglyptini, Plate 21f, 6 mm). Adults of this widespread eastern U. S. species are active from May–July. Larvae mine dead branches of *Carya*, *Quercus* (including *Q. bicolor*), *Betula nigra*, *Crataegus viridis*, *Celtis tenuifolia*, *Prunus americana*, and *Pinus*. Adults can also be collected from beating *Vitis*.
- *Trachyderes mandibularis* **Dupont** (Trachyderini, Plate 15f, 20 mm). This species is widespread throughout the Neotropics and southern U. S. Adults are active from June–August. Larval hosts include *Baccharis*, *Parkinsonia*, *Acacia*, *Pithecellobium*, *Ficus*, and *Salix*. Adults are attracted to rotting fruit, and flowers of *Croton*, *Solidago*, and other genera.
- *Tragidion coquus* (Linnaeus) (Trachyderini, Plate 15k, two variants shown, 19 mm). Adults of this central and eastern U. S. species are active from June–November. Larvae develop in *Quercus* and *Cercis*. Adults can be collected on flowering plants such as *Helianthus annuus* and *Grindelia*.
- *Tylonotus bimaculatus* **Haldeman** (Hesperophanini, Plate 20l, 13 mm). Adults of this widespread but uncommon eastern U. S. species are active from May–August. Larvae feed in live or dying hardwoods, especially *Fraxinus*, but also *Ulmus*, *Carya*, *Juglans nigra*, *J. cinerea*, and *Betula*.
- *Tylonotus masoni* (**Knull**) (Hesperophanini, Plate 19i, 11 mm). Adults of this very rare eastern U. S. species are active from June–August. Larval feeding habits are unknown.
- *Xylotrechus aceris* Fisher (Clytini, Plate 14j, 10 mm). Adults of this uncommon eastern U. S. species are active from June–August. Larvae develop in live *Acer*, inducing galls.

- *Xylotrechus annosus annosus* (Say) (Clytini, Plate 15d, 12 mm). Adults of this uncommon eastern U. S. species are active from May–July. Larvae develop in dead and dying *Salix* and *Populus*.
- *Xylotrechus colonus* (Fabricius) (Clytini, Plate 15a, 12 mm). Adults of this very common and widespread species are active from April–October. Larvae develop in virtually all eastern hardwoods, especially *Carya*, *Fagus*, and also *Pinus*.
- *Xylotrechus convergens* LeConte (Clytini, Plate 14m, 10 mm). Adults of this uncommon eastern U. S. species are active from June–July. Larvae mine in *Crataegus viridis*.
- *Xylotrechus integer* (Haldeman) (Clytini, Plate 15c, 13 mm). Adults of this northeastern long-horned beetle are active from June–July. Larvae feed in *Abies balsamea* and *Tsuga*.
- *Xylotrechus nitidus* (Horn) (Clytini, Plate 14l, 10 mm). Adults of this northeastern and central U. S. species are active from June–July. Larval feeding habits are unknown.
- *Xylotrechus quadrimaculatus* (Haldeman) (Clytini, Plate 14k, 12 mm). Adults of this northeastern U. S. and southern Canada species are active from May–August. Larvae are girdlers of live branches of *Fagus*, *Betula*, *Alnus*, and *Carpinus caroliniana*.
- *Xylotrechus sagittatus* (Germar) (Clytini, Plate 15b, 14 mm). This species is widepread in the U. S. and Mexico. Adults are active from June–September. Larvae feed in conifers, especially *Pinus*.
- *Xylotrechus schaefferi* **Schott** (Clytini, Plate 14n, 7 mm). Adults of this uncommon northeastern U. S. species are active from June–August. Larvae feed in cones of *Pinus rigida* and *P. banksiana*.
- **Zagymnus clerinus** (LeConte) (Agallissini, Plate 18c, two variants shown, 13 mm). Adults of this uncommon Florida species are active in June. Larvae develop in *Sabal palmetto*.

LAMIINAE

- Acanthocinus nodosus (Fabricius) (Acanthocinini, Plate 31a, 24 mm). Adults of this widespread southern U. S. species are active from April–October. Larvae develop in dead or dying pine, especially *Pinus rigida* and *P. taeda*.
- *Acanthocinus obsoletus* (Olivier) (Acanthocinini, Plate 31d, 12 mm). Adults of this uncommon eastern U. S. species are active from April–September. Larvae feed in *Pinus*.
- Acanthocinus pusillus Kirby (Acanthocinini, Plate 31e, 9 mm). Adults of this eastern species are active from May–August. Larvae feed in dead or dying *Pinus* and other conifers, especially *Abies* and *Tsuga*.
- *Adetus brousi* (Horn) (Apomecynini, Plate 26h, 7 mm). Adults of this uncommon central and eastern U. S. species are active from June–July. Known larval hosts inculde *Cucurbita*.
- Aegomorphus modestus (Gyllenhal) (Acanthoderini, Plate 30h, 11 mm). Adults of this widespread eastern U. S. and Bahamian species are active from May–September. Larvae feed in hardwoods such as Acer, Betula, Carya, Castanea, Celtis, Fagus, Fraxinus, and many others, as well as Pinus.
- **Aegomorphus morrisii** (Uhler) (Acanthoderini, Plate 30g, 16 mm). Adults of this rare and localized eastern U. S. species are active from June–July. Larvae feed *Nyssa aquatica*, *N. sylvatica*, and possibly *Liriodendron*.
- Aegomorphus quadrigibbus (Say) (Acanthoderini, Plate 30f, 10 mm). Adults are acive from April–September. Larvae feed in tree genera including Acer, Betula, Carya, Castanea, Celtis, Cercis, Fagus, Liriodendron, Quercus, Ulmus, and others.
- Alcidion umbraticus (Jacquelin du Val) (Acanthocinini, Plate 28f, 7 mm). Adults of this primarily Caribbean species that also occurs in Florida can be collected from June–December. Larval hosts are unknown but may include *Solanum* species.
- Anoplophora glabripennis (Motschulsky) (Lamiini, Plate 24g, 30 mm). This species, widespread in China and Korea, was introduced into New York in 1996 and Chicago in 1998. Referred to as the Asian Longhorned Beetle, adults are active from June–September. Known larval hosts include Acer, Aesculus, Albizia, Betula, Celtis, Platanus, Populus, Salix, Sorbus, and Ulmus, but probably many others can be used by this species.
- Astylidius parvus (LeConte) (Acanthocinini, Plate 29b, 5 mm). Adults of this eastern species are active from May–August. Larvae feed in various hardwoods, shrubs, and vines including *Acer*, *Celtis*, *Diospyros virginiana*, *Morus*, *Ulmus*, and others.

- Astylopsis arcuatus (LeConte) (Acanthocinini, Plate 28i, 8 mm). Adults of this eastern U. S. species are active from May through October. No larval hosts have been determined.
- Astylopsis collaris (Haldeman) (Acanthocinini, Plate 28k, 8 mm). Adults of this eastern U. S. species are active from May–August. Larvae feed in various hardwoods, especially *Quercus* and *Rhus*.
- Astylopsis fascipennis Schiefer (Acanthocinini, Plate 29a, 8 mm). This species is known only from southeastern U. S. Adults are active from June–July. Hosts may include *Ulmus* and *Liquidambar styraciflua*.
- Astylopsis macula (Say) (Acanthocinini, Plate 28g, 8 mm). Adults of this common eastern U. S. species are active from May–September. Larvae feed in hardwoods, shrubs, and vines, especially *Castanea*, *Tilia*, and *Acer*.
- Astylopsis perplexa (Haldeman) (Acanthocinini, Plate 28j, 10 mm). This species occurs in the southeastern U. S. to Texas. The only known larval host is *Baccharis halimifolia*.
- Astylopsis sexguttata (Say) (Acanthocinini, Plate 28h, 7 mm). Adults are active from April–September. Larvae feed under bark of conifers, especially Larix, Picea, and Pinus.
- Ataxia crypta (Say) (Pteropliini, Plate 26m, 14 mm). Adults of this southern U. S. species are active from April—October. Larvae feed in the branches of *Quercus*, *Castanea*, and *Pyrus*, and also in *Xanthium*, *Verbesina*, *Ambrosia*, *Smilax*, and *Gossypium*.
- Ataxia falli Breuning (Pteropliini, Plate 26k, 13 mm). Adults of this uncommon Florida species are active from April–May. This species has been reared from *Piscidia piscipula* in Florida and adults have been collected by beating *Rhizophora mangle* and *Metopium*.
- *Ataxia hubbardi* Fisher (Pteropliini, Plate 26j, 13 mm). Adults of this central U. S. species are active from April–October. Larvae are stem borers of living herbaceous plants, mostly composites, including *Helianthus*.
- Ataxia spinicauda Schaeffer (Pteropliini, Plate 26h, 11 mm). Adults of this uncommon Florida and Caribbean species are active from June–December. Little is known of its habits, although one specimen (along with A. crypta and A. falli) was taken on fresh cut Metopium in June in Florida.
- *Cyrtinus pygmaeus* (Haldeman) (Cyrtinini, Plate 26a, 3 mm). Adults of this widespread eastern U. S. species are active from March–July. Larvae feed in the dry branches of numerous hardwoods, especially *Quercus*. Adults have been collected by beating *Nyssa*, *Sapindus*, and many other trees.
- **Dectes sayi** Dillon & Dillon (Acanthocinini, Plate 30d, 7 mm). Adults of this common central U. S. species are active from May–September. Larvae are stem borers of herbaceous plants, especially *Ambrosia artemisifolia*, but also *Eupatorium*, *Helianthus* and *Xanthium*.
- **Dectes texanus** LeConte (Acanthocinini, Plate 30c, 8 mm). Adults of this common central U. S. species are active from May–September. Larvae are stem borers of *Ambrosia*, *Gaillardia*, *Helianthus*, *Solidago*, and many other herbaceous plants.
- **Desmiphora hirticollis** (Olivier) (Desmiphorini, Plate 26b, 10 mm). This summer active species is widespread throughout the Caribbean and into Mexico, Central and South America, and the southern U. S. Larval hosts include *Cordia*, among others.
- **Dorcaschema alternatum** (Say) (Dorcaschematini, Plate 26c, 10 mm). Adults of this widespread central and eastern U. S. species are active from April–September. Larvae feed in dead or dying branches of *Morus*, *Maclura pomifera*, and *Aesculus glabra*.
- **Dorcaschema cinereum** (Olivier) (Dorcaschematini, Plate 26f, 9 mm). Adults are active from May–August. Larvae feed in dead branches of various hardwoods including *Morus* and *Aesculus glabra*.
- **Dorcaschema nigrum** (Say) (Dorcaschematini, Plate 26d, 9 mm). Adults are active from May–August. Larvae develop in *Carya* and *Aesculus glabra*.
- **Dorcaschema wildii Uhler** (Dorcaschematini, Plate 26e, 16 mm). Adults of this uncommon but widespread central and eastern U. S. species are active from May–October. Larvae develop in live *Morus*, *Maclura pomifera*, and *Aesculus glabra*.
- Dorcasta cinerea (Horn) (Apomecynini, Plate 25m, 8 mm). Adults of this common central and southern U. S. species are active from June–October. Larvae are stem borers of living herbaceous plants including Solanum, Gossypium, Nicotiana, Matelea, Helianthus annuus, Verbesina, Croton capitatus, and others. Adults have also been collected on Medicago.

- *Ecyrus dasycerus* (Say) (Pogonocherini, Plate 261, 7 mm). Adults of this eastern U. S. species are active from April–August. Larvae feed in numerous hardwoods, especially *Quercus* and *Celtis occidentalis*.
- *Eupogonius annulicornis* Fisher (Desmiphorini, Plate 27d, 4 mm). Adults of this generally uncommon eastern U. S. species are active from March–August. Larvae feed in numerous hardwoods, shrubs, and vines including *Bursera simaruba*.
- *Eupogonius pauper* LeConte (Desmiphorini, Plate 30b, 5 mm). Adults of this common eastern U. S. species are active from March–August. Larvae feed in numerous hardwoods, shrubs, and vines.
- *Eupogonius subarmatus* (LeConte) (Desmiphorini, Plate 291, 5 mm). Adults of this common eastern U. S. species are active from May–August. Larvae feed in various hardwoods, including *Tilia* and *Ulmus*.
- *Eupogonius tomentosus* (Haldeman) (Desmiphorini, Plate 30a, 6 mm). Adults of this common eastern U. S. species are active from March–November. Larvae feed in *Pinus*, *Picea*, and *Chamaecyparis*.
- *Eutrichillus biguttatus* (LeConte) (Acanthocinini, Plate 301, 7 mm). Adults of this generally uncommon eastern U. S. species are active from May–August. Larvae develop in *Pinus*.
- *Goes debilis* LeConte (Lamiini, Plate 25f, 12 mm). Adults of this widespread eastern U. S. species are active from May–August. Larvae feed in living *Quercus*. *Carya* and *Castanea* are also mentioned in some literature as potential hosts.
- Goes pulcher (Haldeman) (Lamiini, Plate 25c, 22 mm). Adults of this uncommon species are active from May–August. Larvae feed in living Carya (including C. illinoinensis). Ulmus and Quercus are potential hosts.
- Goes pulverulentus (Haldeman) (Lamiini, Plate 25d, 20 mm). Adults of this widespread but uncommon eastern U. S. species are active from May–August. Larvae feed in *Quercus*, *Fagus*, and *Platanus*. *Pinus* and *Prunus serotina* are potential hosts.
- *Goes tesselatus* (Haldeman) (Lamiini, Plate 25h, 25 mm). Adults of this widespread eastern U. S. species fly from May–September. Larvae feed in living hardwoods, especially *Quercus*.
- Goes tigrinus (DeGeer) (Lamiini, Plate 25i, 29 mm). Adults of this widespread but uncommon eastern U. S. species are active from May–October, but most abundant in June–July. Larvae feed in living hardwoods, especially *Quercus*.
- Goes tumifrons Chemsak & Linsley (Lamiini, Plate 25g, 24 mm). Adults of this uncommon eastern U. S. species are active from July–September. Larval feeding habits are unknown.
- Goes variegatus Linsley & Chemsak (Lamiini, Plate 25e, 12 mm). Adults of this uncommon eastern and central U. S. species are active from June–July. Larval feeding habits are unknown
- *Hebestola nebulosa* Haldeman (Lamiini, Plate 24e, 10 mm). Adults of this uncommon eastern U. S. species are active from May–July. Larvae feed in *Quercus* and *Castanea*.
- *Hemierana marginata* (Fabricius) (Hemilophini, Plate 22h, 6 mm). Adults of this central and eastern U. S. species are active from April–August. Larvae feed in *Vernonia* and *Lithospermum caroliniense*. Adults are associated with *Heliopsis helianthoides* var. *occidentalis*.
- *Hippopsis lemniscata* (Fabricius) (Agapanthiini, Plate 25n, 10 mm). Adults of this common central and southern U. S. species are active from April–October. Larvae are stem borers of living herbaceous plants, mostly composites (especially *Ambrosia*).
- *Hyperplatys aspersa* (Say) (Acanthocinini, Plate 31g, 5 mm). Adults are active from March–September. Larval hosts include various hardwoods as well as *Menispermum* and *Arctium*.
- *Hyperplatys femoralis* Haldeman (Acanthocinini, Plate 31i, 5 mm). This uncommon species occurs in the southeastern U. S. Adults have been collected in April and May. No larval hosts are known.
- Hyperplatys maculata Haldeman (Acanthocinini, Plate 31h, 5 mm). Adults are active from May–October. Larvae feed in various hardwoods, especially Tilia, Menispermum, Amelanchier arborea, and Aesculus pavia.
- Lagocheirus araneiformis stroheckeri Dillon (Acanthocinini, Plate 30j, 18 mm). This southern Florida and Cuban subspecies is active most months of the year. The only known host is Bursera simaruba.

- *Leptostylopsis albofasciatus* (Fisher) (Acanthocinini, Plate 29h, 6 mm). This species is known only from Florida and Cuba. The only known larval host is *Rhizophora mangle*.
- Leptostylopsis argentatus (Jacquelin du Val) (Acanthocinini, Plate 29d, 9 mm). Adults of this very common Florida and Caribbean species can be collected from June–December. Larval hosts include Conocarpus erectus, Zanthoxylum fagara, and Zanthoxylum flavum.
- Leptostylopsis planidorsus (LeConte) (Acanthocinini, Plate 29i, 8 mm). Adults of this southeast U. S. species are active from May–August. Larvae feed in *Betula*, *Cercis canadensis*, *Vitis*, and *Quercus laurifolia*.
- Leptostylopsis terraecolor (Horn) (Acanthocinini, Plate 29g, 9 mm). This species is endemic to Florida. It develops in *Rhizophora mangle*, *Ficus citrifolia*, *Metopium*, *Cojoba arborea*, *Piscidia piscipula*, *Vitis*, *Ficus aurea*, *Bursera simaruba*, *Forestiera segregata*, and *Rhizophora mangle*.
- *Leptostylus asperatus* (Haldeman) (Acanthocinini, Plate 29e, 8 mm). Adults of this southern U. S. species are active from April–August. Larvae feed in *Quercus* and *Rhus*.
- Leptostylus transversus (Gyllenhal) (Acanthocinini, Plate 29f, 8 mm). Adults of this common eastern species are active from March–October. Larvae feed in various hardwoods, vines, and conifers including, among others, Quercus stellata, Q. macrocarpa, Aesculus pavia, and Pinus.
- Lepturges angulatus (LeConte) (Acanthocinini, Plate 32j, 7 mm). Adults of this common eastern and central U. S. species are active from March–August. Larvae feed in various trees including *Pinus*, *Amelanchier arborea*, *Gymnocladus dioicus*, *Celtis tenuifolia*, *Aesculus pavia*, *Celtis laevigata*, and *Parkinsonia aculeata*.
- **Lepturges confluens (Haldeman)** (Acanthocinini, Plate 32k, 7 mm). Adults of this common eastern and central U. S. species are active from May–August. Larvae feed in various hardwoods, including *Carya laciniosa* and *C. illinoinensis*.
- *Lepturges megalops* **Hamilton** (Acanthocinini, Plate 31j, 6 mm). This rare species is known from Florida and Panama. No larval hosts are known.
- Lepturges pictus (LeConte) (Acanthocinini, Plate 32i, 6 mm). Adults of this uncommon eastern U. S. species are active from May–August. Larvae feed in *Celtis* (including *C. occidentalis* and *C. tenuifolia*), *Juglans*, and *Carya*.
- *Lepturges regularis* (LeConte) (Acanthocinini, Plate 31f, 6 mm). Adults are active from May–October. Larvae feed in the branches of *Aesculus pavia* and sometimes *Carya*.
- *Lepturges symmetricus* (Haldeman) (Acanthocinini, Plate 32h, 6 mm). Adults of this uncommon species are active from May–August. Larvae develop in the branches of *Tilia*.
- *Liopinus alpha* (Say) (Acanthocinini, Plate 32b, 5 mm). Adults of this eastern and central U. S. species are active from March–October. Larvae develop in the branches of numerous hardwoods, shrubs, and vines.
- Liopinus mimeticus (Casey) (Acanthocinini, Plate 32c, 5 mm). Adults of this common eastern and central U. S. species are active from April–October. Larvae feed in various hardwoods and shrubs, including Acer, Quercus, Carya, Lysiloma latisiliquum, Gleditsia triacanthos, Ebenopsis ebano, and Parkinsonia aculeata.
- *Liopinus misellus* (LeConte) (Acanthocinini, Plate 32a, 4 mm). Adults of this eastern species are active from April—August. Larvae feed in various hardwoods, including *Acer*, *Quercus* (including *Q. phellos* and *Q. velutina*), *Carya*, *Betula nigra*, and *Crataegus viridis*.
- Liopinus punctatus (Haldeman) (Acanthocinini, Plate 31k, 4 mm). Adults of this common eastern U. S. species are active from March–August. Larvae develop in various hardwoods including Celtis, Diospyros virginiana, Carpinus caroliniana, Carya cordiformis, C. illinoinensis, and Crataegus viridis.
- *Lypsimena fuscata* Haldeman (Pogonocherini, Plate 27e, 9 mm). Adults of this widespread but uncommon eastern U. S. and Caribbean species are active from April–May. Adults have been reared from *Quercus inopina*, *Q. agrifolia*, *Persea*, and *Prunus*.
- *Mecas cana cana* (Newman) (Phytoeciini, Plate 23a, 10 mm). This subspecies occurs only in Florida, and adults are active from April through August. Larval hosts include *Ambrosia* and *Flaveria linearis*.

- *Mecas cana saturnina* LeConte (Phytoeciini, Plate 23b, 12 mm). Adults of this common central and eastern U. S. subspecies are active from April–August. Larvae feed in the stems of living composites, especially *Ambrosia*, *Gaillardia*, and *Helianthus*.
- *Mecas cineracea* Casey (Phytoeciini, Plate 23c, 8 mm). Adults are active from April–August. Larvae feed in the stems of living composites. Adults have been collected sweeping many plants, including *Helenium*.
- *Mecas femoralis* (Haldeman) (Phytoeciini, Plate 22i, 6 mm). This rarely collected southeast U. S. species has been taken on *Aster* in the sandhill–scrub oak community of Florida. No larval hosts are known.
- *Mecas marginella* LeConte (Phytoeciini, Plate 23d, 8 mm). This is a widespread southern U. S. species. Larvae bore in many herbaceous plants. Adults are associated with many wildflowers from April to June.
- *Mecas pergrata* (Say) (Phytoeciini, Plate 22j, 8 mm). Adults are active from April–July. Larvae are stem and root borers of *Aster* and other composites. Adults can be collected on many wildflowers from April to June.
- *Microgoes oculatus* (LeConte) (Lamiini, Plate 24j, 9 mm). Adults are active from June–August. Larvae feed beneath the bark of numerous hardwoods as well as *Pinus*.
- *Monochamus carolinensis* (Olivier) (Lamiini, Plate 25a, 17 mm). Adults of this widespread U. S. species are active from April–September. Larvae feed in dead and dying *Pinus*.
- *Monochamus marmorator* **Kirby** (Lamiini, Plate 24k, 21 mm). Adults of this common U. S. species are active from June–September. Larvae feed in dead and dying *Abies* and *Picea*.
- *Monochamus notatus* (**Drury**) (Lamiini, Plate 24l, 19 mm). Adults are active from May–September. Larvae feed in dead and dying conifers, especially *Pinus*, but also *Abies*, *Picea*, and *Pseudotsuga*.
- **Monochamus scutellatus** (Say) (Lamiini, Plate 24h, 18 mm). Adults of this common and widespread U. S. species are active from April–September. Larvae feed in dead and dying conifers, especially *Pinus*, but also *Abies*, *Larix*, and *Picea*.
- *Monochamus titillator* (Fabricius) (Lamiini, Plate 25b, 24 mm). Adults of this widespread central and eastern U. S. and Bahamian species are active from March–October. Larvae feed in dead and dying *Pinus*, *Abies*, and *Picea*.
- Neoptychodes trilineatus (Linnaeus) (Lamiini, Plate 24i, 21 mm). This species is primarily Caribbean and Neotropical in distribution, but gets into the extreme southeastern U. S. Adults are active in the summer in the U. S. part of the range. Although *Ficus* is the primary host, larvae also develop in *Alnus*, *Morus*, *Salix* and *Celtis*.
- *Nyssodrysina haldemani* (LeConte) (Acanthocinini, Plate 28e, 7 mm). Adults of this widespread Neotropical and eastern U. S. species are active from April–November. Larvae feed in *Celtis*, *Ficus aurea*, *Bursera simaruba*, *Jatropha*, and *Forestiera segregata*.
- *Oberea affinis* Leng & Hamilton (Phytoeciini, Plate 231, 13 mm). Adults of this eastern species are active from June–July. Larvae are stem borers of living *Rubus*.
- *Oberea caseyi* Plavilstshikov (Phytoeciini, Plate 24a, 10 mm). Adults of this uncommon eastern species are active from May–August. Larvae develop in *Salix*.
- *Oberea delongi* Knull (Phytoeciini, Plate 23g, 10 mm). Adults of this widespread eastern and central U. S. and southeastern Canada species are active from May–August. Larvae feed in living *Populus*.
- *Oberea flavipes* **Haldeman** (Phytoeciini, Plate 23h, 10 mm). Adults of this generally uncommon species are active from May–July. Larvae are stem borers of living *Phlox*.
- **Oberea gracilis** (**Fabricius**) (Phytoeciini, Plate 23f, 13 mm). Adults of this widespread eastern U. S. species are active from April–August. Larvae develop in seedling oaks including *Quercus alba* and *Q. falcata*.
- **Oberea myops Haldeman** (Phytoeciini, Plate 24c, 16 mm). Adults of this eastern species are active from May–July. Larvae feed in the branches of living *Rhododendron*, *Vaccinium*, *Azalea*, and other genera.
- *Oberea ocellata* Haldeman (Phytoeciini, Plate 24d, 11 mm). Adults of this widespread eastern and central U. S. species are active from April–September. Larvae feed in the branches of living *Rhus*, *Malus*, *Prunus*, and other hardwoods. Adults are often collected on *Rhus*.

- *Oberea perspicillata* Haldeman (Phytoeciini, Plate 23m, 10 mm). Adults of this widespread eastern and central U. S. and southern Canada species are active from April–August. Larvae are stem borers of living *Rubus* and *Rosa*.
- *Oberea praelonga* Casey (Phytoeciini, Plate 23k, 10 mm). Adults are active from May–July. Larvae feed in *Cornus* and *Viburnum*. Note that this species treatment includes *O. deficiens* Casey which is morphologically identical and a potential synonym of *O. praelonga* Casey.
- *Oberea ruficollis* (Fabricius) (Phytoeciini, Plate 23e, 15 mm). Adults of this widespread eastern U. S. species are active from May–August. Larvae bore in stems and roots of living *Sassafras* and *Lindera*.
- Oberea schaumii LeConte (Phytoeciini, Plate 24b, 12 mm). Adults of this widespread eastern North American species are active from May–July. Larvae are girdlers of the living branches of *Populus*. Note that this species treatment includes *O. pruinosa* Casey which is morphologically identical and a potential synonym of *O. schaumii*.
- Oberea tripunctata (Swederus) (Phytoeciini, Plate 23j, 10 mm). Adults of this widespread eastern and central North American species are active from May–August. Larvae bore in living branches of Cornus, Prunus, Viburnum, Ulmus, Oxydendrum arboreum, Vaccinium, Rhododendron, Populus, Salix, and Morus.
- *Oberea ulmicola* Chittenden (Phytoeciini, Plate 23i, 9 mm). Adults of this uncommon eastern U. S. species are active from May–July. Larvae feed in *Ulmus* and *Prunus*.
- Oncideres cingulata (Say) (Onciderini, Plate 27e, 16 mm). Adults of this common U. S. species are active from April–November. Larvae girdle branches of numerous hardwoods and shrubs, especially *Diospyros virginiana*, Carya, Juglans, Celtis, and Ulmus, but also occasionally Quercus and Prunus.
- *Oplosia nubila* (LeConte) (Acanthoderini, Plate 30i, 10 mm). Adults of this generally uncommon eastern U. S. species are active from May–July. Larvae feed under bark of *Tilia*, *Carya*, and *Fagus*.
- **Parmenonta thomasi** Chemsak & Linsley (Apomecynini, Plate 26g, 7 mm). This rare species is endemic to Florida. Adults have been collected in June and December. Nothing is known of potential host plants.
- *Phaea monostigma* (Haldeman) (Tetraopini, Plate 22b, 9 mm). Adults of this uncommon central and eastern U. S. species are active from May–July. Larvae feed in the living stems of *Ipomoea*.
- *Plectrodera scalator* (Fabricius) (Lamiini, Plate 24f, 30 mm). Adults of this widespread central and southern U. S. species are active from May–September. Larvae develop in *Populus* and *Salix*.
- **Pogonocherus mixtus Haldeman** (Pogonocherini, Plate 29k, 5 mm). Adults of this moderately common North American species are active from May–September. Larvae feed in conifers (especially *Larix*, *Picea*, and *Pinus*) and hardwoods including *Salix* and *Pyrus*
- **Pogonocherus penicillatus LeConte** (Pogonocherini, Plate 29j, 5 mm). Adults of this northern U. S. species are active from July–August. Larvae develop in *Picea*.
- **Psenocerus supernotatus** (Say) (Desmiphorini, Plate 27a, 4 mm). Adults of this common central and eastern U. S. species are active from April–June. Larval hosts include *Liriodendron tulipitera*, Crataegus viridis, and Salix nigra. Adults have also been taken on Acer.
- Saperda candida Fabricius (Saperdini, Plate 27j, 17 mm). Adults of this eastern North American species are active from May–August. Larvae feed in living Malus, Cydonia oblonga, Crataegus, and many related genera, mostly in the family Rosaceae.
- Saperda calcarata Say (Saperdini, Plate 28b, 25 mm). Adults of this widespread central and eastern North American species are active from May–September. Larvae feed in *Populus* (especially *Populus tremuloides* and *Populus deltoides*).
- Saperda cretata Newman (Saperdini, Plate 27k, 14 mm). Adults of this widespread eastern and central U. S. species are active from May–August. Larvae develop in Malus and Crataegus.
- Saperda discoidea Fabricius (Saperdini, Plate 27g, 11 mm). Adults of this widespread eastern and central U. S. and southeast Canada species are active from April–September. Larvae feed in Carya, Juglans cinerea, J. nigra, and occasionally other hardwoods.
- Saperda imitans Felt & Joutel (Saperdini, Plate 27m, 11 mm). Adults of this rare eastern U. S. species are active from May–July. Larvae feed in various dead hardwoods including Carya, Prunus, and Salix.

- Saperda inornata Say (Saperdini, Plate 27h, 10 mm). Adults of this northern U. S. and southern Canada species are active from March–July. Larvae feed in living *Populus* and *Salix*, making galls in the smaller branches and saplings.
- Saperda lateralis Fabricius (Saperdini, Plate 27l, 10 mm). Adults of this widespread eastern and central U. S. species are active from May–August. Larvae feed in various dead hardwoods and shrubs, including Carya, Ulmus, Tilia, Fraxinus, Quercus, Acer, and Toxicodendron. Pinus is a questionable host that has been mentioned in the literature.
- *Saperda mutica* Say (Saperdini, Plate 28c, 11 mm). Adults of this central and eastern U. S. species are active from June–July. Larvae feed in dead *Salix*.
- Saperda obliqua Say (Saperdini, Plate 28d, 15 mm). Adults of this widespread eastern Canada and U. S. species are active from June–August. Larvae feed in living Alnus, Betula, and Corylopsis.
- Saperda populnea moesta LeConte (Saperdini, Plate 27f, 8 mm). Adults of this northern U. S. and southern Canada species are active from June–August. Larvae feed in the living branches of *Populus* and *Salix*.
- Saperda puncticollis Say (Saperdini, Plate 28e, 9 mm). Adults of this eastern North American species are active from May–August. Larvae feed in dead and dying *Parthenocissus quinquefolia*, *Toxicodendron radicans*, and *Vitis*.
- Saperda tridentata Olivier (Saperdini, Plate 27n, 11 mm). Adults of this widespread eastern and central U. S. species are active from April–October. Larvae feed in *Ulmus*.
- Saperda vestita Say (Saperdini, Plate 27i, 15 mm). Adults of this widespread eastern and central U. S. and Canada species are active from May–September. Larvae develop in *Acer*, *Tilia*, and occasionally *Populus*.
- *Spalacopsis chemsaki* Tyson (Agapanthiini, Plate 25j, 6 mm). This rare species has been collected in June from only one locality in southern Florida. No host plants are known.
- Spalacopsis filum costulatum Casey (Agapanthiini, Plate 25k, 7 mm). Adults of this Florida and Caribbean species can be collected in most months, especially June–July. Larval hosts include *Melothria*, but adults have been taken on fresh cut *Ipomoea*, *Cojoba arborea*, and *Bursera simaruba*.
- *Spalacopsis stolata* Newman (Agapanthiini, Plate 25l, 6 mm). Specimens of this uncommon Florida species have been taken on *Chenopodium botrys*, *Verbesina*, *Flaveria linearis*, *Batis maritima*, and *Bursera simaruba* from April through June.
- *Spalacopsis suffusa* **Newman** (Agapanthiini, no figure). This rare species has been collected from March through October in southern Florida. No hosts are known.
- Steirastoma breve (Sulzer) (Acanthoderini, Plate 30e, 23 mm). This species is widely distributed throughout the Neotropics and has been collected in southern Texas and Florida. It is polyphagous with hosts including Cocos nucifera, Adansonia digitata, Bombax ceiba, Cecropia, Hibiscus, Wisteria, Salix, and others.
- Sternidius variegatus (Haldeman) (Acanthocinini, Plate 311, 8 mm). Adults of this common central and eastern U. S. species are active from June–September. Larvae feed in the branches of *Gymnocladus dioicus* and *Aesculus pavia*, among many other hosts.
- Styloleptus biustus (LeConte) (Acanthocinini, Plate 29c, 5 mm). Adults of this southeastern U. S. and Caribbean species are active from May–July. Larvae feed in numerous hardwoods and shrubs, including *Carya*. The *minuens* form (currently a morphologically distinct subspecies of *S. biustus*) may deserve species status (C. J. Micheli, pers. comm.).
- Sybra alternans Wiedemann (Apomecynini, Plate 26i, 8 mm). This widespread Asian and Pacific Island species was discovered in Florida in 1992. Larvae develop in *Ficus* and *Musa*.
- **Tetraopes melanurus Schoenherr** (Tetraopini, Plate 22d, 9 mm). Adults of this southern U. S. species are active from May–September. Larvae feed in *Asclepias tuberosa*.
- **Tetraopes pilosus Chemsak** (Tetraopini, Plate 22c, 10 mm). Adults of this uncommon central U. S. species are active from June–August. Larvae feed in *Asclepias tuberosa* and *A. arenaria*.
- **Tetraopes quinquemaculatus Haldeman** (Tetraopini, Plate 22f, 9 mm). Adults of this uncommon U. S. species are active from June–September. Larvae feed in *Asclepias hirtella* and adults have been associated with other species such as *A. tuberosa* and *A. amplexicaulis*.

- **Tetraopes tetrophthalmus** (**Forster**) (Tetraopini, Plate 22g, 12 mm). Adults of this common U. S. species are active from May–September. Larvae feed in various milkweeds (especially *Asclepias syriaca*) and *Apocynum*.
- **Tetraopes texanus Horn** (Tetraopini, Plate 22e, 12 mm). Adults of this uncommon central U. S. species are active from April–August. Larvae feed in *Asclepias hirtella*.
- *Tetrops praeusta* Linnaeus (Tetraopini, Plate 22a, 4 mm). This European species was introduced into the United States. Adults are active from May–July. Larvae feed in various hardwoods and shrubs, especially *Malus* and *Crataegus*.
- *Urgleptes facetus* (Say) (Acanthocinini, Plate 32d, 4 mm). Adults of this common U. S. species are active from May–August. Larvae feed in the branches of various hardwoods, especially *Quercus*, *Crataegus*, *Maclura*, and *Salix*. Adults have also been collected on *Pinus sylvestris*.
- *Urgleptes foveatocollis* (Hamilton) (Acanthocinini, Plate 32f, 4 mm). Adults of this uncommon southern U. S. species are active from July–September. Larval hosts include *Celtis*, *Lantana*, *Lysiloma*, *Piscidia*, and *Avicennia germinans*.
- *Urgleptes querci* (Fitch) (Acanthocinini, Plate 32e, 4 mm). Adults of this common eastern U. S. species are active from May–September. Larvae feed in the branches of numerous hardwoods, shrubs, and vines, especially *Acer* (including *Acer negundo*), *Aesculus glabra*, *Betula nigra*, *Quercus*, *Prunus serotina*, *Salix exigua*, *Carya*, *Morus*, *Vaccinium*, *Viburnum*, *Toxicodendron*, and others.
- *Urgleptes signatus* (LeConte) (Acanthocinini, Plate 32g, 4 mm). Adults of this eastern U. S. species are active from June–August. Larvae feed in the branches of various genera including *Acer*, *Carpinus caroliniana*, *Carya*, *Cornus*, *Quercus*, and *Tilia*.
- *Urographis despectus* (LeConte) (Acanthocinini, Plate 31b, 10 mm). Adults of this uncommon species are active from May–July. Larvae feed in various hardwoods, especially *Carya*.
- Urographis fasciatus (DeGeer) (Acanthocinini, Plate 31c, 11 mm). Adults of this common eastern U. S. species are active from April—October. Larvae feed in numerous hardwoods including Ouercus, and also Pinus.
- *Urographis triangulifer* (Haldeman) (Acanthocinini, Plate 30k, 15 mm). Adults of this widespread but generally uncommon central and eastern U. S. species are active from May–October. Larvae feed in *Celtis* and *Acer*.
- **Zaplous annulatus** (Chevrolat) (Pogonocherini, Plate 27b, 4 mm). Adults of this uncommon Florida and Cuba species have been collected in June from *Ilex*.

Literature and Websites Cited

- **Bond, W. B. & T. K. Philips. 1999.** Diversity, phenology, and flower hosts of anthophilous long–horned beetles (Coleoptera: Cerambycidae) in a southeastern Ohio forest. Entomological News 110(5):267–278.
- **Browne, D. J. & S. B. Peck. 1996.** The long-horned beetles of south Florida (Cerambycidae: Coleoptera): biogeography and relationships with the Bahama Islands and Cuba. Canadian Journal of Zoology 74:2154–2169.
- **Browne, D. J., S. B. Peck, & M. A. Ivie. 1993.** The Longhorn beetles (Coleoptera Cerambycidae) of the Bahama Islands with an analysis of species—area relationships, distribution patterns, origin of the fauna and an annotated species list. Tropical Zoology 6:27–53.
- Cavey, J. F., E. R. Hoebeke, S. Passoa, & S. W. Lingafelter. 1998. A new exotic threat to North American hardwood forests: An Asian longhorned beetle, *Anoplophora glabripennis* (Coleoptera: Cerambycidae). I. Description and diagnosis of the larva. Proceedings of the Entomological Society of Washington, 100(2):373–381.
- **Giesbert, E. F. 1993.** A new genus and species of clytine cerambycid (Coleoptera) from Florida. Insecta Mundi 7(3):129–131.
- **Gressitt, J. L. 1942.** Destructive Long-horned beetle borers at Canton, China. Special Publication No. 1 of Lingnan Natural History Survey and Museum, Lingnan University, Canton, China: 60pp.
- **Hanley, G. A. 2005.** Cerambycidae of North Dakota. An atlas and identification guide. Cyril Moore Science Center Science Monograph #3, Minot State University, Minot, North Dakota: 105pp.
- **Hoffman, R. L. 2003.** Beetles of the genus *Anthophylax* in Virginia (Coleoptera: Cerambycidae: Lepturinae). Banisteria 22:50–52.
- Hoffman, R. L., S. M. Roble, & W. E. Steiner, Jr. 2002. Thirteen additions to the known beetle fauna of Virginia (Coleoptera: Scirtidae, Bothrideridae, Cleridae, Tenebrionidae, Melyridae, Callirhipidae, Cerambycidae, Chrysomelidae). Banisteria 20:53–61.
- **Hovore, F. T., R. L. Penrose, & R. W. Neck. 1987.** The Cerambycidae or longhorned beetles of Southern Texas: a faunal survey. Proceedings of the California Academy of Science 44(13):283–334.
- Korotyaev, B. A., A. S. Konstantinov, S. W. Lingafelter, M. Y. Mandelshtam, and M. G. Volkovitsh. 2005. Gall–Inducing Coleoptera, pp. 239–271, *in:* A. Raman, C. W. Schaefer, and T. M. Withers, eds, *Biology, Ecology, and Evolution of Gall–inducing Arthropods*. Science Publishers, Inc., Enfield, NH.
- **Lingafelter, S. W. & J. A. Chemsak. 2002.** A new species of *Enaphalodes* Haldeman from Florida (Coleoptera: Cerambycidae) with review of genus, synonymies, and key to species. Coleopterists Bulletin 56(4):569–581.
- **Lingafelter, S. W. & E. H. Nearns. 2006.** Cerambycidae holotypes of the Smithsonian Institution: an online image database. [www.elaphidion.com].
- **Lingafelter, S. W. & E. R. Hoebeke. 2002.** Revision of *Anoplophora* (Coleoptera: Cerambycidae). Entomological Society of Washington, Washington, DC. 236pp.
- **Lingafelter, S. W. & N. V. Horner. 1993.** The Cerambycidae of north-central Texas. Coleopterists Bulletin 47(2):159–191.
- **Lingafelter, S. W. & M. A. Ivie. 2005.** Synonymies and transfers in Elaphidiini mostly relating to the genus *Elaphidion* Audinet-Serville (Coleoptera: Cerambycidae). Journal of the New York Entomological Society 112(2–3):205–211.
- **Linsley, E. G. 1962a.** The Cerambycidae of North America. Part II. Taxonomy and classification of the Parandrinae, Prioninae, Spondylinae and Aseminae. University of California Publications in Entomology, 19:1–102.
- **Linsley, E. G. 1962b.** The Cerambycidae of North America. Part III. Taxonomy and classification of the subfamily Cerambycinae, tribes Opsimini through Megaderini. University of California Publications in Entomology 20:1–188.

- **Linsley, E. G. 1963.** The Cerambycidae of North America. Part IV. Taxonomy and classification of the subfamily Cerambycinae, tribes Elaphidionini through Rhinotragini. University of California Publications in Entomology 21:1–165.
- **Linsley, E. G. 1964.** The Cerambycidae of North America. Part V. Taxonomy and classification of the subfamily Cerambycinae, tribes Callichromini through Ancylocerini. University of California Publications in Entomology 22:1–197 pp.
- **Linsley, E. G., & J. A. Chemsak. 1972.** The Cerambycidae of North America. Part VI, No. 1. Taxonomy and classification of the subfamily Lepturinae. University of California Publications in Entomology 69:1–138.
- **Linsley, E. G., & J. A. Chemsak. 1976.** The Cerambycidae of North America. Part VI, No. 2. Taxonomy and classification of the subfamily Lepturinae. University of California Publications in Entomology 80:1–186.
- **Linsley, E. G., & J. A. Chemsak. 1984.** The Cerambycidae of North America. Part VII, No. 1: Taxonomy and classification of the subfamily Lamiinae, Tribes Parmenini through Acanthoderini. University of California Publications in Entomology 102:1–258.
- **Linsley, E. G., & J. A. Chemsak. 1995.** The Cerambycidae of North America. Part VII, No. 2. Taxonomy and Classification of the Subfamily Lamiinae, Tribes Acanthocinini through Hemilophini. University of California Publications in Entomology 114:1–292.
- **Linsley, E. G., & J. A. Chemsak. 1997.** The Cerambycidae of North America. Part VIII: Bibliography, Index, and Host Plant Index. University of California Publications in Entomology 117:1–534.
- **MacRae, T. C. 1993.** Annotated checklist of the longhorned beetles (Coleoptera: Cerambycidae and Disteniidae) occurring in Missouri. Insecta Mundi 7(4):223–252.
- MacRae, T. C. & M. E. Rice. 2007. Biological and distributional observations on North American Cerambycidae (Coleoptera). Coleopterists Bulletin 61(2).
- Maier, C. T. & C. R. Lemmon. 2000. Discovery of the Small Japanese Cedar Longhorned Beetle, *Callidiellum rufipenne* (Motschulsky) (Coleoptera: Cerambycidae), in live arborvitae in Connecticut. Proceedings of the Entomological Society of Washington 102(3):747–754.
- **Monné, M. A. 2001a.** Catalogue of the Neotropical Cerambycidae with known host plant Part 1: Subfamily Cerambycinae, tribes Achrysonini to Elaphidiini. Publicações Avulsas do Museu Nacional 88:1–108.
- **Monné, M. A. 2001b.** Catalogue of the Neotropical Cerambycidae with known host plant Part II: Subfamily Cerambycinae, tribes Graciliini to Trachyderini. Publicações Avulsas do Museu Nacional 90:1–119.
- **Monné, M. A. 2001c.** Catalogue of the Neotropical Cerambycidae with known host plant Part III: Subfamily Lamiinae, tribes Acanthocinini to Apomecynini. Publicações Avulsas do Museu Nacional 92:1–94.
- **Monné, M. A. 2002a.** Catalogue of the Neotropical Cerambycidae with known host plant Part IV: Subfamily Lamiinae, tribes Batocerini to Xenofreini. Publicações Avulsas do Museu Nacional 94:1–92.
- **Monné, M. A. 2002b.** Catalogue of the Neotropical Cerambycidae with known host plant Part V: Subfamilies Prioninae, Parandrinae, Oxypeltinae, Anoplodermatinae, Aseminae and Lepturinae. Publicações Avulsas do Museu Nacional 96:1–72.
- **Monné**, **M. A. 2005a.** Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part I. Subfamily Cerambycinae. Zootaxa 946:1–765.
- **Monné**, **M. A. 2005b.** Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part II. Subfamily Lamiinae. Zootaxa 1023:1–759.
- Monné, M. A. 2006. Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part III. Subfamilies Parandrinae, Prioninae, Anoplodermatinae, Aseminae, Spondylidinae, Lepturinae, Oxypeltinae, and addenda to the Cerambycinae and Lamiinae. Zootaxa 1212:1–244.
- Monné, M. A. and F. T. Hovore. 2005. Checklist of the Cerambycidae, or longhorned wood-boring beetles (Coleoptera), of the Western Hemisphere. BioQuip Products, Rancho Dominguez, CA. 392 pp.

- **Morris, R. F. II. 2002.** Distribution and biological notes for some Cerambycidae (Coleoptera) occurring in the southeastern United States. Insecta Mundi 16(4):209–213.
- **Perkins, P., P. Naskrecki, & B. Farrell. 2006.** Online database of insect primary types in the collection of the Museum of Comparative Zoology at Harvard University. [http://mcz-28168.oeb. harvard.edu/mcz/index.htm].
- **Rice, M. E. and D. A. Veal. 2006.** New distribution and adult host records for longhorned beetles (Cerambycidae) from Iowa. Coleopterists Bulletin 60(3):255–263.
- **Schiefer, T. L. 1998.** A preliminary list of the Cerambycidae and Disteniidae (Coleoptera) of Mississippi. Transactions of the American Entomological Society 124(2):113–131.
- **Schiefer, T. L. 2000.** A new species of Astylopsis Casey (Coleoptera: Cerambycidae: Acanthocinini) from the southeastern United States. Coleopterists Bulletin 54(4):533–539.
- **Schiefer, T. L. 2001.** Additions and corrections to the list of Cerambycidae (Coleoptera) of Mississippi. Entomological News 112(5):334–336.
- Thomas, M. C., S. Hill, R. F. Morris II, & E. H. Nearns. 2006. The Cerambycidae of Florida. Florida State Collection of Arthropods website. [http://www.fsca-dpi.org/Coleoptera/Mike/FloridaCerambycids/openingpage.htm].
- **USDA NRCS. 2007.** The PLANTS Database. National Plant Data Center, Baton Rouge, LA. [http://plants.usda.gov].
- **Vogt, G. B. 1949.** Notes on Cerambycidae from the lower Rio Grande Valley, Texas. The Pan-Pacific Entomologist 25(3):137-184.
- **Yanega**, **D. 1996.** Field Guide to Northeastern Longhorned Beetles (Coleoptera: Cerambycidae). Illinois Natural History Survey Manual 6, Champaign, Illinois. 184pp.

Plates



Plate 1.

a) Distenia undata (Fabricius), b) Scaphinus muticus (Fabricius), c) Hesperandra polita (Say), d) Neandra brunnea (Fabricius), e) Sphenostethus taslei (Buquet), f) Elateropsis rugosus Gahan, g) Elateropsis scabrosus Gahan, h) Tragosoma depsarius (Linnaeus), i) Strongylaspis corticarius (Erichson).

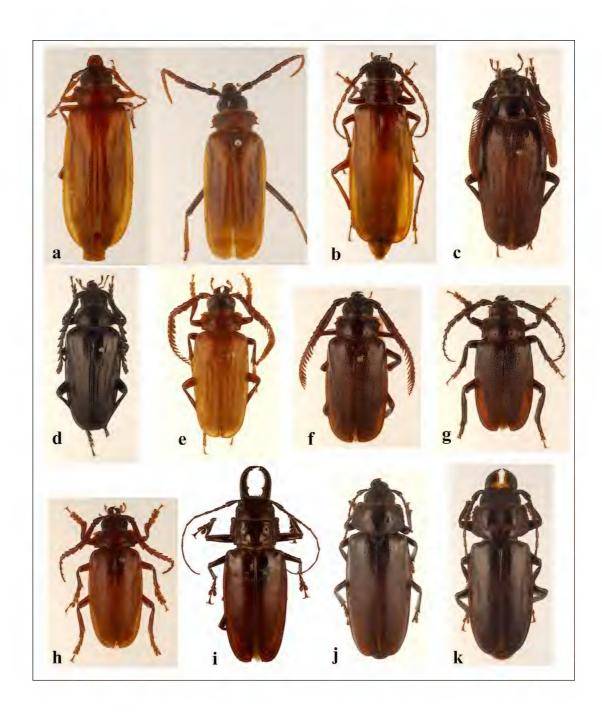


Plate 2.

a) Derobrachus brevicollis Audinet—Serville, b) Orthosoma brunneum (Forster), c) Prionus fissicornis Haldeman, d) Prionus palparis Say, e) Prionus debilis Casey, f) Prionus imbricornis (Linnaeus), g) Prionus laticollis (Drury), h) Prionus pocularis Dalman, i) Stenodontes chevrolati Gahan, j) Archodontes melanopus (Linnaeus), k) Mallodon dasystomus (Say).

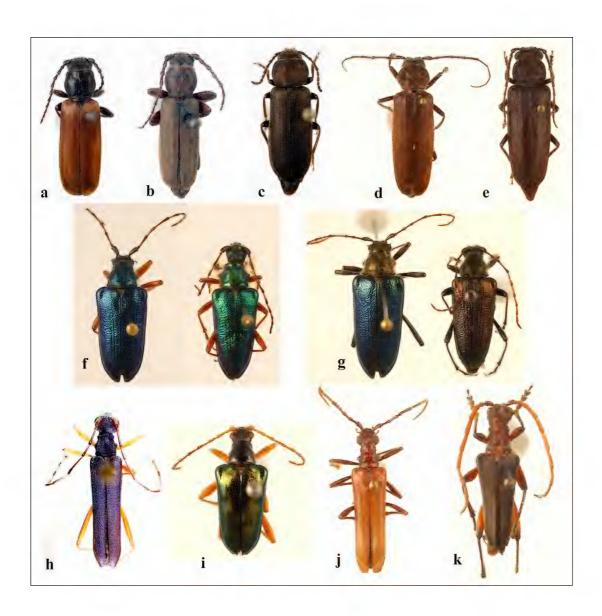


Plate 3.

a) Tetropium cinnamopterum Kirby, b) Tetropium schwarzianum Casey, c) Asemum striatum (Linnaeus), d) Arhopalus foveicollis (Haldeman), e) Arhopalus rusticus (LeConte), f) Anthophylax cyaneus (Haldeman), g) Anthophylax viridis LeConte, h) Encyclops caerulea (Say), i) Gaurotes cyanipennis (Say), j) Centrodera decolorata (Harris), k) Stenocorus cylindricollis (Say).

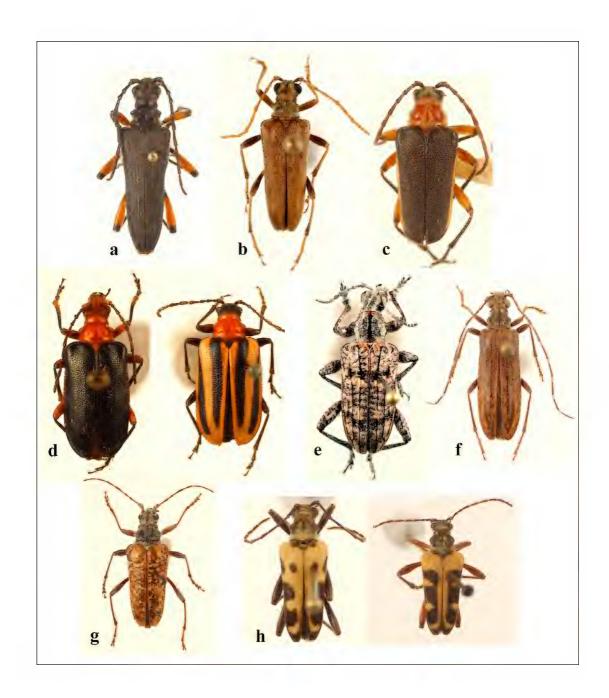


Plate 4.

a) Stenocorus schaumii (LeConte), **b)** Stenocorus cinnamopterus (Randall), **c)** Gaurotes thoracica (Haldeman), **d)** Brachysomida bivittata (Say), **e)** Rhagium inquisitor (Linnaeus), **f)** Centrodera sublineata LeConte, **g)** Anthophylax attenuatus (Haldeman), **h)** Evodinus monticola (Randall).

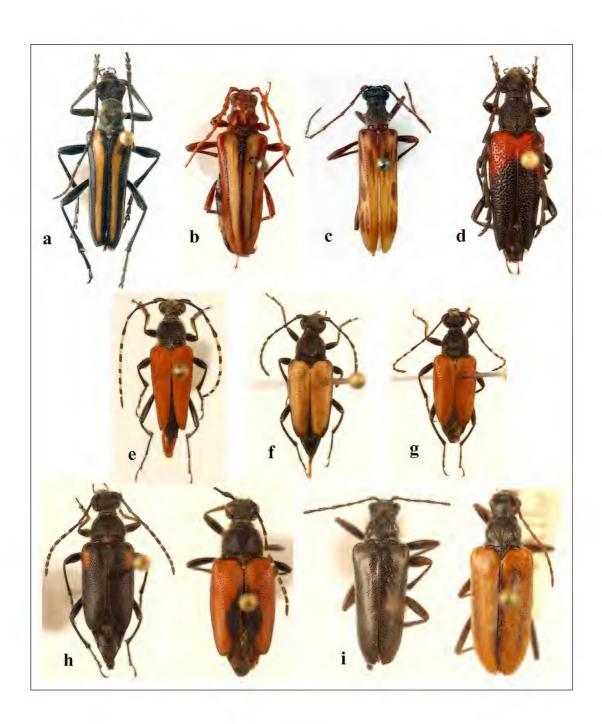


Plate 5.

a) Stenocorus vittiger (Randall),
b) Stenocorus trivittatus (Say),
c) Leptorhabdium pictum (Haldeman),
d) Stictoleptura canadensis (Olivier),
e) Brachyleptura rubrica (Say),
f) Brachyleptura circumdata (Olivier),
g) Brachyleptura champlaini Casey,
h) Brachyleptura vagans (Olivier),
i) Acmaeops proteus (Kirby).

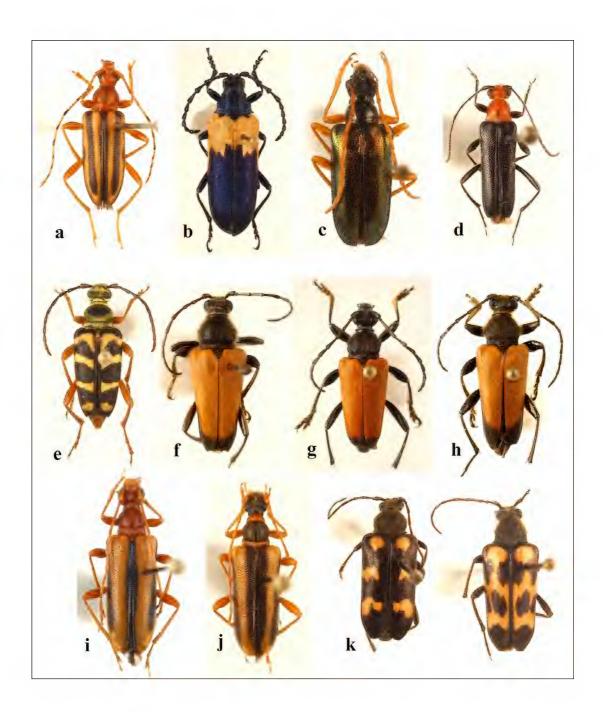


Plate 6.

a) Metacmaeops vittata (Swederus), b) Desmocerus palliatus (Forster), c) Pseudogaurotina abdominalis (Bland), d) Neoalosterna capitata (Newman), e) Strophiona nitens (Forster), f) Trigonarthris minnesotana (Casey), g) Trigonarthris atrata (LeConte), h) Trigonarthris proxima (Say), i) Pidonia aurata (Horn), j) Pidonia densicollis (Casey), k) Judolia montivigans (Couper).

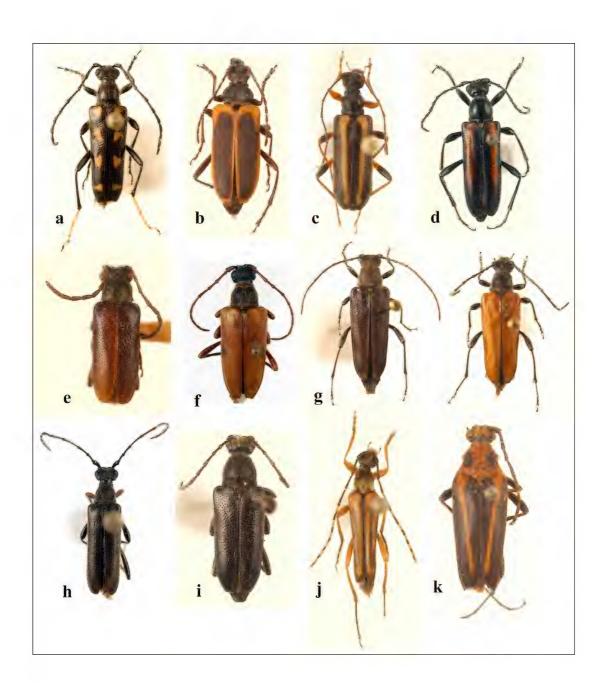


Plate 7.

a) Xestoleptura octonotata (Say),
b) Acmaeops discoideus (Haldeman),
c) Pidonia ruficollis (Say),
d) Strangalepta abbreviata (Germar),
e) Alosternida chalybaea (Haldeman),
f) Lepturopsis biforis (Newman),
g) Trachysida mutabilis (Newman),
h) Idiopidonia pedalis (LeConte),
i) Anoplodera pubera (Say),
j) Analeptura lineola (Say),
k) Leptura abdominalis (Haldeman).

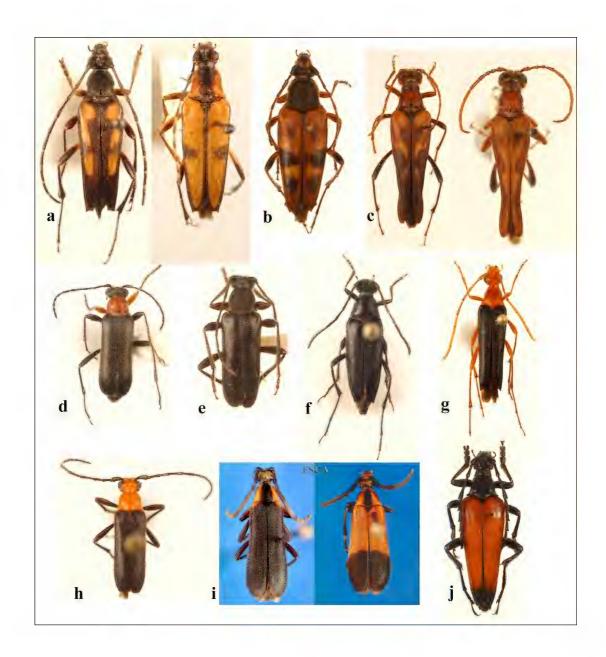


Plate 8.

a) Leptura subhamata Randall, b) Leptura obliterata deleta (LeConte), c) Bellamira scalaris (Say), d) Grammoptera haematites (Newman), e) Grammoptera subargentata (Kirby), f) Typocerus lugubris (Say), g) Strangalia bicolor (Swederus), h) Charisalia americana (Haldeman), i) Lycochoriolaus lateralis (Olivier), j) Stenelytrana emarginata (Fabricius).

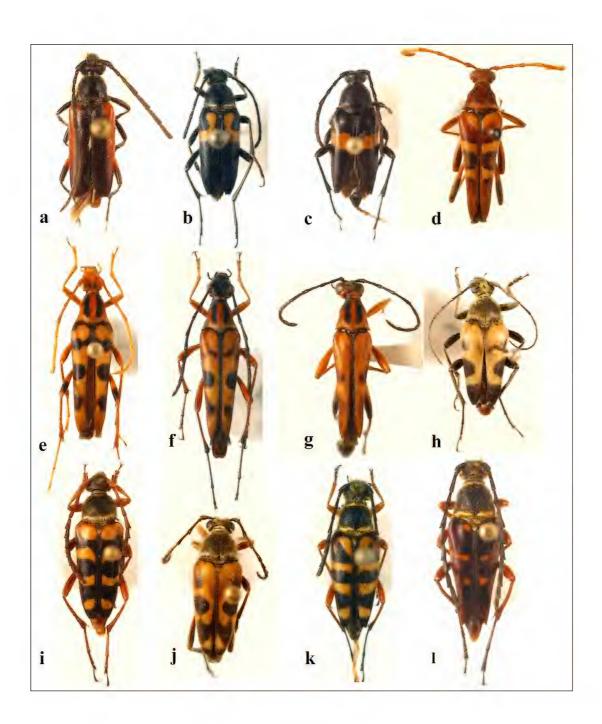


Plate 9.

a) Pseudostrangalia cruentata (Haldeman), b) Typocerus lunulatus (Swederus), c) Typocerus fulvocinctus Knull, d) Strangalia strigosa Newman, e) Strangalia luteicornis (Fabricius), f) Strangalia famelica famelica Newman, g) Strangalia famelica solitaria Haldeman, h) Judolia cordifera (Olivier), i) Typocerus sinuatus (Newman), j) Typocerus octonotatus (Haldeman), k) Typocerus zebra (Olivier), l) Typocerus badius (Newman).

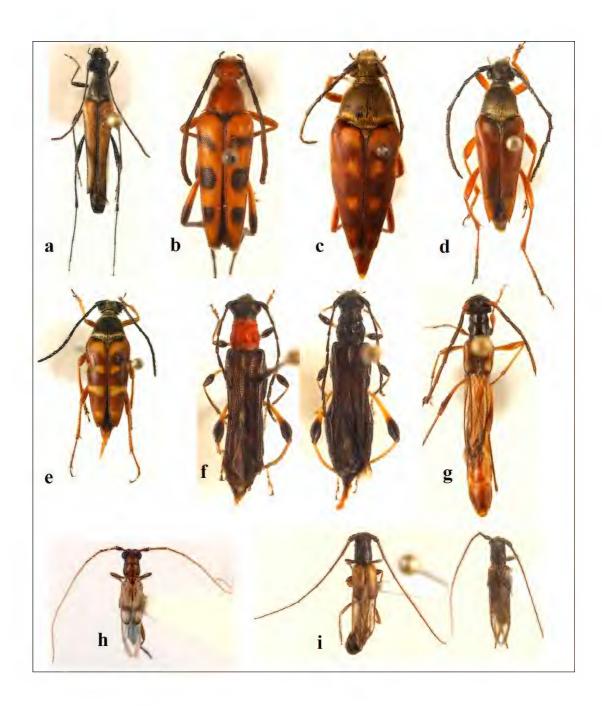


Plate 10.

a) Strangalia acuminata (Olivier), b) Strangalia sexnotata Haldeman, c) Typocerus deceptus Knull, d)
Typocerus acuticauda Casey, e) Typocerus velutinus (Olivier), f) Callimoxys sanguinicollis (Olivier),
g) Necydalis mellita (Say), h) Methia necydalea (Fabricius), i) Tessaropa tenuipes (Haldeman).

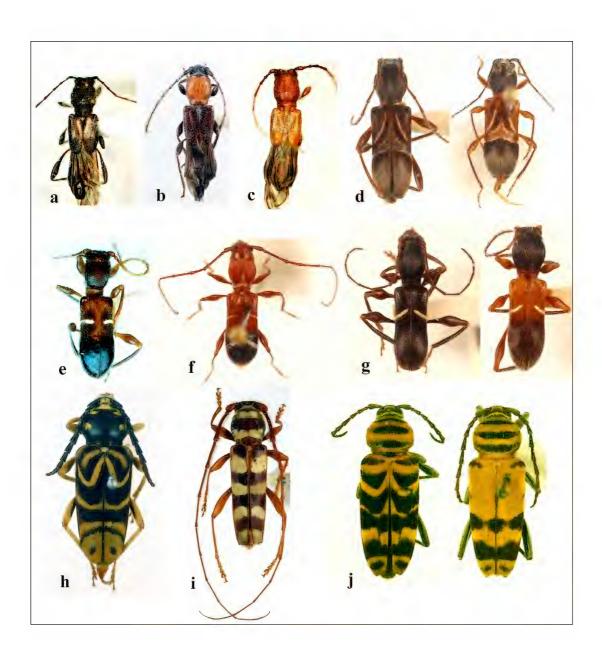


Plate 11.

a) Molorchus bimaculatus bimaculatus Say, b) Molorchus bimaculatus corni Haldeman, c) Molorchus bimaculatus semiustus (Newman), d) Cyrtophorus verrucosus (Olivier), e) Euderces reichei reichei LeConte, f) Euderces pini (Olivier), g) Euderces picipes (Fabricius), h) Glycobius speciosus (Say), i) Dryobius sexnotatus Linsley, j) Megacyllene decora (Olivier).

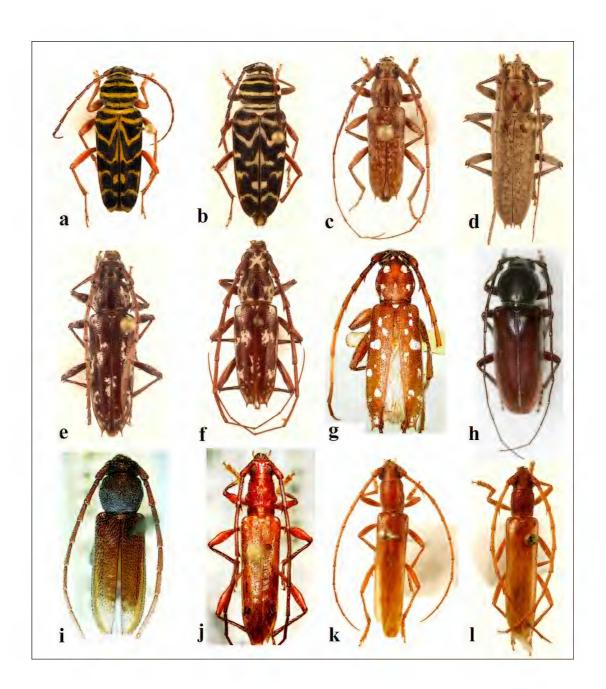


Plate 12.

- $a) \ \textit{Megacyllene robiniae} \ (Forster), \\ b) \ \textit{Megacyllene caryae} \ (Gahan), \\ c) \ \textit{Elaphidion mucronatum} \ (Say),$
- d) Elaphidion tectum LeConte, e) Elaphidion cryptum Linsley, f) Elaphidion irroratum (Linnaeus),
- g) Linsleyonides albomaculatus (Champlain & Knull), h) Romulus globosus Knull, i) Anelaphus moestus (LeConte), j) Stizocera floridana Linsley, k) Aneflomorpha delongi (Champlain & Knull), l) Aneflomorpha subpubescens (LeConte).

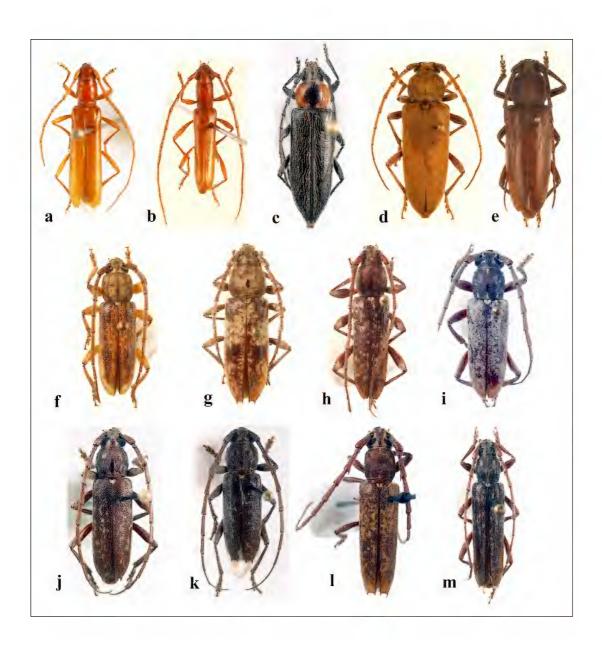


Plate 13.

a) Psyrassa unicolor (Randall), b) Psyrassa pertenuis (Casey), c) Stenosphenus notatus (Olivier), d) Enaphalodes archboldi Lingafelter & Chemsak, e) Enaphalodes hispicornis (Linnaeus), f) Anelaphus cinereus (Olivier), g) Anelaphus mutatum (Gahan), h) Anelaphus pumilus (Newman), i) Anelaphus inermis (Newman), j) Parelaphidion incertum (Newman), k) Parelaphidion aspersum (Haldeman), l) Anelaphus villosus (Fabricius), m) Anelaphus parallelus (Newman).

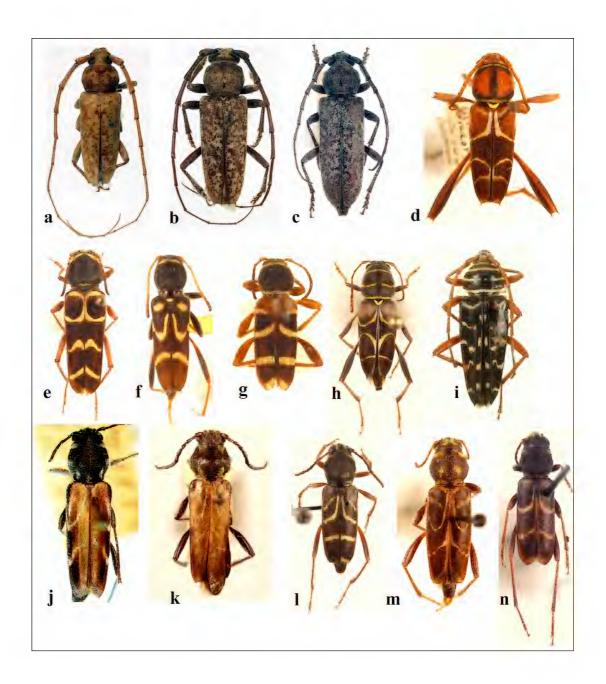


Plate 14.

a) Enaphalodes rufulus (Haldeman), b) Enaphalodes atomarius (Drury), c) Enaphalodes cortiphagus (Craighead), d) Neoclytus mucronatus (Fabricius), e) Neoclytus caprea (Say), f) Clytus ruricola (Olivier), g) Clytus marginicollis Castelnau & Gory, h) Neoclytus scutellaris (Olivier), i) Placosternus difficilis (Chevrolat), j) Xylotrechus aceris Fisher, k) Xylotrechus quadrimaculatus (Haldeman), l) Xylotrechus nitidus (Horn), m) Xylotrechus convergens LeConte, n) Xylotrechus schaefferi Schott.

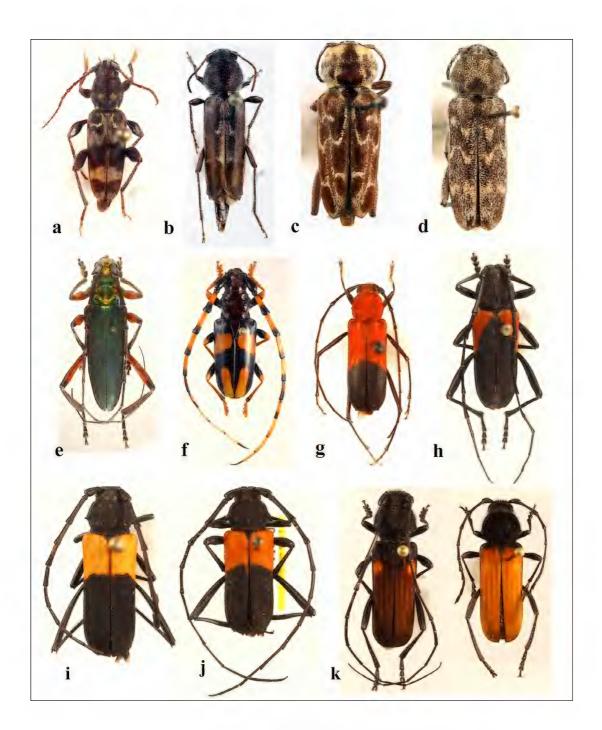


Plate 15.

a) Xylotrechus colonus (Fabricius), b) Xylotrechus sagittatus (Germar), c) Xylotrechus integer (Haldeman), d) Xylotrechus annosus annosus (Say), e) Plinthocoelium suaveolens suaveolens (Linnaeus), f) Trachyderes mandibularis Dupont, g) Heterops dimidiatus (Chevrolat), h) Purpuricenus humeralis (Fabricius), i) Purpuricenus axillaris Haldeman, j) Purpuricenus paraxillaris MacRae, k) Tragidion coquus (Linnaeus).

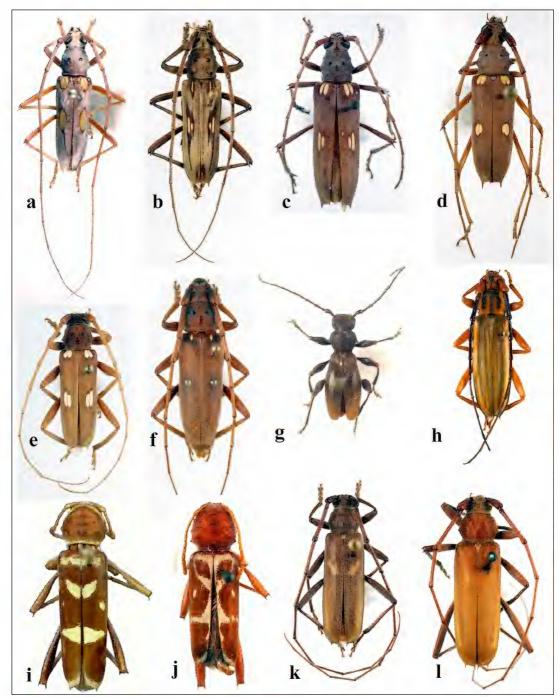


Plate 16.

a) Eburia cinereopilosa Fisher, b) Eburia stigma (Olivier), c) Eburia distincta Haldeman, d) Eburia stroheckeri Knull, e) Eburia quadrigeminata (Say), f) Eburia haldemani LeConte, g) Michthisoma heterodoxum LeConte, h) Chlorida festiva (Linnaeus), i) Neoclytus longipes (Drury), j) Neoclytus cordifer (Klug), k) Knulliana cincta cincta (Drury), l) Knulliana cincta spinifera (Fabricius).

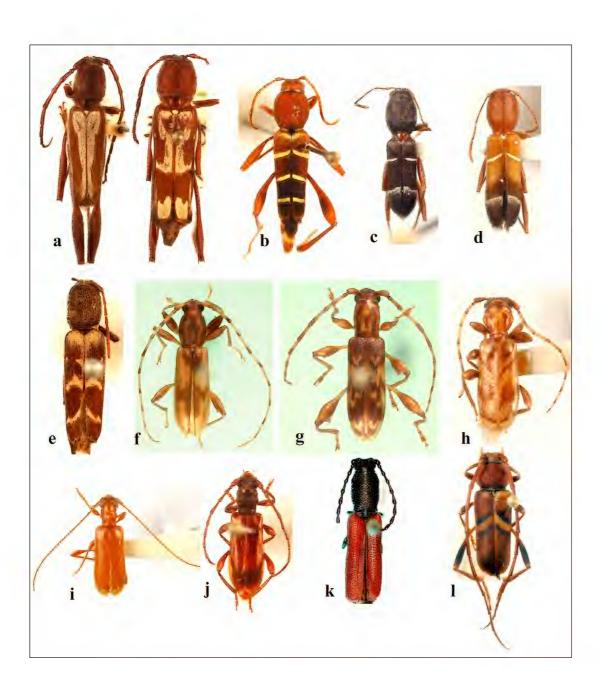


Plate 17.

- a) Euryscelis suturalis (Olivier), b) Neoclytus acuminatus (Fabricius), c) Neoclytus jouteli jouteli Davis,
- d) Neoclytus jouteli simplarius Blatchley, e) Neoclytus horridus (LeConte), f) Curius dentatus Newman,
- **g**) Plectromerus dentipes (Olivier), **h**) Obrium maculatum (Olivier), **i**) Obrium rufulum Gahan, **j**) Obrium rubidum LeConte, **k**) Ancylocera bicolor (Olivier), **l**) Aethecerinus hornii (Lacordaire).

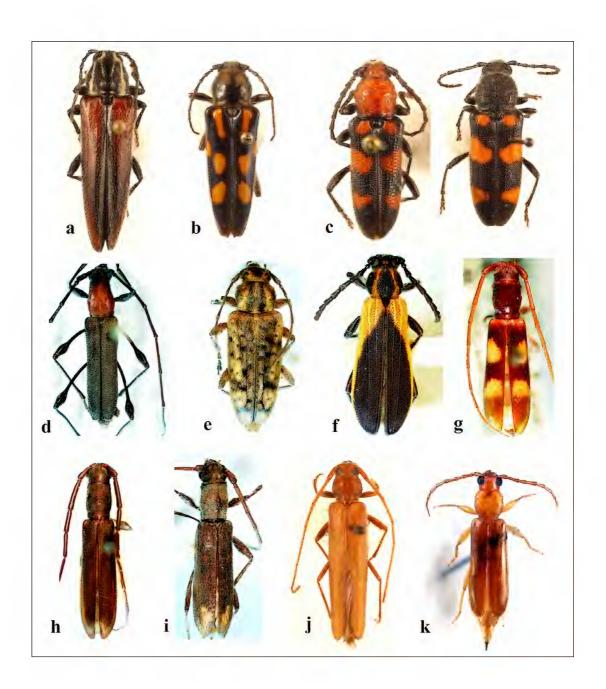


Plate 18.

a) Osmopleura chamaeropis (Horn), b) Agallissus lepturoides (Chevrolat), c) Zagymnus clerinus (LeConte), d) Rhopalophora longipes (Say), e) Atimia confusa (Say), f) Elytroleptus floridanus (LeConte), g) Heterachthes quadrimaculatus Haldeman, h) Heterachthes ebenus Newman, i) Heterachthes sablensis Blatchley, j) Oeme rigida (Say), k) Smodicum cucujiforme (Say).

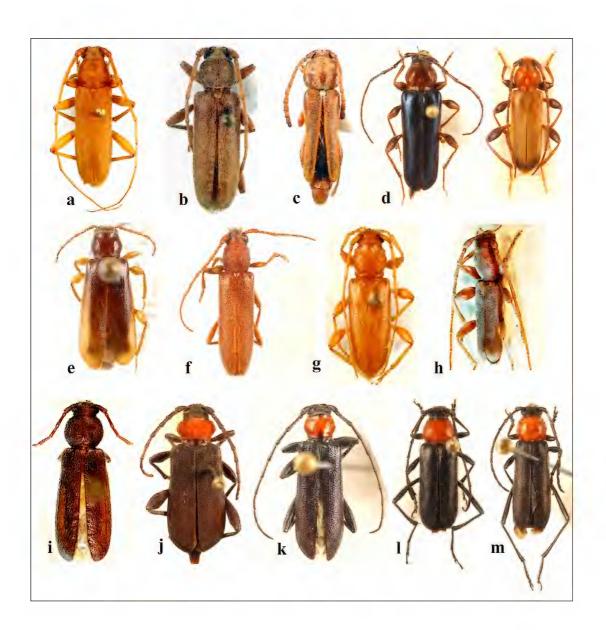


Plate 19.

a) Stromatium fulvum (Villers), b) Hesperophanes pubescens (Haldeman), c) Plesioclytus relictus Giesbert, d) Phymatodes testaceus (Linnaeus), e) Phymatodes aereus (Newman), f) Micranoplium unicolor (Haldeman), g) Curtomerus flavus (Fabricius), h) Gracilia minuta (Fabricius), i) Tylonotus masoni (Knull), j) Ropalopus sanguinicollis (Horn), k) Pronocera collaris (Kirby), l) Batyle ignicollis ignicollis (Say), m) Batyle ignicollis australis Linsley.

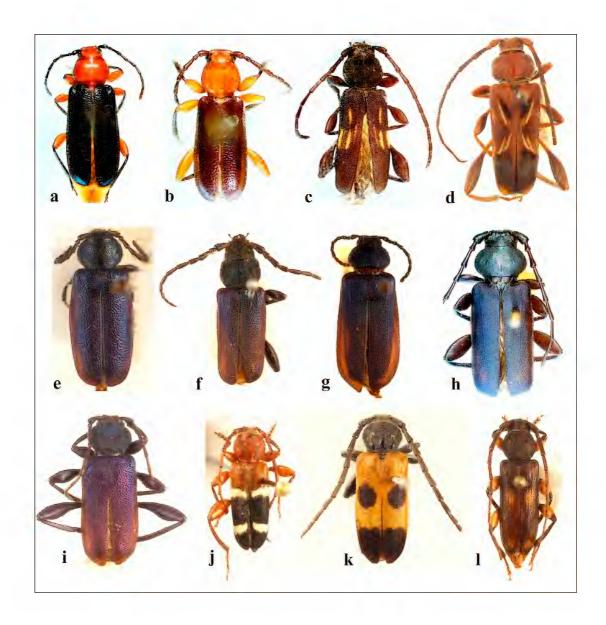


Plate 20.

a) Phymatodes amoenus (Say),
b) Phymatodes lengi Joutel,
c) Physocnemum brevilineum (Say),
d) Physocnemum andreae (Haldeman),
e) Callidium frigidum Casey,
f) Callidium texanum Schaeffer,
g) Callidium schotti Schaeffer,
h) Callidium antennatum Newman,
i) Callidium violaceum (Linnaeus),
j) Phymatodes varius (Fabricius),
k) Semanotus ligneus (Fabricius),
l) Tylonotus bimaculatus Haldeman.

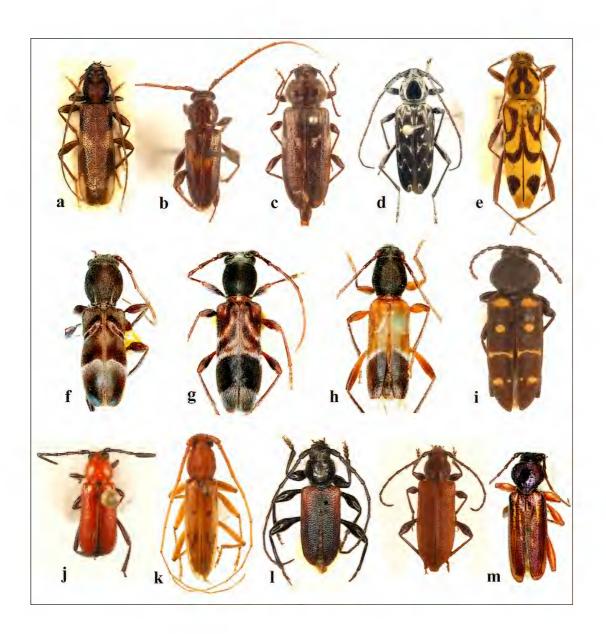


Plate 21.

a) Penichroa fasciata (Stephens), b) Curtomerus fasciatus (Fisher), c) Hylotrupes bajulus (Linnaeus), d) Sarosesthes fulminans (Fabricius), e) Chlorophorus annularis (Fabricius), f) Tilloclytus geminatus (Haldeman), g) Microclytus gazellula (Haldeman), h) Clytoleptus albofasciatus (Castelnau & Gory), i) Calloides nobilis (Harris), j) Batyle suturalis (Say), k) Achryson surinamum (Linnaeus), l) Callidiellum rufipenne (Motschulsky), m) Meriellum proteus (Kirby).

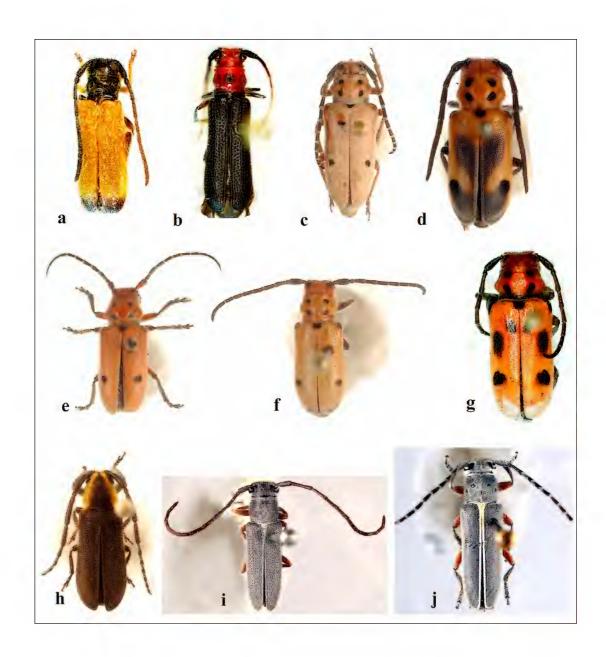


Plate 22.

a) Tetrops praeusta Linnaeus, b) Phaea monostigma (Haldeman), c) Tetraopes pilosus Chemsak, d) Tetraopes melanurus Schoenherr, e) Tetraopes texanus Horn, f) Tetraopes quinquemaculatus Haldeman, g) Tetraopes tetrophthalmus (Forster), h) Hemierana marginata (Fabricius), i) Mecas femoralis (Haldeman), j) Mecas pergrata (Say).

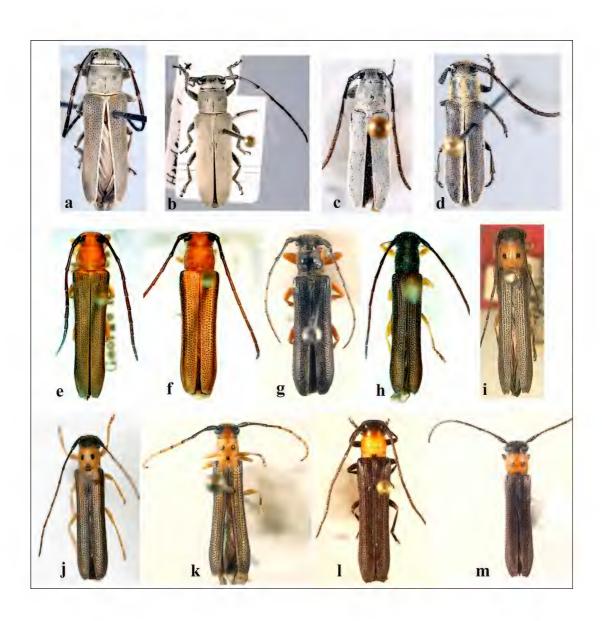


Plate 23.

a) Mecas cana cana (Newman),
b) Mecas cana saturnina LeConte,
c) Mecas cineracea Casey,
d) Mecas marginella LeConte,
e) Oberea ruficollis (Fabricius),
f) Oberea gracilis (Fabricius),
g) Oberea delongi Knull,
h) Oberea flavipes Haldeman,
i) Oberea ulmicola Chittenden,
j) Oberea tripunctata (Swederus),
k) Oberea praelonga Casey,
l) Oberea affinis Leng & Hamilton,
m) Oberea perspicillata Haldeman.

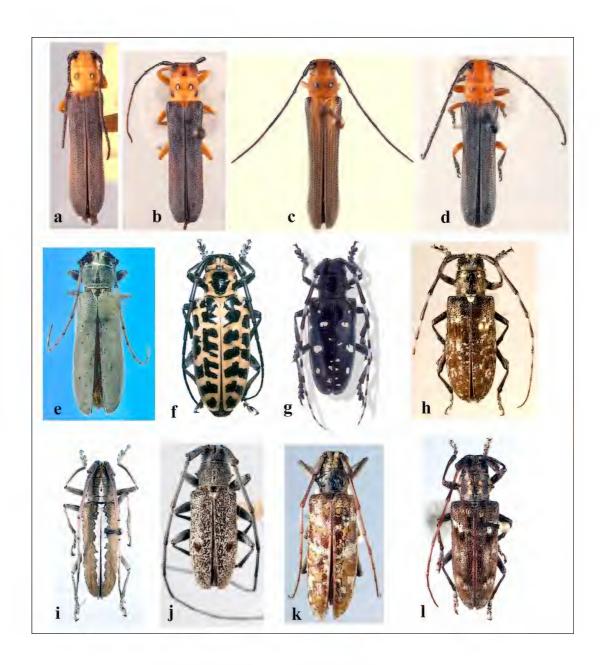


Plate 24.

a) Oberea caseyi Plavilstshikov, b) Oberea schaumii LeConte, c) Oberea myops Haldeman, d) Oberea ocellata Haldeman, e) Hebestola nebulosa Haldeman, f) Plectrodera scalator (Fabricius), g) Anoplophora glabripennis (Motschulsky), h) Monochamus scutellatus (Say), i) Neoptychodes trilineatus (Linnaeus), j) Microgoes oculatus (LeConte), k) Monochamus marmorator Kirby, l) Monochamus notatus (Drury).

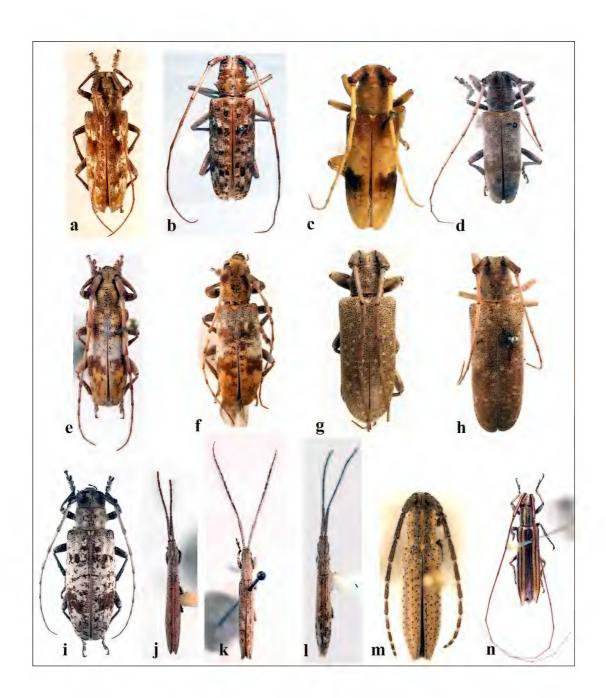


Plate 25.

- a) Monochamus carolinensis (Olivier), b) Monochamus titillator (Fabricius), c) Goes pulcher (Haldeman),
- d) Goes pulverulentus (Haldeman), e) Goes variegatus Linsley & Chemsak, f) Goes debilis LeConte,
- **g**) Goes tumifrons Chemsak & Linsley, **h**) Goes tesselatus (Haldeman), **i**) Goes tigrinus (DeGeer), **j**) Spalacopsis chemsaki Tyson, **k**) Spalacopsis filum costulatum Casey, **l**) Spalacopsis stolata Newman,
- m) Dorcasta cinerea (Horn), n) Hippopsis lemniscata (Fabricius).

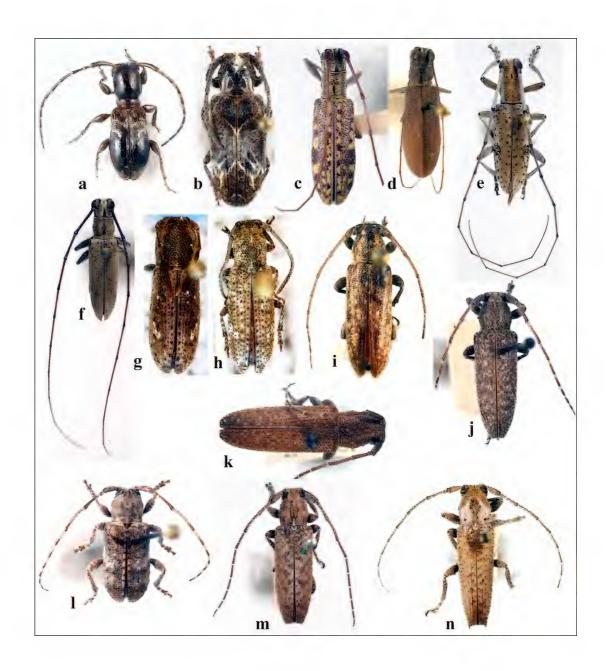


Plate 26.

a) Cyrtinus pygmaeus (Haldeman), b) Desmiphora hirticollis (Olivier), c) Dorcaschema alternatum (Say), d) Dorcaschema nigrum (Say), e) Dorcaschema wildii Uhler, f) Dorcaschema cinereum (Olivier), g) Parmenonta thomasi Chemsak & Linsley, h) Adetus brousi (Horn), i) Sybra alternans Wiedemann, j) Ataxia hubbardi Fisher, k) Ataxia falli Breuning, l) Ecyrus dasycerus (Say), m) Ataxia crypta (Say),

n) Ataxia spinicauda Schaeffer.

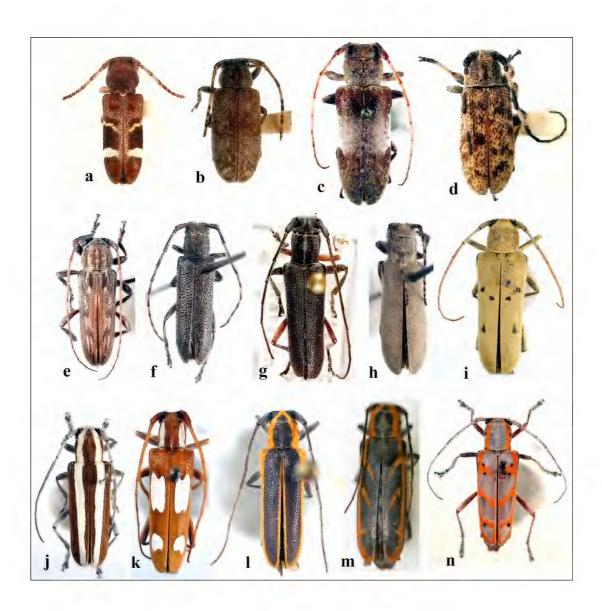


Plate 27.

a) Psenocerus supernotatus (Say), b) Zaplous annulatus (Chevrolat), c) Oncideres cingulata (Say), d) Eupogonius annulicornis Fisher, e) Lypsimena fuscata Haldeman, f) Saperda populnea moesta LeConte, g) Saperda discoidea Fabricius, h) Saperda inornata Say, i) Saperda vestita Say, j) Saperda candida Fabricius, k) Saperda cretata Newman, l) Saperda lateralis Fabricius, m) Saperda imitans Felt & Joutel, n) Saperda tridentata Olivier.

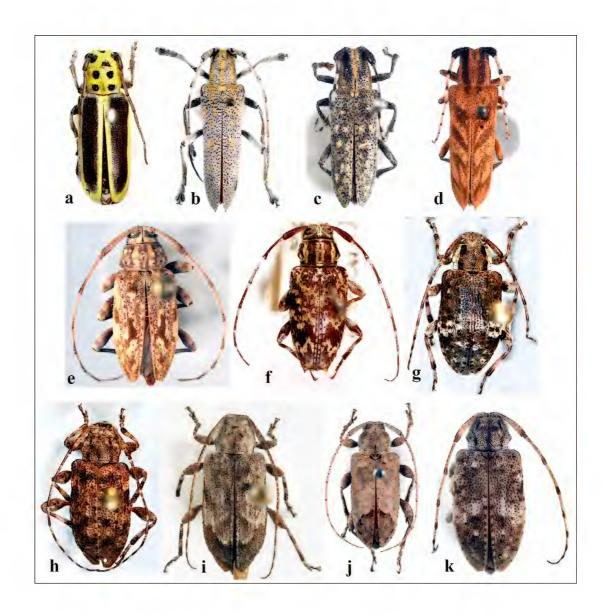


Plate 28.

a) Saperda puncticollis Say, b) Saperda calcarata Say, c) Saperda mutica Say, d) Saperda obliqua Say, e) Nyssodrysina haldemani (LeConte), f) Alcidion umbraticus (Jacquelin du Val), g) Astylopsis macula (Say), h) Astylopsis sexguttata (Say), i) Astylopsis arcuatus (LeConte), j) Astylopsis perplexa (Haldeman), k) Astylopsis collaris (Haldeman).

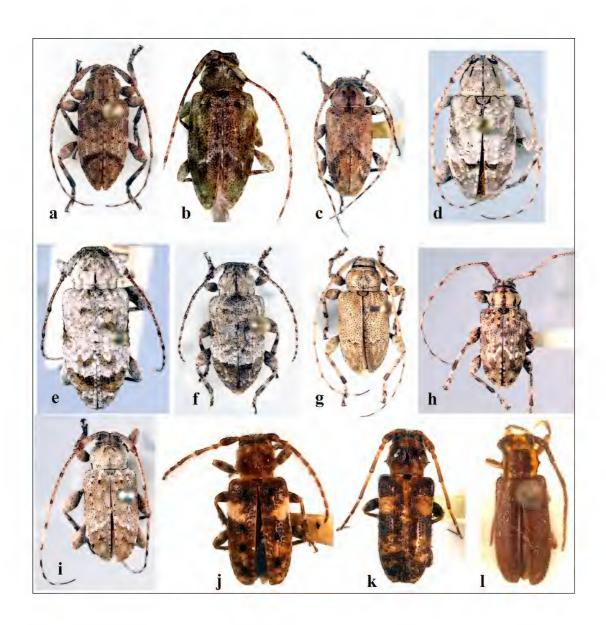


Plate 29.

a) Astylopsis fascipennis Schiefer, b) Astylidius parvus (LeConte), c) Styloleptus biustus (LeConte), d) Leptostylopsis argentatus (Jacquelin du Val), e) Leptostylus asperatus (Haldeman), f) Leptostylus transversus (Gyllenhal), g) Leptostylopsis terraecolor (Horn), h) Leptostylopsis albofasciatus (Fisher), i) Leptostylopsis planidorsus (LeConte), j) Pogonocherus penicillatus LeConte, k) Pogonocherus mixtus Haldeman, l) Eupogonius subarmatus (LeConte).

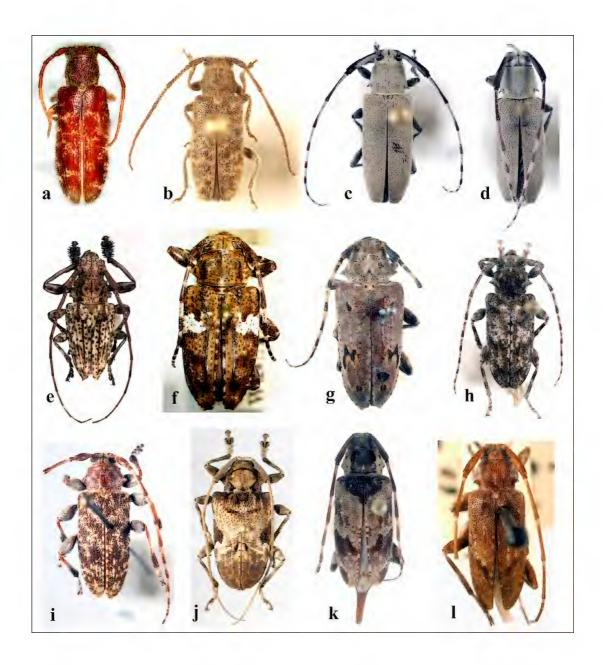


Plate 30.

a) Eupogonius tomentosus (Haldeman), b) Eupogonius pauper LeConte, c) Dectes texanus LeConte, d) Dectes sayi Dillon & Dillon, e) Steirastoma breve (Sulzer), f) Aegomorphus quadrigibbus (Say), g) Aegomorphus morrisii (Uhler), h) Aegomorphus modestus (Gyllenhal), i) Oplosia nubila (LeConte), j) Lagocheirus araneiformis stroheckeri Dillon, k) Urographis triangulifer (Haldeman), l) Eutrichillus biguttatus (LeConte).

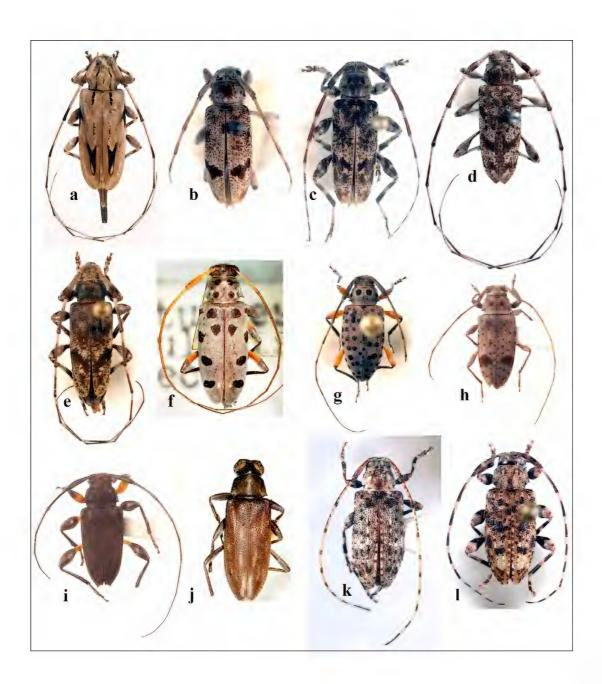


Plate 31.

a) Acanthocinus nodosus (Fabricius), **b)** Urographis despectus (LeConte), **c)** Urographis fasciatus (DeGeer), **d)** Acanthocinus obsoletus (Olivier), **e)** Acanthocinus pusillus Kirby, **f)** Lepturges regularis (LeConte), **g)** Hyperplatys aspersa (Say), **h)** Hyperplatys maculata Haldeman, **i)** Hyperplatys femoralis Haldeman, **j)** Lepturges megalops Hamilton, **k)** Liopinus punctatus (Haldeman), **l)** Sternidius variegatus (Haldeman).

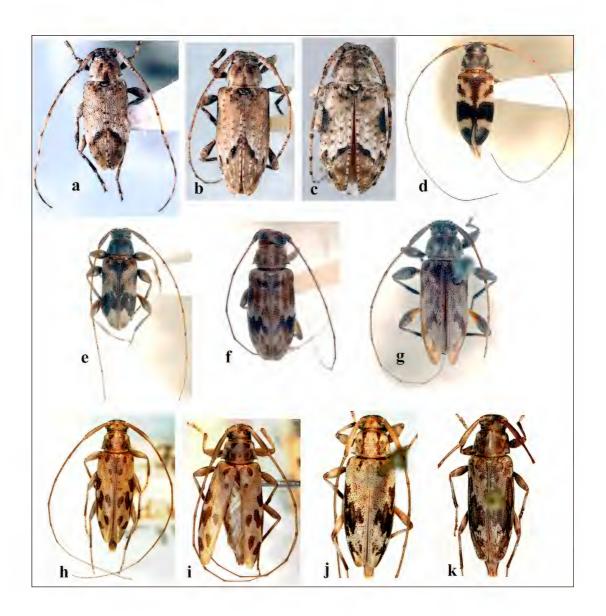


Plate 32.

a) Liopinus misellus (LeConte), b) Liopinus alpha (Say), c) Liopinus mimeticus (Casey), d) Urgleptes facetus (Say), e) Urgleptes querci (Fitch), f) Urgleptes foveatocollis (Hamilton), g) Urgleptes signatus (LeConte), h) Lepturges symmetricus (Haldeman), i) Lepturges pictus (LeConte), j) Lepturges angulatus (LeConte), k) Lepturges confluens (Haldeman).

Appendix 1.

Scientific and Common Names of Host Plants

(authorship and common names from the USDA NRCS PLANTS database [USDA NRCS 2007])

Abies P. Mill. (fir)

Abies balsamea (L.) P. Mill. (balsam fir)

Abies fraseri (Pursh) Poir. (Fraser fir)

Acacia farnesiana (L.) Willd. (sweet acacia;

huisache)

Acer L. (maple)

Acer negundo L. (boxelder)

Acer pseudoplatanus L. (sycamore maple)

Acer saccharum Marsh. (sugar maple)

Acer spicatum Lam. (mountain maple)

Achillea L. (yarrow)

Adansonia digitata L. (baobab)

Aesculus L. (buckeye)

Aesculus glabra Willd. (Ohio buckeye)

Aesculus pavia L. (red buckeye) Agropyron Gaertn. (wheatgrass)

Albizia Durazz. (albizia)

Alnus P. Mill. (alder)
Amelanchier arborea (Michx. f.) Fern. (common

serviceberry)

Ambrosia L. (ragweed)

Ambrosia L. (lagweed)
Ambrosia artemisifolia L. (annual ragweed)

Amorpha fruticosa L. (desert false indigo)

Ampelopsis arborea (L.) Koehne (peppervine)

Andropogon L. (bluestem)
Apocynum L. (dogbane)

Aruncus L. (aruncus)

Aruncus dioicus (Walt.) Fern. (bride's feathers)

Asclepias amplexicaulis Sm. (clasping milkweed)

Asclepias arenaria Torr. (sand milkweed) Asclepias hirtella (Pennell) Woods. (green

milkweed)

Asclepias syriaca L. (common milkweed)

Asclepias tuberosa L. (butterfly milkweed)

Asparagus L. (asparagus) Arctium L. (burdock) Aster L. (aster)

Avicennia germinans (L.) L. (black mangrove)

Baccharis halimifolia L. (eastern baccharis)

Bambusa Schreb. (bamboo)
Batis maritima L. (turtleweed)

Betula L. (birch)

Betula nigra L. (river birch)

Bombax ceiba L. (red silk cottontree)

Bursera simaruba (L.) Sarg. (gumbo limbo)

Carpinus caroliniana Walt. (American hornbeam)

Carya Nutt. (hickory)

Carya aquatica (Michx. f.) Nutt. (water hickory)

Carya cordiformis (Wangenh.) K. Koch (bitternut hickory)

Carya floridana Sarg. (scrub hickory)

Carya glabra (P. Mill.) Sweet (pignut hickory)

Carya illinoinensis (Wangenh.) K. Koch (pecan)

Carya laciniosa (Michx. f.) G. Don (shellbark hickory)

Carya ovata (P. Mill.) K. Koch (shagbark hickory)

Castanea P. Mill. (chestnut)

Castanea dentata (American chestnut)
Casuarina equisetifolia L. (beach sheoak)

Ceanothus L. (New Jersey Tea)

Ceanothus americanus (New Jersey tea)

Cecropia Loefl. (pumpwood)

Celtis L. (hackberry)

Celtis laevigata Willd. (sugarberry)

Celtis laevigata Willd. var. reticulata (Torr.) L.

Benson (netleaf hackberry)

Celtis occidentalis L. (common hackberry)

Celtis tenuifolia Nutt. (dwarf hackberry)

Cercis canadensis L. (eastern redbud)

Chamaecyparis Spach (cedar)

Chenopodium botrys (Jerusalem oak)

Cirsium P. Mill. (thistle)

Citrus L. (citrus)

Coccaloba diversifolia Jacq. (tietongue)

Cocos nucifera L. (coconut palm)

Cojoba arborea (L.) Britt. & Rose (wild tamarind)

Conocarpus erectus L. (button mangrove)

Cordia L. (cordia)

Coreopsis L. (tickseed)

Cornus L. (dogwood)

Cornus asperifolia Michx. (toughleaf dogwood)

Cornus florida L. (flowering dogwood) Corylopsis Sieb. & Zucc. (winter hazel)

Crataegus L. (hawthorn)

Crataegus viridis L. (green hawthorn)

Crossopetalum rhacoma Crantz (maidenberry)

Croton capitatus Michx. (hogwort)

Cryptomeria japonica (L. f.) D. Don (Japanese

cedar)

Cupressus L. (cypress)

Cucurbita L. (gourd)

Cydonia oblonga P. Mill (quince)
Daucus carota L. (Queen Anne's lace)

Delonix regia (Bojer ex Hook.) Raf. (royal

poinciana)

Diospyros virginiana L. (common persimmon)

Appendix 1. Scientific and Common Names of Host Plants

Ebenopsis ebano (Berl.) Barneby & Grimes Lithospermum caroliniense (Walt. ex J. F. Gmel.) (Texas ebony) MacM. (Carolina puccoon) Erigeron L. (fleabane) Maclura pomifera (Raf.) Schneid. (osage orange) Eupatorium L. (thoroughwort) Magnolia L. (magnolia) Fagus L. (beech) Malus P. Mill. (apple) Fagus grandifolia Ehrh. (American beech) Malus angustifolia (Ait.) Michx. (southern Ficus L. (fig) crabapple) Ficus aurea Nutt. (Florida strangler fig) Mangifera indica L. (mango) Ficus citrifolia P. Mill. (wild banyantree) Marshallia Schreb. (Barbara's buttons) Flaveria linearis Lag. (narrowleaf yellowtops) *Matelea* Aubl. (milkvine) Forestiera segregata (Jacq.) Krug & Urban Medicago L. (alfalfa) (Florida swampprivet) Melilotus P. Mill. (sweetclover) Fraxinus L. (ash) Melothria L. (melothria) Fraxinus americana L. (white ash) Menispermum L. (moonseed) Fraxinus quadrangulata Michx. (blue ash) Metopium toxiferum (L.) Krug & Urban (Florida Gaillardia Foug. (blanketflower) poisontree) Geranium maculatum L. (spotted geranium) Morus L. (mulberry) Gleditsia triacanthos L. (honeylocust) Musa L. (banana) Gossypium L. (cotton) *Nicotiana* L. (tobacco) Gossypium thurberi Todaro (Thurber's cotton) Nyssa L. (tupelo) Grindelia Willd. (gumweed) *Nyssa aquatica* L. (water tupelo) Nyssa sylvatica Marsh. (blackgum) Guajacum officinale L. (lignum–vitae) Gymnocladus dioicus (L.) K. Koch (Kentucky *Oenothera* L. (evening–primrose) Opuntia P. Mill (pricklypear) coffeetree) Helenium L. (sneezeweed) Ostrya virginiana (P. Mill.) K. Koch Helianthus L. (sunflower) (hophornbeam) Oxydendrum arboreum (L.) DC. (sourwood) Helianthus annuus L. (common sunflower) Heliopsis helianthoides (L.) Sweet var. occi-Oxypolis Raf. (cowbane) dentalis (T. R. Fisher) Steyermark (smooth Parkinsonia aculeata L. (Jerusalem thorn) Parthenocissus quinquefolia (L.) Planch. oxeye) Heracleum L. (cowparsnip) (Virginia creeper) Hevea Aublet (hevea) Paspalum notatum Flueggé (bahiagrass) *Hibiscus* L. (rosemallow) Persea P. Mill. (bay) Hydrangea arborescens L. (wild hydrangea) Persea borbonia L. Spreng. (redbay) Hymenaea courbaril L. (stinkingtoe) Phlox L. (phlox) Phytolacca L. (pokeweed) *Ilex* L. (holly) *Ilex glabra* (L.) Gray (inkberry) Picea A. Dietr. (spruce) *Ipomoea* L. (morning–glory) Picea rubens Sarg. (red spruce) Jatropha L. (nettlespurge) Pimenta dioica (L.) Merr. (allspice) Juglans L. (walnut) Pinus L. (pine) Juglans cinerea L. (butternut) *Pinus banksiana* Lamb. (jack pine) Juglans nigra L. (black walnut) Pinus caribaea Morelet (Caribbean pine) Juniperus L. (juniper) *Pinus echinata* P. Mill. (shortleaf pine) Juniperus ashei Buchh. (Ashe's juniper) Pinus rigida P. Mill. (pitch pine) Lantana L. (lantana) Pinus sylvestris L. (Scots pine) Laguncularia racemosa (L.) Gaertn. f. (white Pinus taeda L. (loblolly pine) Pinus virginiana P. Mill. (Virginia pine) mangrove) Larix P. Mill. (larch) Pinus strobus L. (eastern white pine) Lindera Thunb. (spicebush) Piscidia L. (piscidia) Liquidambar styraciflua L. (sweetgum) Piscidia piscipula (L.) Sarg. (Florida fishpoison Liriodendron L. (tuliptree) tree; Jamaica dogwood) Lysiloma Benth. (false tamarind) Pithecellobium dulce (Roxb.) Benth. (monkey-Lysiloma latisiliquum (L.) Benth. (false tamarind) pod)

Appendix 1. Scientific and Common Names of Host Plants

Platanus occidentalis L. (American sycamore) Platanus wrightii S. Wats. (Arizona sycamore)

Populus L. (cottonwood)

Populus deltoides Bartr. ex Marsh (eastern cottonwood)

Populus tremuloides Michx. (quaking aspen)

Prosopis juliflora (Sw.) DC. (mesquite)

Prunus L. (plum)

Prunus persica (L.) Batsch (peach)

Prunus serotina Ehrh. (black cherry)

Pseudotsuga menziesii (Mirbel) Franco

(Douglas-fir)

Punica granatum L. (pomegranate)

Pyrus L. (pear) Quercus L. (oak)

Quercus alba L. (white oak)

Quercus bicolor Willd. (swamp white oak)

Quercus falcata Michx. (southern red oak)

Quercus geminata Small (sand live oak)

Quercus inopina Ashe (sandhill oak)

Quercus laevis Walt. (turkey oak)

Quercus laurifolia Michx. (laurel oak)

Quercus lyrata Walt. (overcup oak)

Quercus macrocarpa Michx. (bur oak)

Quercus muhlenbergii Engelm. (chinkapin oak)

Quercus nigra L. (water oak)

Quercus palustris Muenchh. (pin oak)

Quercus phellos L. (willow oak)

Quercus stellata Wangenh. (post oak)

Quercus vaseyana Buckl. (sandpaper oak)

Quercus velutina Lam. (black oak)

Quercus virginiana P. Mill. (live oak)

Rhizophora mangle L. (red mangrove)

Rhododendron L. (rhododendron)

Rhus L. (sumac)

Rhus glabra L. (smooth sumac)

Robinia L. (locust)

Robinia pseudoacacia L. (black locust)

Rosa L. (rose)

Rubus L. (raspberry; blackberry)

Rudbeckia L. (coneflower)

Sabal palmetto (Walt.) Lodd. ex J. A. & J. H.

Schultes (cabbage palmetto)

Salix L. (willow)

Salix exigua Nutt. (narrowleaf willow)

Salix nigra Marsh. (black willow)

Sambucus nigra L. ssp. canadensis (L.) R. Bolli

(common elderberry)

Sapindus saponaria L. (soapberry)

Sassafras Nees & Eberm. (sassafras)

Scalesia Hooker (giant daisy tree)

Schinopsis balansae Engler (quebracho)

Sideroxylon foetidissimum Jacq. (false mastic)

Sideroxylon lanuginosum Michx. ssp. lanuginosum (gum bully)

Sideroxylon tenax (L.) (tough bully)

Sinobambusa gibbosa McClure (gibbous

bamboo)

Smilacina racemosa (false Solomon's seal)

Smilax L. (greenbrier)

Solanum L. (nightshade)

Solidago L. (goldenrod)

Sorghastrum Nash (Indiangrass)

Spiraea L. (spirea)

Spondias L. (mombin)

Spondias purpurea L. (purple mombin)

Sporobolus R. Br. (dropseed)

Swietenia Jacq. (mahogany)

Taxodium distichum (L.) L. C. Rich. (bald cy-

press)

Tectona L. f. (tectona)

Thuja L. (red cedar)

Thuja occidentalis L. (arborvitae)

Tilia L. (basswood; linden)

Toxicodendron radicans (L.) Kuntze (eastern

poison ivy)

Tragopogon L. (goatsbeard)

Tsuga Carr. (hemlock)

Ulmus L. (elm)

Ulmus alata Michx. (winged elm)

Ulmus crassifolia Nutt. (cedar elm)

Ulmus rubra Muhl. (slippery elm)

Vaccinium L. (blueberry)

Vaccinium arboreum Marsh. (farkleberry)

Vernonia Schreb. (ironweed)

Viburnum L. (viburnum)

Vitis L. (grape)

Wisteria Nutt. (wisteria)

Xanthium L. (cocklebur)

Ximenia americana L. (tallow wood)

Zanthoxylum fagara (L.) Sarg. (lime pricklyash)

Zanthoxylum flavum Vahl. (West Indian satinwood)

Zea mays L. (corn)

Acanthocinus nodosus (Fabricius)	couplet 393, pp. 128, 151
Acanthocinus obsoletus (Olivier)	couplet 396, pp. 129, 151
Acanthocinus pusillus Kirby	couplet 396, pp. 129, 151
Achryson surinamum (Linnaeus)	
Acmaeops discoideus (Haldeman)	
Acmaeops proteus (Kirby)	couplets 57, 76, pp. 25, 30, 139
Adetus brousi (Horn)	couplet 329, pp. 108, 151
Aegomorphus modestus (Gyllenhal)	couplet 388, pp. 126, 151
Aegomorphus morrisii (Uhler)	couplet 387, pp. 126, 151
Aegomorphus quadrigibbus (Say)	couplet 386, pp. 125, 151
Aethecerinus hornii (Lacordaire)	couplets 211, 249, pp. 75, 86, 143
Agallissus lepturoides (Chevrolat)	couplet 214, pp. 76, 143
Alcidion umbraticus (Jacquelin du Val)	couplets 354, 398, pp. 115, 129, 151
Alosternida chalybaea (Haldeman)	couplets 72, 88, pp. 29, 33, 139
Analeptura lineola (Say)	couplet 80, pp. 31, 139
Ancylocera bicolor (Olivier)	
Aneflomorpha delongi (Champlain & Knull)	
Aneflomorpha subpubescens (LeConte)	
Anelaphus cinereus (Olivier).	
Anelaphus inermis (Newman)	
Anelaphus moestus (LeConte)	
Anelaphus mutatum (Gahan)	
Anelaphus parallelus (Newman)	
Anelaphus pumilus (Newman)	
Anelaphus villosus (Fabricius)	
Anoplodera pubera (Say)	
Anoplophora glabripennis (Motschulsky)	
Anthophylax attenuatus (Haldeman)	
Anthophylax cyaneus (Haldeman)	
Anthophylax hoffmani Beutenmüller	
Anthophylax viridis LeConte	
Archodontes melanopus (Linnaeus)	
Arhopalus foveicollis (Haldeman)	
Arhopalus rusticus (LeConte)	
Asemum australe LeConte	
Asemum striatum (Linnaeus)	
Astylidius parvus (LeConte)	
Astylopsis collaris (Haldeman)	
Astylopsis fascipennis Schiefer	
Astylopsis macula (Say)	
Astylopsis perplexa (Haldeman)	
Astylopsis sexguttata (Say)	
Ataxia crypta (Say)	
Ataxia falli Breuning	
Ataxia hubbardi Fisher	
Ataxia spinicauda Schaeffer	
Atimia confusa (Say)	
Batyle ignicollis australis Linsley	
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Batyle suturalis (Say)	couplet 261, pp. 89, 144

Bellamira scalaris (Say)	couplet 83, pp. 32, 139
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Brachyleptura vagans (Olivier)	
Brachysomida bivittata (Say)	couplets 39, 47 pp. 19, 22, 140
Callidiellum rufipenne (Motschulsky)	couplet 263, pp. 90, 144
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Callidium frigidum Casey	couplet 244, pp. 85, 144
Callidium schotti Schaeffer	couplet 245, pp. 85, 144
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Centrodera quadrimaculata (Champlain & Knull)	
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Chlorida festiva (Linnaeus)	
Chlorophorus annularis (Fabricius)	
Clytoleptus albofasciatus (Castelnau & Gory)	
Clytus marginicollis Castelnau & Gory	
Clytus ruricola (Olivier)	
Curius dentatus Newman	
Curtomerus fasciatus (Fisher)	
Curtomerus flavus (Fabricius)	
Cyrtinus pygmaeus (Haldeman)	
Cyrtophorus verrucosus (Olivier)	
Dectes sayi Dillon & Dillon	
Dectes texanus LeConte	
Derobrachus brevicollis Audinet-Serville	
Desmiphora hirticollis (Olivier)	
Desmocerus palliatus (Forster)	
Distenia undata (Fabricius)	
Dorcaschema alternatum (Say)	
Dorcaschema cinereum (Olivier)	
Dorcaschema nigrum (Say)	
Dorcaschema wildii Uhler	
Dorcasta cinerea (Horn)	
Dryobius sexnotatus Linsley	
Eburia cinereopilosa Fisher	
Eburia distincta Haldeman	
Eburia haldemani LeConte	
Eburia quadrigeminata (Say)	
Eburia stigma (Olivier)	
Eburia stroheckeri Knull	
Ecyrus dasycerus (Say)	
Elaphidion clavis Linsley (see E. tectum LeConte)	
Elaphidion cryptum Linsley	
Elaphidion irroratum (Linnaeus)	
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Enaphalodes archboldi Lingafelter & Chemsak	
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Gracilia minuta (Fabricius)	
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Grammoptera subargentata (Kirby)	
Hebestola nebulosa Haldeman	
Hemierana marginata (Fabricius)	
Hesperandra polita (Say)	
Hesperophanes pubescens (Haldeman)	
Heterachthes ebenus Newman	
Heterachthes quadrimaculatus Haldeman	
Heterachthes sablensis Blatchley	
Heterops dimidiatus (Chevrolat)	
Hippopsis lemniscata (Fabricius)	
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Leptostylus transversus (Gyllenhal)	
Leptura abdominalis (Haldman)	
Leptura obliterata deleta (LeConte)	
Leptura subhamata Randall	
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Liopinus punctatus (Haldeman)	
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Lypsimena fuscata Haldeman	
Mallodon dasystomus (Say)	
Mecas cana cana (Newman)	
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Mecas cineracea Casey	
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Megacyllene caryae (Gahan)	
Megacyllene decora (Olivier)	
Megacyllene robiniae (Forster)	
Meriellum proteus (Kirby)	
Metacmaeops vittata (Swederus)	
Methia necydalea (Fabricius)	
Michthisoma heterodoxum LeConte	
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Microclytus gazellula (Haldeman)	
Microgoes oculatus (LeConte)	
Molorchus bimaculatus bimaculatus Say	
Molorchus bimaculatus corni Haldeman	
Molorchus bimaculatus semiustus (Newman)	
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Monochamus marmorator Kirby	
Monochamus notatus (Drury)	
Monochamus scutellatus (Say)	
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Sphenostethus taslei (Buquet)	
Steirastoma breve (Sulzer)	
Stenelytrana emarginata (Fabricius)	
Stenocorus cinnamopterus (Randall)	
Stenocorus cylindricollis (Say)	
Stenocorus schaumii (LeConte)	
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